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Drug Repurposing in Medulloblastoma: Challenges and Recommendations

Hussein Hammoud, MSc¹ Zahraa Saker, MSc¹ Hayat Harati, $PhD¹$ Youssef Fares, MD, PhD^{1,2} Hisham F. Bahmad, MD, $MSc^{3,4,*}$ Sanaa Nabha, Ph $D^{1,*}$

Address

*^{,1}Neuroscience Research Center, Faculty of Medical Sciences, Lebanese University, Hadas, Beirut, Lebanon Email: snabha@ul.edu.lb ²Department of Neurosurgery, Faculty of Medical Sciences, Lebanese University, Beirut, Lebanon ³Arkadi M. Rywlin M.D. Department of Pathology and Laboratory Medicine, Mount

Sinai Medical Center, 4300 Alton Rd, Miami Beach, FL, 33140, USA

*^{,4}Herbert Wertheim College of Medicine, Florida International University, Miami, FL, USA Email: hfbahmad@gmail.com

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Hussein Hammoud and Zahraa Saker are authors who contributed equally to this work as co-first authors. Hisham F. Bahmad and Sanaa Nabha are authors who contributed equally to this work as co-senior authors. This article is part of the Topical Collection on Neuro-oncology

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Opinion statement

Medulloblastoma is the most frequently diagnosed primary malignant brain tumor among children. Currently available therapeutic strategies are based on surgical resection, chemotherapy, and/or radiotherapy. However, majority of patients quickly develop therapeutic resistance and are often left with long-term therapy-related side effects and sequelae. Therefore, there remains a dire need to develop more effective therapeutics to overcome the acquired resistance to currently available therapies. Unfortunately, the process of developing novel anti-neoplastic drugs from bench to bedside is highly time-consuming and very expensive. A wide range of drugs that are already in clinical use for treating non-

cancerous diseases might commonly target tumor-associated signaling pathways as well and hence be of interest in treating different cancers. This is referred to as drug repurposing or repositioning. In medulloblastoma, drug repurposing has recently gained a remarkable interest as an alternative therapy to overcome therapy resistance, wherein existing non-tumor drugs are being tested for their potential anti-neoplastic effects outside the scope of their original use.

Introduction

Medulloblastoma (MB) is the most frequently diagnosed primary malignant brain tumor diagnosed in children [[1](#page-22-0)], accounting for around a quarter of all pediatric central nervous system (CNS) neoplasms [\[2\]](#page-22-0). It is a highly aggressive tumor that usually develops early during embryonic life [[3](#page-22-0)•]. It mainly originates from the cerebellar vermis and thus primarily affects children in their first year of life [\[4\]](#page-23-0). Studies have shown that MB arises from remnants of the primitive neuroectoderm within the germinal matrix of the fourth ventricle roof [\[5,6](#page-23-0)] or from the external granular layer precursor cells [[7](#page-23-0),[8](#page-23-0)].

It is approximated that MB affects around 3.9 per million children aged less than 19 years old [[9\]](#page-23-0), and it affects males more than females [\[9\]](#page-23-0). Although MB is aggressive and highly invasive, the 10-year survival rate is around 60–70% [\[9\]](#page-23-0) and the 5-year survival rate ranges between 70 and 85% [[10,11\]](#page-23-0) due to the multimodality treatment approach and improved treatments methods [\[5\]](#page-23-0). Nevertheless, there still exists an imbalance in survival rates where low- to middle-income countries record 33% $[12]$ $[12]$ to 45.6% $[13]$ $[13]$ 5-year progression-free survival rates.

Classification of medulloblastoma subtypes

Histological classification of medulloblastoma

According to the World Health Organization (WHO), MB can be either classified based on a histological classification or according to molecular and genetic features. Histologically, MB can be divided into four distinct subsets [[14\]](#page-23-0): classic MB (CMB), desmoplastic/nodular MB (DMB), MB with extensive nodularity (MBEN), and large cell/anaplastic with subgroups of melanotic MB and medullomyoblastoma which are extremely rare [[15\]](#page-23-0). CMB represents the most common histological subtype of MB and is characterized by sheets of densely packed basophilic small round cells with high nuclear-to-cytoplasmic ratio and showing a mitotic and apoptotic activity [\[14](#page-23-0)]. Similar to CMB, MBEN shows small round tumor cells but is associated with reticulin-free islands within a reticulin-rich stroma [[16](#page-23-0),[17\]](#page-23-0). This subtype is also accompanied by a favorable prognosis [[16\]](#page-23-0). As for DMB, what distinguishes this subtype from CMB is the presence of desmoplasia and marked tendency for neuronal differentiation [[18\]](#page-23-0). Considering the large cell/anaplastic MB, it is a highly aggressive tumor subtype that is usually located in the cerebellar vermis. Tumor cells show extensive and marked nuclear pleomorphism with prominent nucleoli [[18\]](#page-23-0).

Molecular classification of medulloblastoma

At the molecular level, according to the latest consensus nomenclature, 4 major subgroups of MB were identified [[19\]](#page-23-0): MB^{WNT-activated}, MB^{SHH-activated,TP53-mutant}, $MB^{SHH\text{-}activated, TP53-wildtype}$, and $MB^{non-WNT/non-SHH}$. The latter comprises two sub-groups, MB^{non-WNT/non-SHH, Group-3} [\[20](#page-23-0),[21](#page-23-0)] and MB^{non-WNT/non-SHH, Group-4} [\[22,23](#page-23-0)], which are characterized by high rates of disseminated disease and less understood pathology [\[24](#page-24-0)•].

WNT-activated medulloblastoma

The WNT subgroup is the mostly known subgroup, with the best long-term prognosis and survival rates that exceed 90% compared to other molecular subgroups [\[25](#page-24-0)]. MB^{WNT-activated} harbors activating mutations in the Wnt pathway effector β-catenin 1 (CTNNB1) [[26,27\]](#page-24-0) and is characterized by positive nuclear immunohistochemical (IHC) staining for β-catenin [[28](#page-24-0)], AXIN1 mutation and AXIN2 deletion [\[29](#page-24-0)], and loss of chromosome 6 (monosomy six) [[30](#page-24-0),[31\]](#page-24-0) which is considered a defining feature of this MB subgroup that is rarely found in non-WNT MB tumors [[32](#page-24-0)].

SHH-activated medulloblastoma

Regarding the SHH subgroup, it is most common in infants and adults. SHHactivated MB constitutes 30% of all MB cases and is further subdivided into TP53-mutant and TP53-wildtype [[33](#page-24-0)]. It has a good prognosis in infants and is usually caused by aberrant activation of the sonic hedgehog signaling pathway [[34\]](#page-24-0). Infantile MB can be due to germline mutations of the Shh negative regulator SUFU. Somatic mutations, as in PTCH, SMO, and SUFU, and even amplification of GLI1 and GLI2 were also observed in this MB subgroup [[35](#page-24-0),[36,32,37\]](#page-24-0). As for identification methods, immunohistochemical staining for SFRP1 or GAB1, in addition to the deletion of chromosome 9q, is considered to be limited to SHH-MB (the gene for PTCH is located on the chromosome 9q22) [\[32](#page-24-0)].

Non-WNT/non-SHH medulloblastoma

The non-WNT/non-SHH MB group is subdivided into groups 3 and 4, both of which have moderately high overall expression of MYCN [[38\]](#page-24-0). NPR3 has been postulated to be a potential molecular marker for MB^{non-WNT}/non-SHH, Group-3, while KCNA1, CDK6, and MYCN marker genes characterize MB^{non-WNT/non-SHH,} $Group-4$ [[39](#page-24-0)]. In general, unlike the other groups, group 3 is the most common type in infants and is associated with a very poor prognosis [\[32](#page-24-0)]. In non-WNT/ non-SHH MB groups 3 and 4, H3K27me3 is recurrently dysregulated. Normally, H3K27 is methylated by a methyltransferase EZH2 and demethylated by demethylase 6A (KDM6A) and KDM6B. KDM6A and other members of the KDM family were found to be mutated in both groups 3 and 4. Besides, EZH2 overexpression and gain of 7q (where EZH2 is located) were observed in these subgroups. Henceforth, disruption of genes in cooperation with histone methylation might represent major events leading to tumor development [\[40](#page-24-0)–[42](#page-24-0)].

Current treatment modalities for medulloblastoma:

Different treatment modalities are currently in use clinically for MB ranging from surgical intervention to chemotherapy and radiation therapy [[43,](#page-24-0)[44](#page-25-0)]. Those therapeutic options, however, are accompanied with substantial adverse effects including, for instance, 7–50% increased risk of postoperative cerebellar mutism syndrome (pCMS) following surgery [\[45](#page-25-0)–[47\]](#page-25-0). Also, although craniospinal radiotherapy prolongs patient survival, it decreases patients' intelligence quotient (IQ) by 2–4 points per year [\[48\]](#page-25-0) and is accompanied by other toxicities like impaired spinal growth and endocrine dysfunction [[49\]](#page-25-0). Despite that surgery remains the major therapeutic option to remove the malignant mass, radiotherapy is used to destroy tumor cells via proton therapy, and chemotherapy is also given in single-drug or combinatorial approaches [\[50](#page-25-0)]. Other treatments are also available that include molecular targeted therapies, such as silencing endogenous miRNA or targeting specific pathways that underlie MB initiation and growth [[51\]](#page-25-0).

Despite the available therapies, MB like many other CNS tumors [[52,](#page-25-0)[3\]](#page-22-0) recurs in many patients pertaining to development of therapeutic resistance, which necessitates looking for more effective treatments [\[53\]](#page-25-0). Recently, a promising approach has emerged which enables the usage of non-cancerous approved drugs that commonly target specific cancer-related pathways, for cancer [[54](#page-25-0)••]. This approach is known as drug repurposing or repositioning and serves as a potential novel option for treating different tumors and overcoming therapy resistance.

Repurposing approved drugs in cancer

Repurposing drugs in cancer via experimental and computational approaches

Despite the advancement in cancer research, developing anti-tumor therapies is still very challenging due to the different stages that any drug needs to pass through before approving it for clinical use and more importantly due to development of therapeutic resistance. Therefore, new approaches are required to tackle this issue. Drug repurposing is a promising approach recently emerging across different tumor types where existing drugs are being used beyond their original non-cancer indications to be utilized in cancer therapy. Repositioning of FDA-approved drugs for new clinical indications offers advantages of reduced drug development time and cost [\[55](#page-25-0)], taking into consideration that pharmacokinetic, pharmacodynamic, and toxicological data have been already assessed [[56\]](#page-25-0). Drug repurposing has been effective in different oncologic conditions. Anthracyclines, for instance, are class of antibiotics including doxorubicin, daunorubicin, epirubicin, and idarubicin that have been shown to instigate chemotherapeutic potentials in various tumor types such as breast cancer [[57](#page-25-0),[58\]](#page-25-0) and lymphoma [\[59,60](#page-25-0)].

Repurposing FDA-approved drugs usually requires combining both experimental (in vitro and in vivo screening of the potential drugs [\[61\]](#page-25-0)) and computational "in silico" approaches (computationally determine drug interactions with specific targets [\[62](#page-25-0)]). In the pediatric population, children with brain tumors have not shown a significant improvement in their survival [\[63](#page-25-0)], raising the need for identifying more effective drugs to improve patients' quality of life and prolong survival $[64,54\bullet,65]$ $[64,54\bullet,65]$ $[64,54\bullet,65]$ $[64,54\bullet,65]$ $[64,54\bullet,65]$. In this review, we summarize the drug repurposing approaches that have been described so far in MBs (Table [1](#page-5-0)).

Overcoming the challenges in crossing the blood-brain barrier

The blood-brain barrier (BBB) comprises highly specialized microvascular endothelial cells [\[66\]](#page-25-0) that are characterized by specific properties such as lack of fenestration, tight junctions, and presence of solute carriers and transporters [[67](#page-26-0)]. It is implicated in the maintenance of brain homeostasis and protecting the brain tissue from different substances, toxins, and pathogens [\[68](#page-26-0)]. Brain tumors recruit the vascular network of the brain for their continuous growth; however, the BBB hinders delivery of some drugs to the tumor site in the brain [[69](#page-26-0)], particularly some chemotherapeutic agents that are too large or hydrophilic to cross it [\[70,71](#page-26-0)]. Different factors compromise the drugs' ability to reach the brain, such as efflux pumps (P-glycoprotein) that actively transport drugs back into the blood [\[72\]](#page-26-0), physiochemical properties of drugs [[73\]](#page-26-0), and the glymphatic system [\[74,75](#page-26-0)].

MB is subcategorized into different subgroups possessing distinct genomic profiles [[76\]](#page-26-0). Recently, it was declared that the MB genotype dictates the phenotype of BBB, explaining the disparate prognoses pertaining to the differential chemotherapeutic responsiveness seen between the various MB subtypes [[77](#page-26-0)]. For example, while $MB^{WNT-activated}$ has an unfunctional BBB rendering it more vulnerable to chemotherapy, MB^{SHH-activated} has an intact BBB [[77\]](#page-26-0). This heterogeneity in the BBB phenotypes among the different MB subgroups raises a need for identifying novel therapeutic targets and trying new drugs to improve the management of MB. Using non-cancer drugs that have already gained FDA approval and trying them as potential anti-cancer therapies for MB might be a possible option to bypass the timely stages of drug testing needed. Those drugs include antihyperlipidemic agents, medications used for cardiovascular diseases, anthelmintic drugs, antimicrobials, antivirals and antiretroviral drugs, nonsteroidal anti-inflammatory drugs (NSAIDs), medicines used in neurological disorders, and others as outlined below.

Drug repurposing in medulloblastoma

Lipid-lowering agents

Statins are frequently prescribed drugs for the treatment of high cholesterol levels in the blood [[78\]](#page-26-0). Studies have reported anti-tumor effects of statins and decreased cancer-related mortality among patients with different tumor types who are taking statins; hence, they have been proposed for drug repurposing studies [[79\]](#page-26-0). Statins are effective competitive inhibitors of β-hydroxy βmethylglutaryl-CoA (HMG-CoA) reductase [\[80\]](#page-26-0), which is a key enzyme in the cholesterol synthesis pathway [\[81,82](#page-26-0)]. Inhibition of HMG-CoA reductase via statins decreases production of mevalonate derivatives that are essential for many growth regulatory processes [[83](#page-26-0)] such as proliferation, apoptosis, and differentiation [\[84](#page-26-0)].

Anti-cancer effects of lovastatin, a well-known and widely used statin, have been reported in many studies [\[85](#page-26-0)–[87](#page-26-0)]. In one in vitro study, lovastatin induced apoptosis, reduced cell viability, and inhibited cell proliferation of MB cell lines

[[88](#page-26-0)], and in another it induced DNA laddering and shrinkage of MB cells [\[83](#page-26-0)]. The anti-proliferative and apoptotic effects of lovastatin are mediated by modulation of $p27^{K1P1}$, P53 and $p21^{WAF1}$ gene expression [\[89](#page-27-0)]. Takwi et al. also showed that lovastatin upregulates miR-33 gene expression, thus decreasing cell proliferation and inducing apoptosis in vitro, besides reducing tumor growth and improving survival in MB tumor-bearing mice in vivo [\[90](#page-27-0)].

Simvastatin, another antihyperlipidemic drug, was shown to induce apoptotic cell death in a time- and dose-dependent manner in MB cell lines in comparison to the control group via activation of different caspases (Caspase 3, 7, 8 and 9) and down-expression of anti-apoptotic Bcl-2 and Mcl-1 proteins [[91\]](#page-27-0). An in vivo study revealed that simvastatin treatment of Ptch1^{+/-} CB17/SCID mice repressed the Shh signaling pathway with a significant decline in glioma-associated oncogene 1 (Gli1) expression level in MB cells due to disruption of cholesterol biosynthesis [[92\]](#page-27-0). In addition, simvastatin decreased the number of proliferating cells, reduced MB growth, induced apoptosis, and inhibited SHH-MB progression when combined with vismodegib [[92](#page-27-0)].

In the same milieu, fenofibrate—which belongs to the "cholesterol-lowering drugs" family—is usually prescribed as a monotherapy for treating elevated triglyceride levels in patients with severe hypertriglyceridemia [[93\]](#page-27-0). This drug was found to induce peroxisome proliferator activated receptor alpha (PPARα)–mediated cell cycle arrest and apoptosis in BsB8 mouse MB cell line as well as human MB Daoy and D283 cell lines [\[94](#page-27-0)]. Fenofibrate also inhibited IGF-I-induced phosphorylation events, which in turn attenuated the clonogenic growth of MB cells [\[94\]](#page-27-0).

Cardiac glycosides

Cardiac glycosides (CGs) comprise a wide range of naturally derived steroid compounds prescribed for chronic heart failure [\[95\]](#page-27-0). They inhibit the enzyme sodium-potassium ATPase (Na⁺/K⁺-ATPase) [\[96,95](#page-27-0)] leading to depletion of potassium and raising sodium and calcium concentrations within cells [[97](#page-27-0)]. Many studies investigated the anti-neoplastic effects of CGs mainly by inhibiting Na⁺/K⁺-ATPase [[97](#page-27-0),[98](#page-27-0)]. CGs also exhibited antiproliferative and proapoptotic properties in tumor cells by activation of sarcoma (SRC) kinase and epidermal growth factor receptor (EGFR), prompting activation of mitogen-activated protein kinase (MAPK)-ERK1/2 signaling pathway and upregulating cell cycle inhibitor $P21^{CIP1}$ [\[99\]](#page-27-0). Moreover, a systematic in vivo study revealed that digoxin inhibits cell growth, induces apoptosis, and instigates modulation of gene expression related to ERK/AKT signaling pathway in groups 3 and 4 MB cell lines (MED8A and D283). It also significantly prolonged survival in orthotopic PDX mice models when combined with radiation therapy [[100\]](#page-27-0). In the same study, proscillaridin, another CG, was reported to decrease cell viability in both group 3 and 4 MB cell lines (MED8A and D283) [[100\]](#page-27-0). In the same milieu, Wolle et al. studied the interplay between ouabain and EGF signaling and demonstrated that this CG inhibits EGF-induced Erk1/2-Akt activation, and attenuated EGF-induced actin reorganization, prevented stress fiber formation, and inhibited MB cell motility [[101\]](#page-27-0).

erative and cytotoxic properties [[123](#page-28-0)]. Valganciclovir, anti-human cytomegalovirus (HCMV) drug, was shown to inhibit MB tumor growth in vitro and in vivo and reduce tumorigenic and clonogenic capacity of D324 MED, D283 MED, and UW228-3 cell lines by targeting DNA polymerases and decreasing prostaglandin E2 (PGE2) synthesis [[124](#page-28-0)]. Abacavir (ABC), one of the most effective drugs for acquired immunodeficiency syndrome (AIDS) [[125\]](#page-28-0), is well

characterized by its telomerase inhibition activity and termination of DNA elongation [\[126](#page-28-0)]. Rossi et al. showed that abacavir treatment decreased cellular growth, disrupted cell cycle progression, and reduced proliferation of Daoy and D283MED MB cell lines [[127\]](#page-28-0). Interestingly, both MB cell lines showed substantial senescence features after abacavir treatment [\[127](#page-28-0)].

NSAIDs

Nonsteroidal anti-inflammatory drugs (NSAIDs) are highly selective cyclooxygenase (COX) inhibitors that suppress prostaglandin (PG) synthesis [[128](#page-28-0)]. Recent studies have provided evidence that NSAIDs might serve as potential candidates for cancer treatment in various tumor types [\[129](#page-28-0)–[134](#page-29-0)]. Celecoxib, a NSAID, has been used in many treatment protocols for pediatric tumors due to its anti-angiogenesis and immune-modulating effects [\[135](#page-29-0)]. It was shown to induce in vitro and in vivo anti-tumor effects in MB [\[124\]](#page-28-0) via downregulating the expression of phosphorylated-STAT3 and STAT3-related genes (JAK2, BCL2, and c -MYC) [[136\]](#page-29-0). It also enhanced the efficacy of radiotherapy [\[137](#page-29-0)] and synergistically improved survival of MB-bearing mice [[136\]](#page-29-0). Noticeably, in vivo results indicated a potential role of celecoxib in inhibiting angiogenesis gene expression (KDR, VEGFC and PDGFRA) as well as stem cell genes (Nestin, CD133, Sox-2, MSI1, and Bmi-1) [[137\]](#page-29-0). These potential effects of celecoxib in MB were also documented by Baryawno et al., suggesting the prospective benefit of celecoxib for treating patients with MB [\[138\]](#page-29-0).

Diclofenac, another COX-1/COX-2 inhibitor, was shown to reduce cell viability, proliferation, and colony formation in vitro and to further decrease microvascular density and tumor size in vivo [\[138](#page-29-0)]. Tolfenamic acid, another NSAID, also caused a decrease in tumor weight and volume by about 40% in MB athymic nude mice model mediated by a decrease in targeting specificity protein 1 (Sp1) and survivin expression [[139](#page-29-0)] that are implicated in cell proliferation, differentiation growth [\[140](#page-29-0)], and apoptosis [\[141\]](#page-29-0) of MB. Another in vitro study investigated the effectiveness of NSAIDs as antitumor drugs and revealed that flurbiprofen treatment suppressed the growth of MB cells and disrupted cell cycle progression via modulating cyclin B and P53 protein expressions [\[142](#page-29-0)].

Drugs used for neurological disorders

Medications used in neurological disorders have recently gained huge popularity in oncology as treatment options for the complications resulting from cancer and its therapy among this population of patients [[143\]](#page-29-0) as well as being repurposed to treat the cancer itself [\[144](#page-29-0),[145](#page-29-0)]. Ether-a-go-go 2 (EAG2) potassium channels (which are present in brain regions [\[146](#page-29-0)]) are shown to be highly expressed in MB tissues and involved in MB development and progression [[147\]](#page-29-0) and hence are potential targets for the antipsychotic drug thioridazine (a potent EAG2 channel blocker) [[148\]](#page-29-0). Additionally, valproic acid, a histone deacetylase inhibitor (HDACi), decreased cell viability of MB cell lines when combined with cisplatin (CDDP) [[149\]](#page-29-0) and reduced tumorigenicity in vivo at a clinically safe concentration [\[150\]](#page-29-0). The antitumor activity of valproic acid was associated with histone hyperacetylation (H3 and H4) and modulation of several genes expression (P21, c-MYC, CDK4, and TP53) [[150\]](#page-29-0).

Lithium has been characterized by neurotrophic effects and considered as a main therapeutic drug for the treatment of bipolar disorder [\[151](#page-29-0),[152](#page-29-0)]. It has profound effects on cell cycle, metabolism, and cell proliferation by which it potently inhibits cell cycle and accelerates cell death [\[153\]](#page-29-0). An in vitro study revealed that lithium targeted GSK3-β, slowed cell metabolic activity, arrested cell cycle, and induced non-apoptotic cell death in Daoy andD283 MB cell lines [[154\]](#page-30-0). Another study documented that WNT activation by lithium improved the effect of radiotherapy in TP53 mutant MB cells [[155\]](#page-30-0) suggesting a therapeutic property of lithium when combined with radiation for MB treatment but specifically for the TP53-mutant subgroup. Other antipsychotic medicines, such as sertraline, chlorprothixene, and chlorpromazine, were documented to inhibit MB cell growth and sphere formation in Daoy cells via inhibition of REST/NRSF-mSin3 interaction [\[156](#page-30-0)], raising the opportunity of new drug candidates for MB.

Dermatological drugs

Drugs used in dermatological diseases have also shown potential anti-tumor effects in MB. For instance, 13-cis retinoic acid, a dermatological drug, was shown to reduce cell viability and induce apoptosis via activating of bone morphogenetic protein-2 (BMP-2) synergistically with SAHA, a histone deacetylase inhibitor [[157\]](#page-30-0). It also enhanced differentiation of D283 cells and reduced tumor volume in xenograft MB models [[158\]](#page-30-0). All-trans-retinoic acid (ATRA) induced a dose-dependent decrease in cell viability and promoted apoptosis by activating caspase-3/poly(ADP-ribose) polymerase I effector pathway [\[159\]](#page-30-0). In addition, ATRA significantly inhibited cell and clonal proliferation and induced cellular differentiation of MB cells [\[159](#page-30-0)].

Other drugs

The antitumor effects of many other drugs were also studied in MB. Rapamycin, a well-known immunosuppressant drug, instigated a potent effect against cell proliferation and migration [\[160](#page-30-0)]. In other CNS tumors such as glioblastoma and neuroblastoma, rapamycin has proved to be effective in inducing antitumor effects in vitro, by targeting a cancer stem cell (CSC) subpopulation of cells [\[161](#page-30-0)]. In MB, treatment of Daoy cell lines with rapamycin showed a marked anti-tumor effects via inhibition of mTOR signaling [\[162](#page-30-0)]. Fingolimod, another immunosuppressant drug used in multiple sclerosis, was reported to have a therapeutic potential in MB, where it arrested cell cycle, decreased viability and proliferation, and enhanced apoptosis of MB cells, besides decreasing tumor growth in vivo [[163](#page-30-0)].

Unexpectedly, contradictory results were obtained following MB treatment with 17β-estradiol. Mancuso et al. showed an antiproliferative effect of 17βestradiol in Ptch1-associated MB [\[164](#page-30-0)]. However, Belcher et al. documented the potential of estradiol to increase MB cell growth and migration [[165\]](#page-30-0), whereas 17β-estradiol treatment did not affect cell proliferation at any concentration tested in MB cell lines [\[166](#page-30-0)].

Sulfasalazine, an inhibitor of nuclear factor Kappa-B (NF B) signaling, was demonstrated to inhibit cell growth in MB cell lines [[167](#page-30-0)]. The antitumor therapeutic effect of phenformin, an antidiabetic drug, was also tested in vitro and in vivo where phenformin treatment induced a significant inhibition of MB

Conclusions

cell growth and proliferation [\[168](#page-30-0)]. Interestingly, another antidiabetic medication metformin showed anti-tumor effects in glioblastoma and neuroblastoma CNS tumors via targeting CSCs in vitro [\[169](#page-30-0)].

Disulfiram, an FDA-approved drug for treatment of alcoholism [[170\]](#page-30-0), has been repurposed to study its anticancer effects owning to its multiple pharmacological mechanisms in targeting tumor cells and triggering oxidative stress [[171\]](#page-30-0), activating MAPK pathway [\[172](#page-30-0)], and suppressing the proteasome system [[173\]](#page-30-0). In MB, disulfiram was shown to induce apoptosis and decrease cell viability and colony formation in Daoy cell line. It also induced a significant regression of tumor growth in MB xenografts [[174\]](#page-30-0).

In order to provide more comprehensive care for patients with cancer, it is crucial to decipher the mechanisms of action pertaining to the different drugs used clinically and understand the pleiotropic adverse effects and perspective interactions they might have with other medications. However, the dire need for developing and finding more efficient antitumor drugs urges the scientific society to change its approach towards seeking new strategies, most importantly via drug repurposing, to reach its goals in treating cancer patients and improving their quality of life. Repositioning previously FDA-approved drugs is indeed a promising strategy in cancer treatment particularly pediatric tumors owning to its various advantages including cost efficiency and shortened time-frame for safety pharmacology testing in drug development. At a clinical level, many FDA-approved drugs have been put under investigation in clinical trials on medulloblastoma patients [\[175](#page-31-0)], including anthelminthic drug "mebendazole" ([ClinicalTrials.gov;](http://clinicaltrials.gov) phase I clinical trial; NCT02644291), cholinesterase inhibitor "donepezil" that is used to treat Alzheimer's disease [\(ClinicalTrials.gov](http://clinicaltrials.gov); phase I clinical trial; NCT00452868), bradykinin B-2 receptor agonist "lobradimil" [\(ClinicalTrials.gov](http://clinicaltrials.gov); phase II clinical trial; NCT00019422), and acetylcysteine, mannitol, and sodium thiosulfate ([ClinicalTrials.gov](http://clinicaltrials.gov); phase I clinical trial; NCT00238173).

Although treatment by repurposing drugs might seem to be a long way ahead to achieve, it carries potential hope for managing cancer in general and MB in particular. This novel therapeutic approach could help thousands of MB patients suffering worldwide and awaiting more efficient therapies to come up for their disease. Further experimental and clinical studies are needed to establish repurposed drugs as adjuvant remedies for MB and other tumors.

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Not applicable.

Code availability

Not applicable.

Authors' contributions

H.F. Bahmad and S. Nabha conceived the concept and idea of the present review. H.F. Bahmad and S. Nabha worked on the study design strategy and selected the topics to be discussed. H. Hammoud, Z. Saker, H.F. Bahmad, and S. Nabha did literature searches and screened titles and abstracts for relevance. H. Hammoud and Z. Saker abstracted the data from the eligible full text articles, analyzed and interpreted the data, and drafted the manuscript. H. Harati and Y. Fares revised the final draft of the manuscript. H.F. Bahmad and S. Nabha critically revised the manuscript with input from the entire team. All authors have read and approved the final draft.

Compliance with ethical standards

Conflict of interest

The authors declare that they have no conflict of interest.

Ethics approval Not applicable.

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References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- •• Of major importance
- 1. Kumar LP, Deepa SFAJ, Moinca I, Suresh P, Naidu KVJR. Medulloblastoma: a common pediatric tumor: prognostic factors and predictors of outcome. Asian J Neurosurg. 2015;10(1):50. [https://doi.org/10.4103/](http://dx.doi.org/10.4103/1793-5482.151516) [1793-5482.151516](http://dx.doi.org/10.4103/1793-5482.151516).
- 2. Parkes J, Hendricks M, Ssenyonga P, Mugamba J, Molyneux E, Schouten-van Meeteren A, et al. SIOP PODC adapted treatment recommendations for

standard-risk medulloblastoma in low and middle income settings. Pediatric Blood & Cancer. 2015;62(4):553–64. [https://doi.org/10.1002/pbc.](http://dx.doi.org/10.1002/pbc.25313) [25313](http://dx.doi.org/10.1002/pbc.25313).

3.• Bahmad HF, Poppiti RJ (2020) Medulloblastoma cancer stem cells: molecular signatures and therapeutic targets. Journal of clinical pathology 73 (5):243-249. doi:10.1136/jclinpath-2019-206246.

Reason: Comprehensive review providing a synopsis of the novel therapeutic approaches that specifically target medulloblastoma cancer stem cells to attain enhanced anti-tumorous effects and overcome therapy resistance.

- 4. Millard NE, De Braganca KC. Medulloblastoma. J Child Neurol. 2016;31(12):1341–53. [https://doi.org/](http://dx.doi.org/10.1177/0883073815600866) [10.1177/0883073815600866.](http://dx.doi.org/10.1177/0883073815600866)
- 5. Rossi A, Caracciolo V, Russo G, Reiss K, Giordano A. Medulloblastoma: from molecular pathology to therapy. Clin Cancer Res. 2008;14(4):971–6. [https://doi.](http://dx.doi.org/10.1158/1078-0432.CCR-07-2072) [org/10.1158/1078-0432.CCR-07-2072.](http://dx.doi.org/10.1158/1078-0432.CCR-07-2072)
- 6. Gilbertson RJ, Ellison DW. The origins of medulloblastoma subtypes. Annual Review of Pathology: Mechanisms of Disease. 2008;3(1):341–65. [https://](http://dx.doi.org/10.1146/annurev.pathmechdis.3.121806.151518) [doi.org/10.1146/annurev.pathmechdis.3.121806.](http://dx.doi.org/10.1146/annurev.pathmechdis.3.121806.151518) [151518.](http://dx.doi.org/10.1146/annurev.pathmechdis.3.121806.151518)
- 7. Yokota N, Aruga J, Takai S, Yamada K, Hamazaki M, Iwase T, et al. Predominant expression of human Zic in cerebellar granule cell lineage and medulloblastoma. Cancer Research. 1996;56(2):377.
- 8. Behesti H, Marino S. Cerebellar granule cells: insights into proliferation, differentiation, and role in medulloblastoma pathogenesis. The International Journal of Biochemistry & Cell Biology. 2009;41(3):435–45. [https://doi.org/10.1016/j.biocel.2008.06.017.](http://dx.doi.org/10.1016/j.biocel.2008.06.017)
- 9. Ostrom QT, Gittleman H, Truitt G, Boscia A, Kruchko C, Barnholtz-Sloan JS (2018) CBTRUS statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2011-2015. Neuro Oncol 20 (suppl_4):iv1-iv86. doi[:https://doi.org/10.](http://dx.doi.org/10.1093/neuonc/noy131) [1093/neuonc/noy131](http://dx.doi.org/10.1093/neuonc/noy131)
- 10. Esbenshade AJ, Kocak M, Hershon L, Rousseau P, Decarie J-C, Shaw S, Burger P, Friedman HS, Gajjar A, Moghrabi A (2017) A Phase II feasibility study of oral etoposide given concurrently with radiotherapy followed by dose intensive adjuvant chemotherapy for children with newly diagnosed high-risk medulloblastoma (protocol POG 9631): a report from the Children's Oncology Group. Pediatric blood & cancer 64 (6):10.1002/pbc.26373. doi:[https://doi.org/10.1002/](http://dx.doi.org/10.1002/pbc.26373) [pbc.26373](http://dx.doi.org/10.1002/pbc.26373)
- 11. von Bueren AO, Kortmann R-D, von Hoff K, Friedrich C, Mynarek M, Müller K, Goschzik T, zur Mühlen A, Gerber N, Warmuth-Metz M, Soerensen N, Deinlein F, Benesch M, Zwiener I, Kwiecien R, Faldum A, Bode U, Fleischhack G, Hovestadt V, Kool M, Jones D, Northcott P, Kuehl J, Pfister S, Pietsch T, Rutkowski S (2016) Treatment of children and adolescents with metastatic medulloblastoma and prognostic relevance of clinical and biologic parameters. Journal of Clinical Oncology 34 (34):4151-4160. doi[:https://doi.org/10.](http://dx.doi.org/10.1200/JCO.2016.67.2428) [1200/JCO.2016.67.2428](http://dx.doi.org/10.1200/JCO.2016.67.2428)
- 12. Muzumdar D, Deshpande A, Kumar R, Sharma A, Goel N, Dange N, et al. Medulloblastoma in childhood-King Edward Memorial hospital surgical experience and review: comparative analysis of the case series of 365

patients. J Pediatr Neurosci. 2011;6(Suppl 1):S78–85. [https://doi.org/10.4103/1817-1745.85717.](http://dx.doi.org/10.4103/1817-1745.85717)

- 13. Rajagopal R, Abd-Ghafar S, Ganesan D, Bustam Mainudin AZ, Wong KT, Ramli N, et al. Challenges of treating childhood medulloblastoma in a country with limited resources: 20 years of experience at a single tertiary center in Malaysia. J Glob Oncol. 2016;3(2):143–56. [https://doi.org/10.1200/JGO.](http://dx.doi.org/10.1200/JGO.2015.002659) [2015.002659.](http://dx.doi.org/10.1200/JGO.2015.002659)
- 14. Louis DN, Ohgaki H, Wiestler OD, Cavenee WK, Burger PC, Jouvet A, et al. The 2007 WHO classification of tumours of the central nervous system. Acta Neuropathol. 2007;114(2):97–109. [https://doi.org/](http://dx.doi.org/10.1007/s00401-007-0243-4) [10.1007/s00401-007-0243-4.](http://dx.doi.org/10.1007/s00401-007-0243-4)
- 15. Crawford JR, MacDonald TJ, Packer RJ. Medulloblastoma in childhood: new biological advances. The Lancet Neurology. 2007;6(12):1073–85. [https://doi.org/](http://dx.doi.org/10.1016/s1474-4422(07)70289-2) [10.1016/s1474-4422\(07\)70289-2.](http://dx.doi.org/10.1016/s1474-4422(07)70289-2)
- 16. Giangaspero F, Perilongo G, Fondelli MP, Brisigotti M, Carollo C, Burnelli R, et al. Medulloblastoma with extensive nodularity: a variant with favorable prognosis. Journal of Neurosurgery. 1999;91(6):971–7. [https://doi.org/10.3171/jns.1999.91.6.0971.](http://dx.doi.org/10.3171/jns.1999.91.6.0971)
- 17. Lamont JM, McManamy CS, Pearson AD, Clifford SC, Ellison DW. Combined histopathological and molecular cytogenetic stratification of medulloblastoma patients. Clinical Cancer Research. 2004;10(16):5482. [https://doi.org/10.1158/1078-0432.CCR-03-0721](http://dx.doi.org/10.1158/1078-0432.CCR-03-0721).
- 18. Kleihues P, Louis DN, Scheithauer BW, Rorke LB, Reifenberger G, Burger PC, et al. The WHO Classification of tumors of the nervous system. Journal of Neuropathology & Experimental Neurology. 2002;61(3):215–25. [https://doi.org/10.1093/jnen/61.](http://dx.doi.org/10.1093/jnen/61.3.215) [3.215](http://dx.doi.org/10.1093/jnen/61.3.215).
- 19. Louis DN, Ohgaki H, Wiestler OD, Cavenee WK. World health organization classification of tumours of the central nervous system, vol. 1. Revised. 4th ed. Lyon: IARC Press; 2016.
- 20. Kawauchi D, Robinson G, Uziel T, Gibson P, Rehg J, Gao C, et al. A mouse model of the most aggressive subgroup of human medulloblastoma. Cancer cell. 2012;21(2):168–80. [https://doi.org/10.1016/j.ccr.](http://dx.doi.org/10.1016/j.ccr.2011.12.023) [2011.12.023.](http://dx.doi.org/10.1016/j.ccr.2011.12.023)
- 21. Pei Y, Moore CE, Wang J, Tewari AK, Eroshkin A, Cho YJ, et al. An animal model of MYC-driven medulloblastoma. Cancer cell. 2012;21(2):155–67. [https://doi.](http://dx.doi.org/10.1016/j.ccr.2011.12.021) [org/10.1016/j.ccr.2011.12.021](http://dx.doi.org/10.1016/j.ccr.2011.12.021).
- 22. Swartling FJ, Grimmer MR, Hackett CS, Northcott PA, Fan QW, Goldenberg DD, et al. Pleiotropic role for MYCN in medulloblastoma. Genes & development. 2010;24(10):1059–72. [https://doi.org/10.1101/gad.](http://dx.doi.org/10.1101/gad.1907510) [1907510.](http://dx.doi.org/10.1101/gad.1907510)
- 23. Swartling FJ, Savov V, Persson AI, Chen J, Hackett CS, Northcott PA, et al. Distinct neural stem cell populations give rise to disparate brain tumors in response to

N-MYC. Cancer cell. 2012;21(5):601–13. [https://doi.](http://dx.doi.org/10.1016/j.ccr.2012.04.012) [org/10.1016/j.ccr.2012.04.012](http://dx.doi.org/10.1016/j.ccr.2012.04.012).

24.• Menyhárt O, Giangaspero F, Győrffy B (2019) Molecular markers and potential therapeutic targets in non-WNT/non-SHH (group 3 and group 4) medulloblastomas. Journal of Hematology & Oncology 12 (1):29. doi:10.1186/s13045-019-0712-y.

Reason: Review delineating the molecular aberrations involved in MB tumorigenesis, particularly groups 3 and 4, paving the way to identifying potential therapeutic targets for drug repurposing in medulloblastoma.

- 25. Taylor MD, Northcott PA, Korshunov A, Remke M, Cho Y-J, Clifford SC, et al. Molecular subgroups of medulloblastoma: the current consensus. Acta Neuropathol. 2012;123(4):465–72. [https://doi.org/](http://dx.doi.org/10.1007/s00401-011-0922-z) [10.1007/s00401-011-0922-z.](http://dx.doi.org/10.1007/s00401-011-0922-z)
- 26. Mir SE, Smits M, Biesmans D, Julsing M, Bugiani M, Aronica E, Kaspers GJL, Cloos J, Würdinger T, Hulleman E (2017) Trimethylation of H3K27 during human cerebellar development in relation to medulloblastoma. Oncotarget 8 (45):78978-78988. doi:10.18632/oncotarget.20741
- 27. Robbins CJ, Bou-Dargham MJ, Sanchez K, Rosen MC, Sang Q-XA. Decoding somatic driver gene mutations and affected signaling pathways in human medulloblastoma subgroups. J Cancer. 2018;9(24):4596–610. [https://doi.org/10.7150/jca.27993](http://dx.doi.org/10.7150/jca.27993).
- 28. Fattet S, Haberler C, Legoix P, Varlet P, Lellouch-Tubiana A, Lair S, et al. Beta-catenin status in paediatric medulloblastomas: correlation of immunohistochemical expression with mutational status, genetic profiles, and clinical characteristics. The Journal of pathology. 2009;218(1):86–94. [https://doi.org/10.1002/path.](http://dx.doi.org/10.1002/path.2514) [2514](http://dx.doi.org/10.1002/path.2514).
- 29. Yokota N, Nishizawa S Fau Ohta S, Ohta S Fau Date H, Date H Fau - Sugimura H, Sugimura H Fau - Namba H, Namba H Fau - Maekawa M, Maekawa M Role of Wnt pathway in medulloblastoma oncogenesis. (0020-7136 (Print))
- 30. Thompson MC, Fuller C, Hogg TL, Dalton J, Finkelstein D, Lau CC, et al. Genomics identifies medulloblastoma subgroups that are enriched for specific genetic alterations. J Clin Oncol. 2006;24(12):1924– 31. [https://doi.org/10.1200/jco.2005.04.4974.](http://dx.doi.org/10.1200/jco.2005.04.4974)
- 31. Clifford SC, Lusher ME, Lindsey JC, Langdon JA, Gilbertson RJ, Straughton D, et al. Wnt/wingless pathway activation and chromosome 6 loss characterise a distinct molecular sub-group of medulloblastomas associated with a favourable prognosis. Cell Cycle. 2006;5(22):2666–70. [https://doi.org/10.4161/cc.5.22.](http://dx.doi.org/10.4161/cc.5.22.3446) [3446](http://dx.doi.org/10.4161/cc.5.22.3446).
- 32. Northcott PA, Korshunov A, Witt H, Hielscher T, Eberhart CG, Mack S, et al. Medulloblastoma comprises four distinct molecular variants. J Clin Oncol. 2011;29(11):1408–14. [https://doi.org/10.1200/JCO.](http://dx.doi.org/10.1200/JCO.2009.27.4324) [2009.27.4324.](http://dx.doi.org/10.1200/JCO.2009.27.4324)
- 33. Cambruzzi E. Medulloblastoma, WNT-activated/SHHactivated: clinical impact of molecular analysis and histogenetic evaluation. Child's Nervous System. 2018;34(5):809–15. [https://doi.org/10.1007/s00381-](http://dx.doi.org/10.1007/s00381-018-3765-2) [018-3765-2](http://dx.doi.org/10.1007/s00381-018-3765-2).
- 34. Higdon R, Kala J, Wilkins D, Yan JF, Sethi MK, Lin L, et al. Integrated proteomic and transcriptomic-based approaches to identifying signature biomarkers and pathways for elucidation of Daoy and UW228 subtypes. Proteomes. 2017;5(1):5. [https://doi.org/10.](http://dx.doi.org/10.3390/proteomes5010005) [3390/proteomes5010005](http://dx.doi.org/10.3390/proteomes5010005).
- 35. Northcott PA, Shih DJH, Remke M, Cho Y-J, Kool M, Hawkins C, et al. Rapid, reliable, and reproducible molecular sub-grouping of clinical medulloblastoma samples. Acta Neuropathol. 2012;123(4):615–26. [https://doi.org/10.1007/s00401-011-0899-7](http://dx.doi.org/10.1007/s00401-011-0899-7).
- 36. Taylor MD, Liu L, Raffel C, C-c H, Mainprize TG, Zhang X, et al. Mutations in SUFU predispose to medulloblastoma. Nature Genetics. 2002;31(3):306–10. [https://doi.org/10.1038/ng916.](http://dx.doi.org/10.1038/ng916)
- 37. Menyhárt O, Győrffy B. Principles of tumorigenesis and emerging molecular drivers of SHH-activated medulloblastomas. Ann Clin Transl Neurol. 2019;6(5):990–1005. [https://doi.org/10.1002/acn3.](http://dx.doi.org/10.1002/acn3.762) [762](http://dx.doi.org/10.1002/acn3.762).
- 38. Roussel MF, Robinson GW. Role of MYC in medulloblastoma. Cold Spring Harb Perspect Med. 2013;3(11):a014308. [https://doi.org/10.1101/](http://dx.doi.org/10.1101/cshperspect.a014308) [cshperspect.a014308](http://dx.doi.org/10.1101/cshperspect.a014308).
- 39. Tao R, Murad N, Xu Z, Zhang P, Okonechnikov K, Kool M, et al. MYC drives group 3 medulloblastoma through transformation of Sox2(+) astrocyte progenitor cells. Cancer research. 2019;79(8):1967–80. [https://doi.org/10.1158/0008-5472.CAN-18-1787](http://dx.doi.org/10.1158/0008-5472.CAN-18-1787).
- 40. Sengupta S, Pomeranz Krummel D, Pomeroy S (2017) The evolution of medulloblastoma therapy to personalized medicine. F1000Res 6:490-490. doi:10.12688/ f1000research.10859.1
- 41. Park AK, Lee JY, Cheong H, Ramaswamy V, Park S-H, Kool M, et al. Subgroup-specific prognostic signaling and metabolic pathways in pediatric medulloblastoma. BMC Cancer. 2019;19(1):571. [https://doi.org/10.](http://dx.doi.org/10.1186/s12885-019-5742-x) [1186/s12885-019-5742-x](http://dx.doi.org/10.1186/s12885-019-5742-x).
- 42. Thomas A, Noël G. Medulloblastoma: optimizing care with a multidisciplinary approach. J Multidiscip Healthc. 2019;12:335–47. [https://doi.org/10.2147/](http://dx.doi.org/10.2147/JMDH.S167808) [JMDH.S167808.](http://dx.doi.org/10.2147/JMDH.S167808)
- 43. Rutkowski S, Gerber NU, von Hoff K, Gnekow A, Bode U, Graf N, Berthold F, Henze G, Wolff JEA, Warmuth-Metz M, Soerensen N, Emser A, Ottensmeier H, Deinlein F, Schlegel P-G, Kortmann R-D, Pietsch T, Kuehl J, German Pediatric Brain Tumor Study G. Treatment of early childhood medulloblastoma by postoperative chemotherapy and deferred radiotherapy. Neuro Oncol. 2009;11(2):201–10. [https://doi.org/](http://dx.doi.org/10.1215/15228517-2008-084) [10.1215/15228517-2008-084.](http://dx.doi.org/10.1215/15228517-2008-084)
- 44. Srinivasan VM, Ghali MGZ, North RY, Boghani Z, Hansen D, Lam S. Modern management of medulloblastoma: Molecular classification, outcomes, and the role of surgery. Surg Neurol Int. 2016;7(Suppl 44):S1135–41. [https://doi.org/10.4103/2152-7806.](http://dx.doi.org/10.4103/2152-7806.196922) [196922.](http://dx.doi.org/10.4103/2152-7806.196922)
- 45. Pols S, van Veelen MLC, Aarsen FK, Gonzalez Candel A, Catsman-Berrevoets CE. Risk factors for development of postoperative cerebellar mutism syndrome in children after medulloblastoma surgery. Journal of neurosurgery Pediatrics. 2017;20(1):35–41. [https://doi.org/](http://dx.doi.org/10.3171/2017.2.peds16605) [10.3171/2017.2.peds16605](http://dx.doi.org/10.3171/2017.2.peds16605).
- 46. Giambelli C, Fei DL, Wang H, Robbins DJ. Repurposing an old anti-fungal drug as a Hedgehog inhibitor. Protein & Cell. 2010;1(5):417–8. [https://doi.](http://dx.doi.org/10.1007/s13238-010-0063-5) [org/10.1007/s13238-010-0063-5](http://dx.doi.org/10.1007/s13238-010-0063-5).
- 47. Kim J, Tang JY, Gong R, Kim J, Lee JJ, Clemons KV, et al. Itraconazole, a commonly used antifungal that inhibits Hedgehog pathway activity and cancer growth. Cancer cell. 2010;17(4):388–99. [https://doi.org/10.1016/j.ccr.](http://dx.doi.org/10.1016/j.ccr.2010.02.027) [2010.02.027.](http://dx.doi.org/10.1016/j.ccr.2010.02.027)
- 48. Mulhern RK, Palmer SL, Merchant TE, Wallace D, Kocak M, Brouwers P, et al. Neurocognitive consequences of risk-adapted therapy for childhood medulloblastoma. J Clin Oncol. 2005;23(24):5511–9. [https://doi.org/10.1200/jco.2005.00.703.](http://dx.doi.org/10.1200/jco.2005.00.703)
- 49. Ivanov DP, Coyle B, Walker DA, Grabowska AM. In vitro models of medulloblastoma: Choosing the right tool for the job. Journal of Biotechnology. 2016;236:10–25. [https://doi.org/10.1016/j.jbiotec.](http://dx.doi.org/10.1016/j.jbiotec.2016.07.028) [2016.07.028.](http://dx.doi.org/10.1016/j.jbiotec.2016.07.028)
- 50. Duffner Pk Fau Cohen ME, Cohen Me Fau Thomas PR, Thomas Pr Fau - Sinks LF, Sinks Lf Fau - Freeman AI, Freeman AI Combination chemotherapy in recurrent medulloblastoma. (0008-543X (Print))
- 51. Mollashahi B, Aghamaleki FS, Movafagh A (2019) The roles of miRNAs in medulloblastoma: a systematic review. J Cancer Prev 24 (2):79-90. doi:10.15430/ JCP.2019.24.2.79
- 52. Bahmad HF, Chamaa F, Assi S, Chalhoub RM, Abou-Antoun T, Abou-Kheir W. Cancer stem cells in neuroblastoma: expanding the therapeutic frontier. Frontiers in molecular neuroscience. 2019;12:131. [https://doi.](http://dx.doi.org/10.3389/fnmol.2019.00131) [org/10.3389/fnmol.2019.00131.](http://dx.doi.org/10.3389/fnmol.2019.00131)
- 53. Ramaswamy V, Remke M, Bouffet E, Faria CC, Perreault S, Cho Y-J, et al. Recurrence patterns across medulloblastoma subgroups: an integrated clinical and molecular analysis. Lancet Oncol. 2013;14(12):1200–7. [https://doi.org/10.1016/S1470-](http://dx.doi.org/10.1016/S1470-2045(13)70449-2) [2045\(13\)70449-2.](http://dx.doi.org/10.1016/S1470-2045(13)70449-2)
- 54.•• Bahmad HF, Elajami MK, El Zarif T, Bou-Gharios J, Abou-Antoun T, Abou-Kheir W (2020) Drug repurposing towards targeting cancer stem cells in pediatric brain tumors. Cancer metastasis reviews 39 (1):127-148. doi:10.1007/s10555-019-09840-2.

Reason: review elucidating the drug repurposing methodologies that have been used in pediatric brain tumors and how this selective compilation of approaches could elevate drug repurposing to the next level.

- 55. Prasad V, Mailankody S. Research and development spending to bring a single cancer drug to market and revenues after approval. JAMA Intern Med. 2017;177(11):1569–75. [https://doi.org/10.1001/](http://dx.doi.org/10.1001/jamainternmed.2017.3601) [jamainternmed.2017.3601.](http://dx.doi.org/10.1001/jamainternmed.2017.3601)
- 56. Wu CH, Bai LY, Tsai MH, Chu PC, Chiu CF, Chen MY, et al. Pharmacological exploitation of the phenothiazine antipsychotics to develop novel antitumor agentsa drug repurposing strategy. Sci Rep. 2016;6:27540. [https://doi.org/10.1038/srep27540.](http://dx.doi.org/10.1038/srep27540)
- 57. Aggarwal S, Verma SS, Aggarwal S, Gupta SC. Drug repurposing for breast cancer therapy: old weapon for new battle. Seminars in Cancer Biology. 2019. [https://](http://dx.doi.org/10.1016/j.semcancer.2019.09.012) [doi.org/10.1016/j.semcancer.2019.09.012.](http://dx.doi.org/10.1016/j.semcancer.2019.09.012)
- 58. Giordano SH, Lin YL, Kuo YF, Hortobagyi GN, Goodwin JS. Decline in the use of anthracyclines for breast cancer. J Clin Oncol. 2012;30(18):2232–9. [https://doi.org/10.1200/jco.2011.40.1273.](http://dx.doi.org/10.1200/jco.2011.40.1273)
- 59. Nabhan C, Byrtek M, Rai A, Dawson K, Zhou X, Link BK, et al. Disease characteristics, treatment patterns, prognosis, outcomes and lymphoma-related mortality in elderly follicular lymphoma in the United States. Br J Haematol. 2015;170(1):85–95. [https://doi.org/10.](http://dx.doi.org/10.1111/bjh.13399) [1111/bjh.13399](http://dx.doi.org/10.1111/bjh.13399).
- 60. Chihara D, Westin JR, Oki Y, Ahmed MA, Do B, Fayad LE, et al. Management strategies and outcomes for very elderly patients with diffuse large B-cell lymphoma. Cancer. 2016;122(20):3145–51. [https://doi.org/10.](http://dx.doi.org/10.1002/cncr.30173) [1002/cncr.30173](http://dx.doi.org/10.1002/cncr.30173).
- 61. Pasquier E, Ciccolini J, Carre M, Giacometti S, Fanciullino R, Pouchy C, Montero MP, Serdjebi C, Kavallaris M, André N (2011) Propranolol potentiates the anti-angiogenic effects and anti-tumor efficacy of chemotherapy agents: implication in breast cancer treatment. Oncotarget 2 (10):797-809. doi:10.18632/ oncotarget.343
- 62. Dubus E, Ijjaali I, Barberan O, Petitet F. Drug repositioning using in silico compound profiling. Future Medicinal Chemistry. 2009;1(9):1723–36. [https://doi.](http://dx.doi.org/10.4155/fmc.09.123) [org/10.4155/fmc.09.123](http://dx.doi.org/10.4155/fmc.09.123).
- 63. Smith MA, Seibel NL, Altekruse SF, Ries LA, Melbert DL, O'Leary M, et al. Outcomes for children and adolescents with cancer: challenges for the twenty-first century. J Clin Oncol. 2010;28(15):2625–34. [https://](http://dx.doi.org/10.1200/JCO.2009.27.0421) [doi.org/10.1200/JCO.2009.27.0421](http://dx.doi.org/10.1200/JCO.2009.27.0421).
- 64. Brinkman TM, Krasin MJ, Liu W, Armstrong GT, Ojha RP, Sadighi ZS, et al. Long-term neurocognitive functioning and social attainment in adult survivors of pediatric CNS tumors: results from the St Jude Lifetime Cohort Study. J Clin Oncol. 2016;34(12):1358–67. [https://doi.org/10.1200/JCO.2015.62.2589.](http://dx.doi.org/10.1200/JCO.2015.62.2589)
- 65. Saker Z, Bahmad HF, Fares Y, Al Najjar Z, Saad M, Harati H, et al. Prognostic impact of adenylyl cyclaseassociated protein 2 (CAP2) in glioma: a clinicopathological study. Heliyon. 2020;6(1):e03236. [https://](http://dx.doi.org/10.1016/j.heliyon.2020.e03236) [doi.org/10.1016/j.heliyon.2020.e03236](http://dx.doi.org/10.1016/j.heliyon.2020.e03236).
- 66. Janzer RC, Raff MC. Astrocytes induce blood–brain barrier properties in endothelial cells. Nature.

1987;325(6101):253–7. [https://doi.org/10.1038/](http://dx.doi.org/10.1038/325253a0) [325253a0.](http://dx.doi.org/10.1038/325253a0)

- 67. Biegel D, Spencer DD, Pachter JS. Isolation and culture of human brain microvessel endothelial cells for the study of blood-brain barrier properties in vitro. Brain Res. 1995;692(1-2):183–9. [https://doi.org/10.1016/](http://dx.doi.org/10.1016/0006-8993(95)00511-n) [0006-8993\(95\)00511-n](http://dx.doi.org/10.1016/0006-8993(95)00511-n).
- 68. Ben-Zvi A, Lacoste B, Kur E, Andreone BJ, Mayshar Y, Yan H, et al. Mfsd2a is critical for the formation and function of the blood–brain barrier. Nature. 2014;509(7501):507–11. [https://doi.org/10.1038/](http://dx.doi.org/10.1038/nature13324) [nature13324](http://dx.doi.org/10.1038/nature13324).
- 69. Thorsen F, Fite B, Mahakian LM, Seo JW, Qin S, Harrison V, et al. Multimodal imaging enables early detection and characterization of changes in tumor permeability of brain metastases. J Control Release. 2013;172(3):812–22. [https://doi.org/10.1016/j.](http://dx.doi.org/10.1016/j.jconrel.2013.10.019) [jconrel.2013.10.019.](http://dx.doi.org/10.1016/j.jconrel.2013.10.019)
- 70. Carbonell WS, Ansorge O, Sibson N, Muschel R. The vascular basement membrane as "soil" in brain metastasis. PLOS ONE. 2009;4(6):e5857. [https://doi.org/](http://dx.doi.org/10.1371/journal.pone.0005857) [10.1371/journal.pone.0005857.](http://dx.doi.org/10.1371/journal.pone.0005857)
- 71. Palmieri D, Chambers AF, Felding-Habermann B, Huang S, Steeg PS. The biology of metastasis to a sanctuary site. Clinical Cancer Research. 2007;13(6):1656–62. [https://doi.org/10.1158/1078-](http://dx.doi.org/10.1158/1078-0432.ccr-06-2659) [0432.ccr-06-2659](http://dx.doi.org/10.1158/1078-0432.ccr-06-2659).
- 72. Kakee A, Terasaki T, Sugiyama Y. Brain efflux index as a novel method of analyzing efflux transport at the blood-brain barrier. J Pharmacol Exp Ther. 1996;277(3):1550–9.
- 73. Wager TT, Hou X, Verhoest PR, Villalobos A. Moving beyond rules: the development of a central nervous system multiparameter optimization (CNS MPO) approach to enable alignment of druglike properties. ACS Chemical Neuroscience. 2010;1(6):435–49. [https://](http://dx.doi.org/10.1021/cn100008c) [doi.org/10.1021/cn100008c.](http://dx.doi.org/10.1021/cn100008c)
- 74. Lilius TO, Blomqvist K, Hauglund NL, Liu G, Stæger FF, Bærentzen S, Du T, Ahlström F, Backman JT, Kalso EA (2019) Dexmedetomidine enhances glymphatic brain delivery of intrathecally administered drugs.
- 75. Iliff JJ, Wang M, Liao Y, Plogg BA, Peng W, Gundersen GA, et al. A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid β. Sci Transl Med 4 (147):147ra111. 2012. [https://doi.org/10.1126/](http://dx.doi.org/10.1126/scitranslmed.3003748) [scitranslmed.3003748](http://dx.doi.org/10.1126/scitranslmed.3003748).
- 76. Jones DT, Jager N, Kool M, Zichner T, Hutter B, Sultan M, et al. Dissecting the genomic complexity underlying medulloblastoma. Nature. 2012;488(7409):100–5. [https://doi.org/10.1038/nature11284](http://dx.doi.org/10.1038/nature11284).
- 77. Phoenix TN, Patmore DM, Boop S, Boulos N, Jacus MO, Patel YT, et al. Medulloblastoma genotype dictates blood brain barrier phenotype. Cancer cell. 2016;29(4):508–22. [https://doi.org/10.1016/j.ccell.](http://dx.doi.org/10.1016/j.ccell.2016.03.002) [2016.03.002.](http://dx.doi.org/10.1016/j.ccell.2016.03.002)
- 78. Baigent C, Keech A, Kearney PM, Blackwell L, Buck G, Pollicino C, et al. Efficacy and safety of cholesterollowering treatment: prospective meta-analysis of data

from 90,056 participants in 14 randomised trials of statins. Lancet. 2005;366(9493):1267–78. [https://doi.](http://dx.doi.org/10.1016/S0140-6736(05)67394-1) [org/10.1016/S0140-6736\(05\)67394-1](http://dx.doi.org/10.1016/S0140-6736(05)67394-1).

- 79. Nielsen SF, Nordestgaard BG, Bojesen SE. Statin use and reduced cancer-related mortality. New England Journal of Medicine. 2012;367(19):1792–802. [https://](http://dx.doi.org/10.1056/NEJMoa1201735) [doi.org/10.1056/NEJMoa1201735.](http://dx.doi.org/10.1056/NEJMoa1201735)
- 80. Ginsberg HN, Le NA, Short MP, Ramakrishnan R, Desnick RJ. Suppression of apolipoprotein B production during treatment of cholesteryl ester storage disease with lovastatin. Implications for regulation of apolipoprotein B synthesis. J Clin Invest. 1987;80(6):1692–7. [https://doi.org/10.1172/](http://dx.doi.org/10.1172/JCI113259) [JCI113259.](http://dx.doi.org/10.1172/JCI113259)
- 81. Endo A, Tsujita Y, Kuroda M, Tanzawa K. Inhibition of cholesterol synthesis in vitro and in vivo by ML-236A and ML-236B, competitive inhibitors of 3-hydroxy-3 methylglutaryl-coenzyme A reductase. Eur J Biochem. 1977;77(1):31–6. [https://doi.org/10.1111/j.1432-](http://dx.doi.org/10.1111/j.1432-1033.1977.tb11637.x) [1033.1977.tb11637.x](http://dx.doi.org/10.1111/j.1432-1033.1977.tb11637.x).
- 82. Tobert JA, Hitzenberger G, Kukovetz WR, Holmes IB, Jones KH. Rapid and substantial lowering of human serum cholesterol by mevinolin (MK-803), an inhibitor of hydroxymethylglutaryl-coenzyme A reductase. Atherosclerosis. 1982;41(1):61–5. [https://doi.org/10.](http://dx.doi.org/10.1016/0021-9150(82)90070-3) [1016/0021-9150\(82\)90070-3.](http://dx.doi.org/10.1016/0021-9150(82)90070-3)
- 83. Wang W, Macaulay RJ. Mevalonate prevents lovastatininduced apoptosis in medulloblastoma cell lines. Can J Neurol Sci. 1999;26(4):305–10. [https://doi.org/10.](http://dx.doi.org/10.1017/s0317167100000433) [1017/s0317167100000433](http://dx.doi.org/10.1017/s0317167100000433).
- 84. Tanaka T, Tatsuno I, Uchida D, Moroo I, Morio H, Nakamura S, et al. Geranylgeranyl-pyrophosphate, an isoprenoid of mevalonate cascade, is a critical compound for rat primary cultured cortical neurons to protect the cell death induced by 3-hydroxy-3 methylglutaryl-CoA reductase inhibition. Journal of Neuroscience. 2000;20(8):2852–9.
- 85. Dimitroulakos J, Yeger H. HMG-CoA reductase mediates the biological effects of retinoic acid on human neuroblastoma cells: lovastatin specifically targets Pglycoprotein-expressing cells. Nat Med. 1996;2(3):326–33. [https://doi.org/10.1038/nm0396-](http://dx.doi.org/10.1038/nm0396-326) [326](http://dx.doi.org/10.1038/nm0396-326).
- 86. Jones KD, Couldwell WT, Hinton DR, Su Y, He S, Anker L, et al. Lovastatin induces growth inhibition and apoptosis in human malignant glioma cells. Biochem Biophys Res Commun. 1994;205(3):1681–7. [https://](http://dx.doi.org/10.1006/bbrc.1994.2861) [doi.org/10.1006/bbrc.1994.2861](http://dx.doi.org/10.1006/bbrc.1994.2861).
- 87. Miller AC, Samid D. Tumor resistance to oxidative stress: association with ras oncogene expression and reversal by lovastatin, an inhibitor of p21ras isoprenylation. Int J Cancer. 1995;60(2):249–54. [https://doi.org/10.1002/ijc.2910600220.](http://dx.doi.org/10.1002/ijc.2910600220)
- 88. Macaulay RJ, Wang W, Dimitroulakos J, Becker LE, Yeger H. Lovastatin-induced apoptosis of human medulloblastoma cell lines in vitro. J Neurooncol. 1999;42(1):1–11. [https://doi.org/10.1023/](http://dx.doi.org/10.1023/a:1006164406202) [a:1006164406202](http://dx.doi.org/10.1023/a:1006164406202).
- 89. Wang W, Macaulay RJ. Cell-cycle gene expression in lovastatin-induced medulloblastoma apoptosis. Can J Neurol Sci. 2003;30(4):349–57. [https://doi.org/10.](http://dx.doi.org/10.1017/s0317167100003061) [1017/s0317167100003061](http://dx.doi.org/10.1017/s0317167100003061).
- 90. Takwi AA, Li Y, Becker Buscaglia LE, Zhang J, Choudhury S, Park AK, et al. A statin-regulated microRNA represses human c-Myc expression and function. EMBO Mol Med. 2012;4(9):896–909. [https://doi.org/10.1002/emmm.201101045](http://dx.doi.org/10.1002/emmm.201101045).
- 91. Sheikholeslami K, Ali Sher A, Lockman S, Kroft D, Ganjibakhsh M, Nejati-Koshki K, et al. Simvastatin induces apoptosis in medulloblastoma brain tumor cells via mevalonate cascade prenylation substrates. Cancers (Basel). 2019;11(7). [https://doi.org/10.3390/](http://dx.doi.org/10.3390/cancers11070994) [cancers11070994.](http://dx.doi.org/10.3390/cancers11070994)
- 92. Gordon RE, Zhang L, Peri S, Kuo YM, Du F, Egleston BL, et al. Statins synergize with hedgehog pathway inhibitors for treatment of medulloblastoma. Clin Cancer Res. 2018;24(6):1375–88. [https://doi.org/10.](http://dx.doi.org/10.1158/1078-0432.CCR-17-2923) [1158/1078-0432.CCR-17-2923.](http://dx.doi.org/10.1158/1078-0432.CCR-17-2923)
- 93. Ling H, Luoma JT, Hilleman D. A review of currently available fenofibrate and fenofibric acid formulations. Cardiol Res. 2013;4(2):47–55. [https://doi.org/10.](http://dx.doi.org/10.4021/cr270w) [4021/cr270w](http://dx.doi.org/10.4021/cr270w).
- 94. Urbanska K, Pannizzo P, Grabacka M, Croul S, Del Valle L, Khalili K, et al. Activation of PPARalpha inhibits IGF-I-mediated growth and survival responses in medulloblastoma cell lines. Int J Cancer. 2008;123(5):1015–24. [https://doi.org/10.1002/ijc.](http://dx.doi.org/10.1002/ijc.23588) [23588](http://dx.doi.org/10.1002/ijc.23588).
- 95. Schwinger RH, Bohm M, Erdmann E. Effectiveness of cardiac glycosides in human myocardium with and without "downregulated" beta-adrenoceptors. J Cardiovasc Pharmacol. 1990;15(5):692–7. [https://doi.](http://dx.doi.org/10.1097/00005344-199005000-00002) [org/10.1097/00005344-199005000-00002.](http://dx.doi.org/10.1097/00005344-199005000-00002)
- 96. Johansson S, Lindholm P, Gullbo J, Larsson R, Bohlin L, Claeson P. Cytotoxicity of digitoxin and related cardiac glycosides in human tumor cells. Anticancer Drugs. 2001;12(5):475–83. [https://doi.org/10.1097/](http://dx.doi.org/10.1097/00001813-200106000-00009) [00001813-200106000-00009.](http://dx.doi.org/10.1097/00001813-200106000-00009)
- 97. Perne A, Muellner MK, Steinrueck M, Craig-Mueller N, Mayerhofer J, Schwarzinger I, et al. Cardiac glycosides induce cell death in human cells by inhibiting general protein synthesis. PLoS One. 2009;4(12):e8292. [https://doi.org/10.1371/journal.pone.0008292](http://dx.doi.org/10.1371/journal.pone.0008292).
- 98. Osman MH, Farrag E, Selim M, Osman MS, Hasanine A, Selim A. Cardiac glycosides use and the risk and mortality of cancer; systematic review and metaanalysis of observational studies. PLoS One. 2017;12(6):e0178611. [https://doi.org/10.1371/](http://dx.doi.org/10.1371/journal.pone.0178611) [journal.pone.0178611.](http://dx.doi.org/10.1371/journal.pone.0178611)
- 99. Kometiani P, Liu L, Askari A. Digitalis-induced signaling by Na+/K+-ATPase in human breast cancer cells. Mol Pharmacol. 2005;67(3):929–36. [https://doi.org/](http://dx.doi.org/10.1124/mol.104.007302) [10.1124/mol.104.007302.](http://dx.doi.org/10.1124/mol.104.007302)
- 100. Huang L, Garrett Injac S, Cui K, Braun F, Lin Q, Du Y, Zhang H, Kogiso M, Lindsay H, Zhao S, Baxter P, Adekunle A, Man T-K, Zhao H, Li X-N, Lau CC, Wong STC (2018) Systems biology-based drug

repositioning identifies digoxin as a potential therapy for groups 3 and 4 medulloblastoma. Science translational medicine 10 (464):eaat0150. doi:[https://doi.](http://dx.doi.org/10.1126/scitranslmed.aat0150) [org/10.1126/scitranslmed.aat0150](http://dx.doi.org/10.1126/scitranslmed.aat0150)

- 101. Wolle D, Lee SJ, Li Z, Litan A, Barwe SP, Langhans SA. Inhibition of epidermal growth factor signaling by the cardiac glycoside ouabain in medulloblastoma. Cancer Med. 2014;3(5):1146–58. [https://doi.org/10.](http://dx.doi.org/10.1002/cam4.314) [1002/cam4.314.](http://dx.doi.org/10.1002/cam4.314)
- 102. Schmidt WF, Huber KR, Ettinger RS, Neuberg RW. Antiproliferative effect of verapamil alone on brain tumor cells in vitro. Cancer Res. 1988;48(13):3617–21.
- 103. Ingram WJ, Crowther LM, Little EB, Freeman R, Harliwong I, Veleva D, et al. ABC transporter activity linked to radiation resistance and molecular subtype in pediatric medulloblastoma. Exp Hematol Oncol. 2013;2(1):26. [https://doi.org/10.1186/2162-3619-2-](http://dx.doi.org/10.1186/2162-3619-2-26) [26](http://dx.doi.org/10.1186/2162-3619-2-26).
- 104. Chen Y, Bai N, Bi J, Zhang J, Li X, Zhang L, et al. Propranolol induces apoptosis in endothelial cells by inhibiting AKt and ERK phosphorylation and MAPK signaling pathway. International Journal of Clinical and Experimental Medicine. 2017;10:13167–73.
- 105. Annabi B, Vaillancourt-Jean E, Weil AG, Beliveau R. Pharmacological targeting of beta-adrenergic receptor functions abrogates NF-kappaB signaling and MMP-9 secretion in medulloblastoma cells. Onco Targets Ther. 2010;3:219–26. [https://doi.org/10.2147/OTT.](http://dx.doi.org/10.2147/OTT.S14503) [S14503](http://dx.doi.org/10.2147/OTT.S14503).
- 106. Smout MJ, Kotze AC, McCarthy JS, Loukas A. A novel high throughput assay for anthelmintic drug screening and resistance diagnosis by real-time monitoring of parasite motility. PLoS Negl Trop Dis. 2010;4(11):e885. [https://doi.org/10.1371/journal.](http://dx.doi.org/10.1371/journal.pntd.0000885) [pntd.0000885.](http://dx.doi.org/10.1371/journal.pntd.0000885)
- 107. Thorne CA, Hanson AJ, Schneider J, Tahinci E, Orton D, Cselenyi CS, et al. Small-molecule inhibition of Wnt signaling through activation of casein kinase 1alpha. Nat Chem Biol. 2010;6(11):829–36. [https://](http://dx.doi.org/10.1038/nchembio.453) [doi.org/10.1038/nchembio.453](http://dx.doi.org/10.1038/nchembio.453).
- 108. Lum L, Yao S, Mozer B, Rovescalli A, Von Kessler D, Nirenberg M, et al. Identification of Hedgehog pathway components by RNAi in Drosophila cultured cells. Science. 2003;299(5615):2039–45. [https://doi.](http://dx.doi.org/10.1126/science.1081403) [org/10.1126/science.1081403.](http://dx.doi.org/10.1126/science.1081403)
- 109. Li B, Fei DL, Flaveny CA, Dahmane N, Baubet V, Wang Z, et al. Pyrvinium attenuates Hedgehog signaling downstream of smoothened. Cancer Res. 2014;74(17):4811–21. [https://doi.org/10.1158/](http://dx.doi.org/10.1158/0008-5472.CAN-14-0317) [0008-5472.CAN-14-0317](http://dx.doi.org/10.1158/0008-5472.CAN-14-0317).
- 110. De Witt M, Gamble A, Hanson D, Markowitz D, Powell C, Al Dimassi S, et al. Repurposing mebendazole as a replacement for vincristine for the treatment of brain tumors. Mol Med. 2017;23:50–6. [https://doi.org/10.2119/molmed.2017.00011.](http://dx.doi.org/10.2119/molmed.2017.00011)
- 111. Bai RY, Staedtke V, Rudin CM, Bunz F, Riggins GJ. Effective treatment of diverse medulloblastoma models with mebendazole and its impact on tumor

angiogenesis. Neuro Oncol. 2015;17(4):545–54. [https://doi.org/10.1093/neuonc/nou234.](http://dx.doi.org/10.1093/neuonc/nou234)

- 112. Dakshanamurthy S, Issa NT, Assefnia S, Seshasayee A, Peters OJ, Madhavan S, et al. Predicting new indications for approved drugs using a proteochemometric method. J Med Chem. 2012;55(15):6832–48. [https://](http://dx.doi.org/10.1021/jm300576q) [doi.org/10.1021/jm300576q.](http://dx.doi.org/10.1021/jm300576q)
- 113. Larsen AR, Bai RY, Chung JH, Borodovsky A, Rudin CM, Riggins GJ, et al. Repurposing the antihelmintic mebendazole as a hedgehog inhibitor. Mol Cancer Ther. 2015;14(1):3–13. [https://doi.org/10.1158/](http://dx.doi.org/10.1158/1535-7163.Mct-14-0755-t) [1535-7163.Mct-14-0755-t](http://dx.doi.org/10.1158/1535-7163.Mct-14-0755-t).
- 114. Bai RY, Staedtke V, Wanjiku T, Rudek MA, Joshi A, Gallia GL, et al. Brain penetration and efficacy of different mebendazole polymorphs in a mouse brain tumor model. Clin Cancer Res. 2015;21(15):3462– 70. [https://doi.org/10.1158/1078-0432.CCR-14-](http://dx.doi.org/10.1158/1078-0432.CCR-14-2681) [2681](http://dx.doi.org/10.1158/1078-0432.CCR-14-2681).
- 115. Koto KS, Lescault P, Brard L, Kim K, Singh RK, Bond J, et al. Antitumor activity of nifurtimox is enhanced with tetrathiomolybdate in medulloblastoma. Int J Oncol. 2011;38(5):1329–41. [https://doi.org/10.](http://dx.doi.org/10.3892/ijo.2011.971) [3892/ijo.2011.971](http://dx.doi.org/10.3892/ijo.2011.971).
- 116. Söderlund J, Erhardt S, Kast RE. Acyclovir inhibition of IDO to decrease Tregs as a glioblastoma treatment adjunct. J Neuroinflammation. 2010;7:44. [https://](http://dx.doi.org/10.1186/1742-2094-7-44) [doi.org/10.1186/1742-2094-7-44.](http://dx.doi.org/10.1186/1742-2094-7-44)
- 117. Zhang B, Wang X, Cai F, Chen W, Loesch U, Bitzer J, et al. Effects of salinomycin on human ovarian cancer cell line OV2008 are associated with modulating p38 MAPK. Tumour Biol. 2012;33(6):1855–62. [https://](http://dx.doi.org/10.1007/s13277-012-0445-9) [doi.org/10.1007/s13277-012-0445-9.](http://dx.doi.org/10.1007/s13277-012-0445-9)
- 118. Ketola K, Hilvo M, Hyotylainen T, Vuoristo A, Ruskeepaa AL, Oresic M, et al. Salinomycin inhibits prostate cancer growth and migration via induction of oxidative stress. Br J Cancer. 2012;106(1):99–106. [https://doi.org/10.1038/bjc.2011.530.](http://dx.doi.org/10.1038/bjc.2011.530)
- 119. Lu D, Choi MY, Yu J, Castro JE, Kipps TJ, Carson DA. Salinomycin inhibits Wnt signaling and selectively induces apoptosis in chronic lymphocytic leukemia cells. Proc Natl Acad Sci U S A. 2011;108(32):13253– 7. [https://doi.org/10.1073/pnas.1110431108](http://dx.doi.org/10.1073/pnas.1110431108).
- 120. Zhou S, Wang F, Zhang Y, Johnson MR, Qian S, Wu M, et al. Salinomycin suppresses PDGFRbeta, MYC, and Notch signaling in human medulloblastoma. Austin J Pharmacol Ther. 2014;2(3):1020.
- 121. Natarajan S, Li Y, Miller EE, Shih DJ, Taylor MD, Stearns TM, et al. Notch1-induced brain tumor models the sonic hedgehog subgroup of human medulloblastoma. Cancer Res. 2013;73(17):5381–90. [https://doi.org/10.1158/0008-5472.CAN-13-0033](http://dx.doi.org/10.1158/0008-5472.CAN-13-0033).
- 122. Pace JR, Jog R, Burgess DJ, Hadden MK. Formulation and evaluation of itraconazole liposomes for hedgehog pathway inhibition. J Liposome Res. 2019:1-7. [https://doi.org/10.1080/08982104.2019.1668011](http://dx.doi.org/10.1080/08982104.2019.1668011).
- 123. Shaimerdenova M, Karapina O, Mektepbayeva D, Alibek K, Akilbekova D. The effects of antiviral treatment on breast cancer cell line. Infect Agent Cancer.

2017;12:18. [https://doi.org/10.1186/s13027-017-](http://dx.doi.org/10.1186/s13027-017-0128-7) [0128-7.](http://dx.doi.org/10.1186/s13027-017-0128-7)

- 124. Baryawno N, Rahbar A, Wolmer-Solberg N, Taher C, Odeberg J, Darabi A, et al. Detection of human cytomegalovirus in medulloblastomas reveals a potential therapeutic target. J Clin Invest. 2011;121(10):4043– 55. [https://doi.org/10.1172/JCI57147.](http://dx.doi.org/10.1172/JCI57147)
- 125. Wang LH, Chittick GE, McDowell JA. Single-dose pharmacokinetics and safety of abacavir (1592U89), zidovudine, and lamivudine administered alone and in combination in adults with human immunodeficiency virus infection. Antimicrob Agents Chemother. 1999;43(7):1708–15.
- 126. Yegorov YE, Chernov DN, Akimov SS, Bolsheva NL, Krayevsky AA, Zelenin AV. Reverse transcriptase inhibitors suppress telomerase function and induce senescence-like processes in cultured mouse fibroblasts. FEBS Lett. 1996;389(2):115–8. [https://doi.org/](http://dx.doi.org/10.1016/0014-5793(96)00533-9) [10.1016/0014-5793\(96\)00533-9](http://dx.doi.org/10.1016/0014-5793(96)00533-9).
- 127. Rossi A, Russo G, Puca A, La Montagna R, Caputo M, Mattioli E, et al. The antiretroviral nucleoside analogue Abacavir reduces cell growth and promotes differentiation of human medulloblastoma cells. Int J Cancer. 2009;125(1):235–43. [https://doi.org/10.](http://dx.doi.org/10.1002/ijc.24331) [1002/ijc.24331](http://dx.doi.org/10.1002/ijc.24331).
- 128. Amano H, Hayashi I, Endo H, Kitasato H, Yamashina S, Maruyama T, et al. Host prostaglandin E(2)-EP3 signaling regulates tumor-associated angiogenesis and tumor growth. J Exp Med. 2003;197(2):221–32. [https://doi.org/10.1084/jem.20021408.](http://dx.doi.org/10.1084/jem.20021408)
- 129. Leidgens V, Seliger C, Jachnik B, Welz T, Leukel P, Vollmann-Zwerenz A, et al. Ibuprofen and diclofenac restrict migration and proliferation of human glioma cells by distinct molecular mechanisms. PLoS One. 2015;10(10):e0140613. [https://doi.org/10.1371/](http://dx.doi.org/10.1371/journal.pone.0140613) [journal.pone.0140613](http://dx.doi.org/10.1371/journal.pone.0140613).
- 130. Shi J, Leng W, Zhao L, Xu C, Wang J, Chen X, Wang Y, Peng X (2017) Nonsteroidal anti-inflammatory drugs using and risk of head and neck cancer: a doseresponse meta analysis of prospective cohort studies. Oncotarget 8 (58):99066-99074. doi:10.18632/ oncotarget.21524
- 131. Harris RE, Chlebowski RT, Jackson RD, Frid DJ, Ascenseo JL, Anderson G, et al. Breast cancer and nonsteroidal anti-inflammatory drugs: prospective results from the Women's Health Initiative. Cancer Res. 2003;63(18):6096–101.
- 132. Trabert B, Ness RB, Lo-Ciganic WH, Murphy MA, Goode EL, Poole EM, Brinton LA, Webb PM, Nagle CM, Jordan SJ, Risch HA, Rossing MA, Doherty JA, Goodman MT, Lurie G, Kjær SK, Hogdall E, Jensen A, Cramer DW, Terry KL, Vitonis A, Bandera EV, Olson S, King MG, Chandran U, Anton-Culver H, Ziogas A, Menon U, Gayther SA, Ramus SJ, Gentry-Maharaj A, Wu AH, Pearce CL, Pike MC, Berchuck A, Schildkraut JM, Wentzensen N (2014) Aspirin, nonaspirin nonsteroidal anti-inflammatory drug, and acetaminophen use and risk of invasive epithelial ovarian cancer: a pooled analysis in the Ovarian Cancer

Association Consortium. J Natl Cancer Inst 106 (2):djt431. doi:[https://doi.org/10.1093/jnci/djt431](http://dx.doi.org/10.1093/jnci/djt431)

- 133. Vidal AC, Howard LE, Moreira DM, Castro-Santamaria R, Andriole GL, Freedland SJ. Aspirin, NSAIDs, and risk of prostate cancer: results from the REDUCE study. Clin Cancer Res. 2015;21(4):756–62. [https://doi.org/10.1158/1078-0432.CCR-14-2235](http://dx.doi.org/10.1158/1078-0432.CCR-14-2235).
- 134. Friis S, Riis AH, Erichsen R, Baron JA, Sørensen HT. Low-dose aspirin or nonsteroidal anti-inflammatory drug use and colorectal cancer risk: a populationbased, case-control study. Ann Intern Med. 2015;163(5):347–55. [https://doi.org/10.7326/m15-](http://dx.doi.org/10.7326/m15-0039) [0039](http://dx.doi.org/10.7326/m15-0039).
- 135. Verschuur A, Heng-Maillard MA, Dory-Lautrec P, Truillet R, Jouve E, Chastagner P, et al. Metronomic four-drug regimen has anti-tumor activity in pediatric low-grade glioma; the results of a phase II clinical trial. Front Pharmacol. 2018;9:00950. [https://doi.](http://dx.doi.org/10.3389/fphar.2018.00950) [org/10.3389/fphar.2018.00950](http://dx.doi.org/10.3389/fphar.2018.00950).
- 136. Yang MY, Lee HT, Chen CM, Shen CC, Ma HI. Celecoxib suppresses the phosphorylation of STAT3 protein and can enhance the radiosensitivity of medulloblastoma-derived cancer stem-like cells. Int J Mol Sci. 2014;15(6):11013–29. [https://doi.org/10.](http://dx.doi.org/10.3390/ijms150611013) [3390/ijms150611013.](http://dx.doi.org/10.3390/ijms150611013)
- 137. Chen KH, Hsu CC, Song WS, Huang CS, Tsai CC, Kuo CD, et al. Celecoxib enhances radiosensitivity in medulloblastoma-derived CD133-positive cells. Childs Nerv Syst. 2010;26(11):1605–12. [https://doi.](http://dx.doi.org/10.1007/s00381-010-1190-2) [org/10.1007/s00381-010-1190-2.](http://dx.doi.org/10.1007/s00381-010-1190-2)
- 138. Baryawno N, Sveinbjornsson B, Eksborg S, Orrego A, Segerstrom L, Oqvist CO, et al. Tumor-growthpromoting cyclooxygenase-2 prostaglandin E2 pathway provides medulloblastoma therapeutic targets. Neuro Oncol. 2008;10(5):661–74. [https://doi.org/](http://dx.doi.org/10.1215/15228517-2008-035) [10.1215/15228517-2008-035](http://dx.doi.org/10.1215/15228517-2008-035).
- 139. Eslin D, Lee C, Sankpal UT, Maliakal P, Sutphin RM, Abraham L, et al. Anticancer activity of tolfenamic acid in medulloblastoma: a preclinical study. Tumour Biol. 2013;34(5):2781–9. [https://doi.org/10.1007/](http://dx.doi.org/10.1007/s13277-013-0836-6) [s13277-013-0836-6](http://dx.doi.org/10.1007/s13277-013-0836-6).
- 140. Abdelrahim M, Baker CH, Abbruzzese JL, Safe S. Tolfenamic acid and pancreatic cancer growth, angiogenesis, and Sp protein degradation. J Natl Cancer Inst. 2006;98(12):855–68. [https://doi.org/10.1093/](http://dx.doi.org/10.1093/jnci/djj232) [jnci/djj232.](http://dx.doi.org/10.1093/jnci/djj232)
- 141. Shelake S, Sankpal UT, Paul Bowman W, Wise M, Ray A, Basha R. Targeting specificity protein 1 transcription factor and survivin using tolfenamic acid for inhibiting Ewing sarcoma cell growth. Invest New Drugs. 2017;35(2):158–65. [https://doi.org/10.1007/](http://dx.doi.org/10.1007/s10637-016-0417-9) [s10637-016-0417-9](http://dx.doi.org/10.1007/s10637-016-0417-9).
- 142. King JG Jr, Khalili K. Inhibition of human brain tumor cell growth by the anti-inflammatory drug, flurbiprofen. Oncogene. 2001;20(47):6864–70. [https://doi.org/10.1038/sj.onc.1204907](http://dx.doi.org/10.1038/sj.onc.1204907).
- 143. Ng CG, Boks MP, Smeets HM, Zainal NZ, de Wit NJ. Prescription patterns for psychotropic drugs in cancer patients; a large population study in the Netherlands.

Psychooncology. 2013;22(4):762–7. [https://doi.org/](http://dx.doi.org/10.1002/pon.3056) [10.1002/pon.3056.](http://dx.doi.org/10.1002/pon.3056)

- 144. Mortensen PB. The incidence of cancer in schizophrenic patients. J Epidemiol Community Health. 1989;43(1):43–7. [https://doi.org/10.1136/jech.43.1.](http://dx.doi.org/10.1136/jech.43.1.43) [43](http://dx.doi.org/10.1136/jech.43.1.43).
- 145. Chen JJ, Cai N, Chen GZ, Jia CC, Qiu DB, Du C, Liu W, Yang Y, Long ZJ, Zhang Q (2017) The neuroleptic drug pimozide inhibits stem-like cell maintenance and tumorigenicity in hepatocellular carcinoma. Oncotarget 8 (11):17593-17609. doi:10.18632/ oncotarget.4307
- 146. Ludwig J, Weseloh R, Karschin C, Liu Q, Netzer R, Engeland B, et al. Cloning and functional expression of rat eag2, a new member of the ether-à-go-go family of potassium channels and comparison of its distribution with that of eag1. Mol Cell Neurosci. 2000;16(1):59–70. [https://doi.org/10.1006/mcne.](http://dx.doi.org/10.1006/mcne.2000.0851) [2000.0851.](http://dx.doi.org/10.1006/mcne.2000.0851)
- 147. Huang X, Dubuc AM, Hashizume R, Berg J, He Y, Wang J, et al. Voltage-gated potassium channel EAG2 controls mitotic entry and tumor growth in medulloblastoma via regulating cell volume dynamics. Genes & development. 2012;26(16):1780–96. [https://doi.org/10.1101/gad.193789.112](http://dx.doi.org/10.1101/gad.193789.112).
- 148. Huang X, He Y, Dubuc AM, Hashizume R, Zhang W, Reimand J, et al. EAG2 potassium channel with evolutionarily conserved function as a brain tumor target. Nat Neurosci. 2015;18(9):1236–46. [https://doi.org/](http://dx.doi.org/10.1038/nn.4088) [10.1038/nn.4088](http://dx.doi.org/10.1038/nn.4088).
- 149. Mascaro-Cordeiro B, Oliveira ID, Tesser-Gamba F, Pavon LF, Saba-Silva N, Cavalheiro S, et al. Valproic acid treatment response in vitro is determined by TP53 status in medulloblastoma. Childs Nerv Syst. 2018;34(8):1497–509. [https://doi.org/10.1007/](http://dx.doi.org/10.1007/s00381-018-3817-7) [s00381-018-3817-7](http://dx.doi.org/10.1007/s00381-018-3817-7).
- 150. Li XN, Shu Q, Su JM, Perlaky L, Blaney SM, Lau CC. Valproic acid induces growth arrest, apoptosis, and senescence in medulloblastomas by increasing histone hyperacetylation and regulating expression of p21Cip1, CDK4, and CMYC. Mol Cancer Ther. 2005;4(12):1912–22. [https://doi.org/10.1158/1535-](http://dx.doi.org/10.1158/1535-7163.MCT-05-0184) [7163.MCT-05-0184](http://dx.doi.org/10.1158/1535-7163.MCT-05-0184).
- 151. Sassi RB, Nicoletti M, Brambilla P, Mallinger AG, Frank E, Kupfer DJ, et al. Increased gray matter volume in lithium-treated bipolar disorder patients. Neurosci Lett. 2002;329(2):243–5. [https://doi.org/](http://dx.doi.org/10.1016/s0304-3940(02)00615-8) [10.1016/s0304-3940\(02\)00615-8.](http://dx.doi.org/10.1016/s0304-3940(02)00615-8)
- 152. Moore GJ, Bebchuk JM, Hasanat K, Chen G, Seraji-Bozorgzad N, Wilds IB, et al. Lithium increases Nacetyl-aspartate in the human brain: in vivo evidence in support of bcl-2's neurotrophic effects? Biol Psychiatry. 2000;48(1):1–8. [https://doi.org/10.1016/](http://dx.doi.org/10.1016/s0006-3223(00)00252-3) [s0006-3223\(00\)00252-3](http://dx.doi.org/10.1016/s0006-3223(00)00252-3).
- 153. Mao CD, Hoang P, DiCorleto PE. Lithium inhibits cell cycle progression and induces stabilization of p53 in bovine aortic endothelial cells. J Biol Chem. 2001;276(28):26180–8. [https://doi.org/10.1074/jbc.](http://dx.doi.org/10.1074/jbc.M101188200) [M101188200](http://dx.doi.org/10.1074/jbc.M101188200).
- 154. Ronchi A, Salaroli R, Rivetti S, Della Bella E, Di Tomaso T, Voltattorni M, et al. Lithium induces mortality in medulloblastoma cell lines. Int J Oncol. 2010;37(3):745–52. [https://doi.org/10.3892/ijo_](http://dx.doi.org/10.3892/ijo_00000724) [00000724](http://dx.doi.org/10.3892/ijo_00000724).
- 155. Zhukova N, Ramaswamy V, Remke M, Martin DC, Castelo-Branco P, Zhang CH, et al. WNT activation by lithium abrogates TP53 mutation associated radiation resistance in medulloblastoma. Acta Neuropathol Commun. 2014;2:174. [https://doi.org/](http://dx.doi.org/10.1186/s40478-014-0174-y) [10.1186/s40478-014-0174-y.](http://dx.doi.org/10.1186/s40478-014-0174-y)
- 156. Kurita JI, Hirao Y, Nakano H, Fukunishi Y, Nishimura Y. Sertraline, chlorprothixene, and chlorpromazine characteristically interact with the REST-binding site of the corepressor mSin3, showing medulloblastoma cell growth inhibitory activities. Sci Rep. 2018;8(1):13763. [https://doi.org/10.1038/s41598-](http://dx.doi.org/10.1038/s41598-018-31852-1) [018-31852-1.](http://dx.doi.org/10.1038/s41598-018-31852-1)
- 157. Spiller SE, Ditzler SH, Pullar BJ, Olson JM. Response of preclinical medulloblastoma models to combination therapy with 13-cis retinoic acid and suberoylanilide hydroxamic acid (SAHA). J Neurooncol. 2008;87(2):133–41. [https://doi.org/10.](http://dx.doi.org/10.1007/s11060-007-9505-1) [1007/s11060-007-9505-1.](http://dx.doi.org/10.1007/s11060-007-9505-1)
- 158. Hallahan AR, Pritchard JI, Chandraratna RA, Ellenbogen RG, Geyer JR, Overland RP, et al. BMP-2 mediates retinoid-induced apoptosis in medulloblastoma cells through a paracrine effect. Nat Med. 2003;9(8):1033–8. [https://doi.org/10.1038/nm904](http://dx.doi.org/10.1038/nm904).
- 159. Gumireddy K, Sutton LN, Phillips PC, Reddy CD. Alltrans-retinoic acid-induced apoptosis in human medulloblastoma: activation of caspase-3/poly(ADP-ribose) polymerase 1 pathway. Clin Cancer Res. 2003;9(11):4052–9.
- 160. Mancini D, Pinney S, Burkhoff D, LaManca J, Itescu S, Burke E, et al. Use of rapamycin slows progression of cardiac transplantation vasculopathy. Circulation. 2003;108(1):48–53. [https://doi.org/10.1161/01.Cir.](http://dx.doi.org/10.1161/01.Cir.0000070421.38604.2b) [0000070421.38604.2b.](http://dx.doi.org/10.1161/01.Cir.0000070421.38604.2b)
- 161. Bahmad HF, Mouhieddine TH, Chalhoub RM, Assi S, Araji T, Chamaa F, Itani MM, Nokkari A, Kobeissy F, Daoud G, Abou-Kheir W (2018) The Akt/mTOR pathway in cancer stem/progenitor cells is a potential therapeutic target for glioblastoma and neuroblastoma. Oncotarget 9 (71):33549-33561. doi:10.18632/ oncotarget.26088
- 162. Mohan AL, Friedman MD, Ormond DR, Tobias M, Murali R, Jhanwar-Uniyal M. PI3K/mTOR signaling pathways in medulloblastoma. Anticancer Res. 2012;32(8):3141–6.
- 163. Garner EF, Williams AP, Stafman LL, Aye JM, Mroczek-Musulman E, Moore BP, et al. FTY720 decreases tumorigenesis in group 3 medulloblastoma patient-derived xenografts. Sci Rep. 2018;8(1):6913. [https://doi.org/10.1038/s41598-018-25263-5.](http://dx.doi.org/10.1038/s41598-018-25263-5)
- 164. Mancuso M, Leonardi S, Ceccarelli M, Pasquali E, De Stefano I, Prisco MG, et al. Protective role of 17 βestradiol on medulloblastoma development in Patched 1 heterozygous mice. Int J Cancer.

2010;127(12):2749–57. [https://doi.org/10.1002/ijc.](http://dx.doi.org/10.1002/ijc.25293) [25293](http://dx.doi.org/10.1002/ijc.25293).

- 165. Belcher SM, Ma X, Le HH. Blockade of estrogen receptor signaling inhibits growth and migration of medulloblastoma. Endocrinology. 2009;150(3):1112–21. [https://doi.org/10.1210/en.](http://dx.doi.org/10.1210/en.2008-1363) [2008-1363](http://dx.doi.org/10.1210/en.2008-1363).
- 166. Ciucci A, Meco D, De Stefano I, Travaglia D, Zannoni GF, Scambia G, et al. Gender effect in experimental models of human medulloblastoma: does the estrogen receptor β signaling play a role? PloS one. 2014;9(7):e101623. [https://doi.org/10.1371/journal.](http://dx.doi.org/10.1371/journal.pone.0101623) [pone.0101623](http://dx.doi.org/10.1371/journal.pone.0101623).
- 167. Spiller SE, Logsdon NJ, Deckard LA, Sontheimer H. Inhibition of nuclear factor kappa-B signaling reduces growth in medulloblastoma in vivo. BMC Cancer. 2011;11:136. [https://doi.org/10.1186/1471-2407-](http://dx.doi.org/10.1186/1471-2407-11-136) [11-136.](http://dx.doi.org/10.1186/1471-2407-11-136)
- 168. Di Magno L, Manni S, Di Pastena F, Coni S, Macone A, Cairoli S, Sambucci M, Infante P, Moretti M, Petroni M, Nicoletti C, Capalbo C, De Smaele E, Di Marcotullio L, Giannini G, Battistini L, Goffredo BM, Iorio E, Agostinelli E, Maroder M, Canettieri G (2020) Phenformin inhibits hedgehog-dependent tumor growth through a complex I-independent redox/ corepressor module. Cell Rep 30 (6):1735- 1752.e1737. doi[:https://doi.org/10.1016/j.celrep.](http://dx.doi.org/10.1016/j.celrep.2020.01.024) [2020.01.024](http://dx.doi.org/10.1016/j.celrep.2020.01.024)
- 169. Mouhieddine TH, Nokkari A, Itani MM, Chamaa F, Bahmad H, Monzer A, et al. Metformin and Ara-a effectively suppress brain cancer by targeting cancer stem/progenitor cells. Frontiers in neuroscience. 2015;9:442. [https://doi.org/10.3389/fnins.2015.](http://dx.doi.org/10.3389/fnins.2015.00442) [00442](http://dx.doi.org/10.3389/fnins.2015.00442).
- 170. Skinner MD, Lahmek P, Pham H, Aubin HJ. Disulfiram efficacy in the treatment of alcohol dependence: a meta-analysis. PLoS One. 2014;9(2):e87366. [https://doi.org/10.1371/journal.pone.0087366](http://dx.doi.org/10.1371/journal.pone.0087366).
- 171. Zha J, Chen F, Dong H, Shi P, Yao Y, Zhang Y, et al. Disulfiram targeting lymphoid malignant cell lines via ROS-JNK activation as well as Nrf2 and NF-kB pathway inhibition. J Transl Med. 2014;12:163. [https://doi.org/10.1186/1479-5876-12-163.](http://dx.doi.org/10.1186/1479-5876-12-163)
- 172. Yip NC, Fombon IS, Liu P, Brown S, Kannappan V, Armesilla AL, et al. Disulfiram modulated ROS– MAPK and NFκB pathways and targeted breast cancer cells with cancer stem cell-like properties. British Journal of Cancer. 2011;104(10):1564–74. [https://](http://dx.doi.org/10.1038/bjc.2011.126) [doi.org/10.1038/bjc.2011.126.](http://dx.doi.org/10.1038/bjc.2011.126)
- 173. Chen D, Cui QC, Yang H, Dou QP. Disulfiram, a clinically used anti-alcoholism drug and copperbinding agent, induces apoptotic cell death in breast cancer cultures and xenografts via inhibition of the proteasome activity. Cancer Res. 2006;66(21):10425–33. [https://doi.org/10.1158/](http://dx.doi.org/10.1158/0008-5472.CAN-06-2126) [0008-5472.CAN-06-2126](http://dx.doi.org/10.1158/0008-5472.CAN-06-2126).
- 174. Madala HR, Punganuru SR, Ali-Osman F, Zhang R, Srivenugopal KS (2017) Brain- and brain tumorpenetrating disulfiram nanoparticles: sequence of

cytotoxic events and efficacy in human glioma cell lines and intracranial xenografts. Oncotarget 9 (3):3459-3482. doi:10.18632/oncotarget.23320

175. Zhang Z, Zhou L, Xie N, Nice EC, Zhang T, Cui Y, et al. Overcoming cancer therapeutic bottleneck by drug repurposing. Signal Transduction and Targeted Therapy. 2020;5(1):113. [https://doi.org/10.1038/s41392-](http://dx.doi.org/10.1038/s41392-020-00213-8) [020-00213-8.](http://dx.doi.org/10.1038/s41392-020-00213-8)

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