NEURO-ONCOLOGY (Y UMEMURA, SECTION EDITOR)

Medulloblastoma: an Old Diagnosis with New Promises

Luca Szalontay¹ · Yasmin Khakoo^{1,2}

© Springer Science+Business Media, LLC, part of Springer Nature 2020

Abstract



Purpose of Review Molecular subtyping in medulloblastoma (MB) has diagnostic and prognostic values which impact therapy. This paper provides guidance for the clinician caring for pediatric and adult patients with medulloblastoma in the modern era. **Recent Findings** Medulloblastoma comprises four molecularly distinct subgroups: wingless activated (WNT), sonic hedgehog activated (SHH), group 3, and group 4. Risk stratification before and after the discovery of molecular subgroups aims at minimizing toxicity by reducing radiation and chemotherapy doses in low-risk patients while maintaining favorable overall survival (OS). The mainstay of newly diagnosed medulloblastoma treatment is surgery, radiation therapy, and chemotherapy, except for children under 6 years of age, where high-dose chemotherapy with autologous stem cell rescue is used to avoid or delay radiotherapy, preventing neurocognitive sequelae. Management of recurrent/refractory medulloblastoma remains a challenge with immunotherapy and small-molecule inhibitors forming the backbone of novel strategies.

Summary Recent innovations in medulloblastoma research allow us to better understand pathogenesis and molecular characteristics resulting in advanced risk stratification models, new therapeutic approaches, and overall improved survival and quality of life.

Keywords Medulloblastoma \cdot Molecular subtyping \cdot Genomics \cdot Neurosurgery \cdot Radiation therapy \cdot Chemotherapy \cdot Targeted therapy \cdot Clinical trials

Introduction

In 1925, Harvey Cushing, MD and Percival Bailey, MD coined the term medulloblastoma after reviewing a series of cases of central nervous system (CNS) tumors and identifying a subtype of posterior fossa tumors seen in younger children [1]. They also recognized that more extensive tumor excision correlated with longer survival. In the 1950s, the introduction of craniospinal irradiation (CSI) with a posterior fossa boost further improved outcome, but at a price: increased risk of neurocognitive impairment, endocrine disfunction, and secondary malignancies were commonly seen [2]. In the 1970s, adjuvant chemotherapy was introduced in an effort to improve

This article is part of the Topical Collection on Neuro-oncology

Yasmin Khakoo khakooy@mskcc.org survival, decrease radiation dose, and subsequently reduce deleterious late effects [3]. While surgery, external beam radiation therapy (in patients over 6 years of age), and chemotherapy continue to be the mainstay of medulloblastoma treatment, the discovery of molecular subtyping in 2000, led to the identification of four distinctive subgroups of medulloblastoma incorporated into the 2016 WHO classification of CNS tumors [4]. The advent of newer technologies to further subclassify tumors has set the stage for novel therapeutic approaches.

Epidemiology

CNS tumors are the most common form of solid tumors in children and adolescents [5] and the leading cause of cancer-related death in this age group [6]. Embryonal tumors account for 10%, with medulloblastoma being the most common, representing 63%. In children age 0-19 years, the incidence is 0.39 per 100,000 population, and it decreases with age [7]. As with most childhood cancers, medulloblastoma is more common in

¹ Department of Pediatrics, Memorial Sloan Kettering Cancer Center, 1275 York Ave., New York, NY 10065, USA

² Department of Pediatrics, Weill Cornell Medical College, New York, NY, USA

males [8], though there may be variability depending upon MB subtypes as discussed below [9•]. While much is known about the MB biology including molecular characteristics and genetic syndromes, to date, the role of environmental factors is unknown.

Genetic Syndromes

The association between certain hereditary cancer predisposition syndromes and medulloblastoma is well recognized. Table 1 summarizes the most important familial cancer predisposition syndromes associated with medulloblastoma. In 2018, Waszak et al. reported that germline mutations accounted for 6% of medulloblastoma diagnoses [10••]. The investigation also found that the risk of medulloblastoma is highest in carriers of SUFU and TP53 mutations and BRCA2 and PALB2 biallelic mutations [10••]. In over half of the cases, where an underlying genetic predisposition syndrome was identified after germline testing had been performed, medical and family histories were unrevealing emphasizing the need of new guidelines for genetic counseling and testing. Understanding the molecular characteristics of these tumors played a significant role in the identification of the medulloblastoma subgroups.

Subgroups—Signal Pathways

Pomeroy et al. demonstrated in 2002 that medulloblastomas are molecularly distinct from other brain tumors [11]. Further investigation by Taylor et al. defined four distinct molecular entities: wingless activated (WNT 10%), sonic hedgehog activated (SHH 30%), group 3 (20–25%), and group 4 (35–40%) [12, 13]. It has been shown that the molecular subgroup is defined at the time of tumor initiation and is not affected by genetic evolution, therapy, or other factors [14]. Figure 1 summarizes the characteristics of each MB subgroup.

WNT Subgroup

Ten percent of all medulloblastomas belong to the WNT subgroup and occur in children over 3 years of age with a 1:1 M:F ratio. If diagnosed before age 16 years, prognosis is excellent [15, 16].

The Wnt/ β -catenin pathway plays a central role during embryonal development, regulating stem cell pluripotency and driving cell growth and proliferation. Nearly 90% of WNT medulloblastomas harbor mutations in exon 3 of the *CTNNB1* gene, resulting in reduced cytoplasmic degradation and nuclear accumulation of the transcription factor coactivator β -catenin [17••]. Other commonly mutated genes in the WNT subgroup include *DDX3X*, *SMARCA4*, *TP53*,

Mutated gene	Cancer predisposition syndrome	Clinical features	Role of mutated gene	Subgroup association
APC	Turcot syndrome	Multiple adenomatous colon polyps, increased risk of colorectal cancer and CNS tumors	Negative regulator that controls β-catenin concentrations	WNT, rarely SHH
PTCH1 SUFU	Gorlin syndrome	Developmental abnormalities, bone cysts, increased risk of basal cell carcinoma and medulloblastoma	Receptor for sonic hedgehog Negative regulator in the hedgehog/smoothened signaling pathway	SHH
TP53	Li Fraumeni syndrome	Multiple cancer types and primary sites (breast, sarcomas, brain tumors, adrenocortical carcinoma) at an early onset	Tumor suppressor protein regulates genes involved in cell cycle arrest, apoptosis, senescence, DNA repair, and changes in metabolism	SHH
BRCA2/FANCD1	Fanconi anemia	Developmental abnormalities, bone marrow failure, predisposition to medulloblastoma	Double-strand DNA-break repair, vital in homologous recombination	SHH if compound heterozygous, groups 3 and 4 if heterozygous mutation
PALB2			Partner and localizer of BRCA2	SHH, rarely group 3 and group 4
CREBBP	Rubinstein Taybi syndrome	Microcephaly, growth deficiency, dysmorphic features, intellectual disability, increased risk of brain tumors	CREB binding protein—co-activator of transcription factors	

Table 1 Cancer predisposition syndromes in medulloblastoma

Subgroup		WNT		SHH			Group 3			Group 4			
S	ubtype	WNT α	WNT β	SHH α	SHH β	SHH y	SHH δ	Group 3a	Group 3β	Group 3y	Group 4a	Group 4β	Group 4y
Subtype proportion		αδ			3β 3α 3γ			4β 4α 4γ					
Subtype relationship		α β Υ δ			α β γ			β α γ					
ata	Age	††	∱ ∯	††	÷	÷	Ń	÷ †	† †	÷ †	††	††	††
Clinical d	Metastases	8.6%	21.4%	20%	33%	8.9%	9.4%	43.4%	20%	39.4%	40%	40.7%	38.7%
	Survival at 5 years	97%	100%	69.8%	67.3%	88%	88.5%	66.2%	55.8%	41.9%	66.8%	75.4%	82.5%
Copy number	Broad	6		9q [°] , 10q [°] , 17p [°]		Balanced genome		7 ⁺ , 8 ⁻ , 10 ⁻ , 11 ⁻ , i17q		8 [†] , i17q	7q ⁺ , 8p [−] , i17q	i17q	7q ⁺ , 8p ⁻ , i17q (less)
	Focal			MYCN amp, GLI2 amp, YAP1 amp	PTEN loss		10q22 ⁻ , 11q23.3 ⁻		<i>OTX2</i> gain, <i>DDX31</i> loss	MYC amp	MYCN amp, CDK6 amp	SNCAIP dup	CDK6 amp
0	ther events			<i>TP53</i> mutations			TERT promoter mutations		High <i>GFI1/1B</i> expression				
Tumor location/ enhancement patterns Cerebellopontine angle		Cerebellar hemisphere			Midline, ill-define margins		Midline, no enhancement						
Origin Cells in the lower rhombic lip		Cerebellar granule neuron progenitors (CGNPs)			Uncertain		Uncertain						
Histology Classic			Desmoplastic nodular MBEN - infant LC/A - TP53 mutant			Classic LC/A - infant		Classic					
Risk of CMS 21%		7%			31%		35%						

Age (years): * 0-3 * >3-10 * >10-17 * >17

Fig. 1 Summary of medulloblastoma subgroups adapted from Cavalli et al. Intertumoral Heterogeneity within Medulloblastoma Subgroups. Cell 2017;31:75 [12]

and *CSNK2B*, which also contribute to tumorigenesis. Gibson et al. developed a transgenic mouse model of WNT MB using constitutively active Ctnnb1 and absent p53 expressions [18]. The addition of constitutively active Pik3ca increased tumor development from 20 to 100% and reduced the latency of tumor onset in the mice from 6 to 3 months of age [19]. The 10-15% of WNT MB lacking *CTNNB1* mutations often harbor pathogenic variants of the *APC* gene [10••]. Another hallmark of WNT-driven MB is monosomy 6, found in about 85% of the cases. A small-subset WNT medulloblastomas acquire subclonal genetic alteration which secondarily activate the SHH pathway [20]. The prognostic and therapeutic significance of this finding remains unclear.

SHH Subgroup

The sonic hedgehog pathway regulates cellular differentiation, organ formation, and post-embryonic tissue regeneration and repair in multicellular organisms [21]. The key components of the pathway include the transmembrane receptor Patched1

(PTCH1), the signal transducer Smoothened (SMO), GLI transcription factors, and Suppressor of fused homolog (SUFU), which normally suppresses GLI [22]. SHH binds PTCH1, which releases SMO resulting in the dissociation of the GLI transcription factors from SUFU. The unbound GLI translocates to the nucleus turning on gene transcription. [23].

Disruption of this pathway may result in SHH-driven medulloblastoma as follows: loss of function mutations or deletions in *PTCH1* (43%) and *SUFU* (10%), activation of mutations in *SMO* (9%), or amplifications of *GL11* or *GL12* (9%) [17••]. Wetmore et al. demonstrated that germline deletion of *PTCH1* generated MB in 15–20% of mice, and the additional deletion of *TP53* produced tumors in 100% [24].

More recent studies identified four SHH MB subtypes, each with a specific phenotype and different prognosis [12]. For example, SHH α with germline or somatic *TP53* mutations occurs in older children and adolescents and predicts poor outcome. Both SSH β and SSH γ tumors develop primarily in infants, with SSH γ conferring a better prognosis [25]. Finally, SSH δ , commonly seen in adults, has a prognosis somewhere in the middle and is almost always associated with somatic *TERT* promoter mutations [26].

Group 3 Subgroup

Group 3 medulloblastoma, seen primarily in infants and older children with a male predominance, has a 40-65% OS depending upon the subtype. These patients may present with disseminated disease, which have a guarded prognosis [13], especially in the setting of MYC amplification, representing 15–20% of the cases [27]. Other cytogenetic features include the presence of isochromosome 17q and gain of chromosome 8q [12]. Gene-level mutations are rare, primarily occurring in SMARCA4, KBTBD4, CTDNEP1, and KMT2D [17••]. Enhancer hijacking is a well described epigenetic phenomenon, whereby regulatory elements are translocated close to coding genomic regions, resulting in gene overexpression and tumor formation. Northcott et al. observed that enhancer hijacking occurs in one-third of group 3 MB (resulting in GFI1/GFI1B oncogene activation) and in 17% of group 4 MB as described next [28, 29].

Group 4 Subgroup

Group 4 is most commonly driven by the enhancer-hijackingmediated overexpression of *PRDM6*, strongly associated with focal tandem duplication of *SNCAIP*. Mutations in histonemodifying genes *KDM6A*, *ZMYM3*, *KMT2C*, and *KBTBD4* have also been described [17••]. Cytogenetic aberrations are frequent in this subgroup, with gain of chromosome 7 (40%) or 17q (>80%), and deletion of chromosome 8 (40%), 11 (> 30%) or 17p (>75%), and the most common, isochromosome 17q (80%). High-level amplifications of *MYCN* and *CDK6* are also observed [27].

Its prognosis is intermediate: patients with metastasis have a higher risk of relapse, while those tumors with loss of chromosome 11 or gain of chromosome 17 are associated with good outcome [30•].

Clinical Presentation

Medulloblastoma typically originates from the posterior fossa, and most clinical symptoms at presentation result from increased intracranial pressure (ICP) due to obstructive hydrocephalus and cerebellar dysfunction. In a study from 2012 by Brasme et al., the most frequent symptoms leading to a MB diagnosis were vomiting, headache, ataxia, and neuropsychological symptoms depending upon age at presentation [31]. In this report, psychomotor regression was seen early in children under 3 years of age, while declining school performance often prompted investigation in older patients. The median time from symptom onset to definitive diagnosis was 65 days; however, survival and neurological outcome were independent in this report, which predates molecular risk stratification [31]. Despite medulloblastoma being a rapidly growing tumor, obtaining a final diagnosis can be challenging due to non-specific symptoms or a completely normal neurological exam.

Diagnosis

Once symptoms are recognized, neuroimaging is obtained; typically, a non-contrast head CT scan, which does not require anesthesia, can be safely performed on critically ill children and is widely available. In most instances, a hyperdense mass in the posterior fossa surrounded by vasogenic edema and hydrocephalus is seen.

Thereafter, confirmatory magnetic resonance imaging (MRI) is performed to provide further detail and identify metastases. Tumors are typically hypointense to grey matter on T1-weighted imaging with heterogenous gadolinium enhancement. A recent article by Perreault et al. demonstrated a correlation between tumor location/ enhancement patterns and molecular subgroups [32•]. Most WNT tumors occurred along the cerebellar peduncle/cerebellopontine angle, while SHH medulloblastoma occurred in the cerebellar hemisphere. Group 3 and group 4 tumors were found in the midline, occupying the fourth ventricle. Group 3 tumors had illdefined margins, and group 4 MB did not enhance (Fig. 2). Differential diagnosis of a posterior fossa mass includes pilocytic astrocytoma, ependymoma, atypical teratoid/rhabdoid tumor, exophytic brainstem glioma, embryonal tumor with multi-layered rosettes, and choroid plexus papilloma. In adults, metastatic disease of an extracranial primary site should be considered. Spectroscopy can be helpful in distinguishing highfrom low-grade tumors; tractography may help surgeons minimize postoperative morbidity [33].

Staging

In 1969, Chang proposed the following staging criteria for medulloblastoma: 1) M0: no evidence of metastatic disease; 2) M1: positive cerebrospinal fluid (CSF) cytology without gross tumor visible on MRI; 3) M2: intracranial metastasis; 4): M3: metastasis within the spinal subarachnoid space and; and 5) M4: disease outside of the neuraxis, most common in the bone and bone marrow [34].

Ideally, extent of disease evaluation should be performed preoperatively and include pre- and post-contrast brain and spine MRI and lumbar puncture (LP) only if the patient is stable without signs or symptoms of increased Fig. 2 Characteristic MRI images of medulloblastoma subgroups and the histopathological variants commonly associated with them. a Characteristic location of WNT MB in the cerebellopontine angle region showing e classic histology. b SHH MB in a hemispheric location with fdesmoplastic nodular histology. c Group 3 MB with contrast enhancement in the midline/ fourth ventricle showing g large cell/anaplastic features. d Group 4 MB showing no enhancement in the midline/fourth ventricle, often associated with h classic histology



ICP. If deferred, LP should be performed no sooner than 14 days postoperatively to avoid false-positive results, but prior to the initiation of therapy, preferably 30 days from diagnosis.

Histopathology

The 2016 edition of the WHO CNS classification recognizes that both histology and genetic signature are important in defining MB subgroups, all of which are grade IV and characterized by small round blue cells usually of cerebellar origin. Histopathologic variants of MB include: 1) classic with minimal or no differentiation; 2) desmoplastic/nodular (DNMB) which has nodules of differentiated cells with internodular fibrosis; 3) medulloblastoma with extensive nodularity (MBEN) with more significant nodularity; and 4) large cell/anaplastic (LC/A-MB) characterized by bizarre atypical polymorphic cells, nuclear molding, high mitotic rate, and apoptotic bodies (Fig. 2) [4]. Classic histology is seen in almost all WNT medulloblastomas [35], the majority of Group 3 and 4 tumors [13], and occasionally in SHH-driven tumors which are largely desmoplastic/nodular [36]. MBEN, typically seen in very young children with SHH/TP53WT, can be associated with Gorlin syndrome [37]. LC/A-MB is most often seen in SHH/TP53 mutant and infant group 3 MB [38, 39] (Fig. 2).

The origin of medulloblastoma cells has long been debated. Gibson et al. has shown that WNT subgroup tumors derive from cells in the lower rhombic lip, while cerebellar granule neuron progenitors (CGNPs) are the probable cell of origin for SSH medulloblastoma [18]. As of today, it is still uncertain where group 3 and group 4 tumors originate from.

Risk Stratification

Risk stratification guidelines include age, extent of resection, presence of metastasis, and histological subtypes. Averagerisk patients are older than 3 years of age and have M0 disease with less than 1.5 cm² of residual tumor after resection. Highrisk children have microscopic or macroscopic metastasis, more than 1.5 cm² of mass present after surgery, and/or large cell anaplastic histology. Children under 3 years of age represent a distinctive-risk group and require a different treatment approach, which omits upfront radiation due to the unacceptable risk of neurocognitive impairment.

Experts convened at a consensus conference in 2015 in Heidelberg to refine risk stratification in the context of molecular subtypes [40••]. The low-risk group (>90% survival) includes non-metastatic WNT medulloblastoma diagnosed before age 16 years, and non-metastatic group 4 tumors with loss of chromosome 11 or gain of chromosome 17. The standard-risk group (75-90% survival) consists of nonmetastatic SHH medulloblastoma without TP53 mutation or MYCN amplification; non-metastatic group 4 tumors without chromosome 11 loss; and non-metastatic, non-MYC-amplified group 3 tumors. The high-risk group (50-75% survival) includes metastatic non-infant TP53 wild type and nonmetastatic MYCN-amplified SHH medulloblastoma, and metastatic group 4 medulloblastoma. The very high-risk group (< 50% survival) is comprised of TP53-mutated SHH medulloblastoma and metastatic group 3 tumors.

The transcriptome of adult medulloblastomas differs considerably from pediatric counterparts, both in tumor biology and prognostic impact [41]. Therefore, age-specific classification is necessary when designing clinical trials for adult medulloblastoma.

Treatment

Surgery

The first step in the management of medulloblastoma is maximal safe surgical resection, which is usually performed by posterior craniotomy. A recent retrospective analysis of almost 800 medulloblastoma patients found a progression-free survival benefit between gross total and subtotal resection, but not between gross total and near total resection, which questions the role of second-look surgeries to remove small residual tumors (less than 1.5 cm²), especially when the risk of neurological sequelae is increased [42•]. For increased extent of resection, the study did not observe significant survival benefit in WNT, SHH, group 3, or M0 group 4 patients.

While extensive neurosurgical procedures carry inherent risks, cerebellar mutism syndrome (CMS) is a complication unique to posterior fossa surgery. One to two days postoperatively, patients develop speech impairment, oral apraxia, emotional lability, hypotonia, and ataxia. Neuropsychologic testing reveals difficulty with executive function, linguistic processing, visual spatial abilities, and affective modulation [43]. In 2006, the Children's Oncology Group (COG) reported an incidence of 25% in all patients with medulloblastoma; risk factors included younger age, larger tumor volume, and midline tumor location [44]. Tractography reveals disruption of cerebrocerebellar pathways [33]. Most recently, Jabarkheel et al. found that independent of tumor volume and location, subgroup of MB is predictive of CMS as follows: group 4 tumors (35%), group 3 (31%), WNT (21%), and SHH (7%) [45]. Patients with moderate to severe CMS often have long-term neurocognitive effects, which may be compounded by other effects of treatment for medulloblastoma [46].

Radiation

Because survivors of medulloblastoma treated with radiation therapy (RT) are often left with neurocognitive, neuroendocrine, and auditory sequelae, decreasing radiation dose and volume while maintaining survival is crucial [47]. Averagerisk patients over 6 years of age receive 23.4-Gy craniospinal irradiation (CSI), a tumor bed boost to 55.8 Gy, and weekly vincristine during radiotherapy, beginning roughly 30 days post-resection [48, 49]. Initiating RT sooner than 3 weeks pos-operatively, delaying treatment, or attempting to further reduce the CSI dose to 18 Gy resulted in inferior OS [50, 51], while hyperfractionation had no effect [52]. For high-risk patients, 36-Gy CSI and additional boost to metastatic sites besides the tumor bed are standard. Concomitant carboplatin as a radiosensitizer did not alter outcomes for this patient population [53].

While RT has improved overall survival in MB patients, dose-dependent late effects primarily related to CSI and seen typically 5 years post-treatment need to be monitored. The COG has published long-term follow-up guidelines to aid the clinicians [54•]. Radiation to the hypothalamic-pituitary axis, associated with hypothyroidism, growth hormone and ACTH deficiency, and altered metabolism, may result in obesity and delayed puberty [55, 56].

A systematic review in 2015 by Hanzlik et al. found that even with reduced-dose RT, pediatric medulloblastoma survivors had lower IQ, memory, and executive function scores with ongoing decline compared with normal population and patients receiving only focal radiation [57].

The advent of proton beam RT has significantly improved both acute and long-term morbidity associated with photons without reducing OS [58•]. Acute toxicities (fatigue, esophagitis, and nausea) and late effects (neurocognitive and ototoxicity) are reduced with proton beam RT. However, alopecia and radiation necrosis may occur more often with proton radiation. [59, 60]. Other late effects including cavernomas, meningiomas, and high-grade second malignancies may also develop following radiation (from either CSI or the boost), but risk factors, other than familial cancer predisposition syndromes, require further investigation [61, 62].

Chemotherapy

Chemotherapy was first added to medulloblastoma treatment in the 1970s, significantly improving both progression-free survival (PFS) and OS rates, allowing reduction of CSI dose for average-risk patients [63]. All studies investigating optimal time for chemotherapy concluded that surgery-radiationchemotherapy sequence is favored over "sandwich chemotherapy," or pre-radiation chemotherapy only [64–67]. Alkylators [68] represent the backbone of maintenance chemotherapy, and currently, the "Packer regimen" (vincristine and cisplatin plus either lomustine or cyclophosphamide) is the standard of care for standard-risk patients [48]. For patients with high-risk disease, maintenance chemotherapy includes higher cumulative dose cyclophosphamide [69]. The incorporation of high-dose chemotherapy with autologous stem cell transplantation (thiotepa-containing regimens) did not improve outcomes for high-risk patients but was associated with significant neuro- and hepatotoxicity [70, 71]. Acute and long-term side effects of chemotherapy are well described. While both pediatric and adult MB protocols traditionally include vincristine, adults may develop neuropathy sooner, with more severity, requiring dose reduction or complete omission of the drug [72]. Patients receiving alkylating agents are at risk of developing secondary malignancies; the cumulative 10-year incidence of treatment-related tumors was 4.2% on the standard-risk COG protocol A9961 [73]. Grade 3 or 4 ototoxicity due to both radiotherapy and the use of cisplatin affects 20–25% of the patients [48].

Infant Medulloblastoma

The Head Start group of protocols have shown that chemotherapy improves survival while avoiding or delaying radiotherapy to preserve the intellect of young children [74, 75]. In 2005, Rutkowski et al. showed that the addition of intrathecal methotrexate improved survival in infants with desmoplastic variants of MB [76]. Several subsequent studies confirmed that infants with DNMB and MBEN have outcomes around 80% [74, 77, 78]. A recently closed COG study included systemic high-dose methotrexate during induction plus three cycles of thiotepa and carboplatin followed by stem cell rescue in children less than 3 years of age. Incorporating molecular subgroups as part of risk stratification (as in the Head Start 4 protocol) is being investigated.

Adult Medulloblastoma

Medulloblastoma accounts for less than 1% of adult intracranial tumors [79]. Seventy percent of cases are classified as SHH (desmoplastic/nodular or extensive nodularity histology), followed by group 4 and WNT, each around 15%, and rarely group 3 [39]. WNT is the most favorable subtype, with an 80% 5-year OS, slightly lower than 90% observed in children younger than 16 years [16•]. This difference is largely unexplained due to the rarity of the tumor and the heterogeneity of the treatment. Craniospinal irradiation with a boost to the tumor bed is considered the standard of care, and while maintenance chemotherapy is feasible, its role has not yet been investigated in a prospective randomized study [80–82].

Refractory and Recurrent Disease

Refractory and recurrent medulloblastomas are responsible for 95% of medulloblastoma-associated death [83]. Aside from the very young patients who can be salvaged with radiation therapy following a treatment with a chemotherapy-only approach, survival after relapsed medulloblastoma is grim [84]. The combination of temozolomide and irinotecan has been utilized in the relapsed medulloblastoma setting due to its favorable toxicity profile and the feasibility in heavily pretreated patients, and showed a 30% response rate with a median survival of 16 months [85, 86]. The most recent COG

Table 2 Ongoing clinical trials for newly diagnosed medulloblastoma and ongoing clinical trials for relapsed/progressive medulloblastoma

ClinicalTrials. gov Identifier	Phase	Treatment	MB subgroup (if applicable)	Sponsor
Ongoing clinical	trials for	newly diagnosed medulloblastoma		
NCT01878617	II	Clinical and molecular risk-directed therapy for newly diagnosed MB	WNT: 3 treatment arms SHH: 2 treatment arms Non-WNT/non-SHH: 3 treatment arms	St. Jude Children's Research Hospital
NCT02875314	IV	HeadStart4: newly diagnosed children (< 10 years old) with medulloblastoma	Non-WNT/non-SHH	Nationwide Children's Hospital
NCT02724579	Π	ACNS 1422: reduced CSI and chemotherapy in younger patients with newly diagnosed WNT-driven medulloblastoma	WNT	Children's Oncology Group
NCT02066220	II/III	SIOP PNET 5 medulloblastoma	WNT	Universitatsklinikum Hamburg-Eppendorf
NCT02025881	I/II	Study of sequential high-dose chemotherapy in children with high-risk MB (HR MB-5)	N/A	Gustave Roussy, Cancer Campus, Grand Paris
NCT01857453	Π	Dose decrease for RT associated with chemotherapy for treatment of standard-risk adult medulloblastoma	N/A	Central Hospital, Nancy, France
Ongoing clinical	trials for	relapsed/progressive medulloblastoma		
NCT01356290	II	Metronomic and targeted anti-angiogenesis therapy (MEMMAT)	N/A	Medical University of Vienna
NCT03904862	I/II	CX-4945 (silmitasertib), casein kinase II inhibitor	SHH	Pediatric Brain Tumor Consortium
NCT03936465	Ι	BMS-986158 bromodomain (BRD) and extra-terminal domain (BET) inhibitor	MYC or MYCN amplification or translocation, or high copy number gain, BRD3 or BRD4 translocation	Dana-Farber Cancer Institute
NCT02359565	Ι	Pembrolizumab	N/A	National Cancer Institute
NCT03173950	II	Nivolumab	N/A	National Cancer Institute
NCT03389802	Ι	APX055M monoclonal antibody binding to CD40	N/A	Pediatric Brain Tumor Consortium
NCT00089245	Ι	Intrathecal radiolabeled monoclonal antibody against B7-H3 (I-8H9–omburtamab)	N/A	Y-mAbs Therapeutics
NCT03911388	Ι	G207-oncolytic herpes simplex virus 1	N/A	University of Alabama at Birmingham
NCT02962167	Ι	MV-NIS-modified measles virus	N/A	Sabine Muller, MD, PhD
NCT03043391	Ι	PVSRIPO-oncolytic poliovirus	N/A	Istari Oncology, Inc.
NCT03299309	Ι	PEP-CMV-cytomegalovirus-specific peptide vaccine	N/A	Gary Archer, PhD
NCT03500991	Ι	HER2-specific CAR T cell locoregional immunotherapy	N/A	Seattle Children's Hospital
NCT03638167	Ι	EGFR806-specific CAR T cell locoregional immunotherapy	N/A	Seattle Children's Hospital
NCT03387020	Ι	Ribociclib (cyclin D1/CDK4 and CDK6 inhibitor and everolimus	N/A	Pediatric Brain Tumor Consortium
NCT03434262	Ι	Ribociclib and gemcitabine	Groups 3 and 4 MB	St. Jude Children's Research
		Ribociclib and trametinib	WNT and SHH MB	Hospital
		Ribociclib and sonidegib	SHH	
NCT03598244	Ι	Volitinib–c-Met inhibitor	N/A	National Cancer Institute
NCT02095132	I/II	Adavosertib (MK-1775)–WEE1 inhibitor and irinotecan	N/A	National Cancer Institute
NCT03213678	II	Samotolisib (LY3023414)–PI3K/mTOR inhibitor Pediatric MATCH Treatment Trial	TSC or PI3K/mTOR mutations	National Cancer Institute
NCT03213665	II	Tazemetostat-EZH2 inhibitor	EZH2, SMARCA4, or SMARCB1 mutation	National Cancer Institute
NCT03233204	II	Olaparib-Pediatric MATCH Treatment Trial	Defect in DNA damage repair pathway	National Cancer Institute
NCT04023669	Ι	Prexasertib (LY2606368)–CHK1/2 inhibitor and cyclophosphamide	SHH, groups 3 and 4, indeterminate MB	St. Jude Children's Research Hospital
		Prexasertib (LY2606368)–CHK1/2 inhibitor and gemcitabine	Groups 3 and 4 MB	
NCT02644291	Ι	Mebendazole	N/A	Sidney Kimmel Comprehensive Cancer Center at Johns Hopkins

phase II trial is investigating the synergistic effect of bevacizumab when added to temozolomide and irinotecan (ACNS 0821), with preliminary results showing a median event-free survival (EFS) of 9 months vs. 6 months and a median OS of 19 months vs. 16 months when compared with the standard arm [87].

New Approaches

The recent refinement of risk stratification provides an opportunity to personalize therapy, while the use of targeted therapy allows dose reduction of cytotoxic chemotherapy, resulting in the mitigation of treatment-related toxicity and maintaining favorable OS. Table 2 summarizes ongoing trials for both newly diagnosed and recurrent/progressive medulloblastomas.

Phoenix et al. has shown that WNT medulloblastomas contain aberrant vascular networks and disrupted blood-brainbarrier (BBB), which allow for increased concentrations of systemic chemotherapy; de-escalation of therapy in low-risk patients is tested in multiple clinical trials [88].

In adult SHH medulloblastoma inhibiting SMO and blocking downstream signaling (e.g., vismodegib) can be a successful strategy, as these tumors frequently (80%) harbor PTCH1 or SMO mutations. Of note, the use of these inhibitors is associated with disrupted bone homeostasis, limiting its use in skeletally immature patients. A current St. Jude upfront MB trial adds vismodegib as maintenance therapy for skeletally mature patients with SHH MB. Some tumors, however, present with inherent resistance to SMO inhibitors by harboring either certain SMO mutations, which impair drug binding or genetic alterations in the downstream effectors SUFU or Gli [89]. Alternatively, prolonged exposure to smoothened inhibitors may result in acquired resistance by reactivation of the hedgehog pathway. To overcome these limitations, silmitasertib, a casein kinase 2 inhibitor which blocks signaling at the most terminal component Gli, is currently in a phase I/II trial for patients over 3 years with recurrent/relapsed SHH medulloblastoma.

The aforementioned St. Jude trial includes strata for group 3 and group 4 intermediate- and high-risk medulloblastoma patients, adding pemetrexed and gemcitabine to the standard therapy. Based upon promising preclinical data, trials targeting the PI3K, mTOR, cyclin-dependent kinase (CDK), or BET bromodomain pathways are ongoing for recurrent and/or refractory group 3 MB [90–92].

Immunotherapy and small-molecule inhibitors form the backbone of novel strategies for many refractory tumors. The programmed cell death protein 1 (PD1) inhibitors pembrolizumab and nivolumab, successful at treating lung and breast cancer CNS metastases, are now being investigated in the treatment of refractory primary CNS tumors. Their therapeutic benefit in this population must be balanced with immune-related adverse events including encephalitis, meningitis, and myelitis [93]. Antibodies against the molecules of the co-stimulatory pathways represent the next generation of immune checkpoint inhibitors. APX005M, a monoclonal antibody against CD40, is currently in a phase I trial. Kramer et al. recently reported that intrathecal B7-H3, another immune checkpoint protein expressed in embryonal tumors, was well tolerated in patients with leptomeningeal disease including a subset of MB patients and possibly demonstrating improved outcome in the relapsed/refractory setting [94, 95].

Oncolytic viral therapy, whereby viruses selectively inhibit malignant cells by producing a robust immune response, is another promising strategy for recurrent/progressive MB [96]. These viruses, including the herpes simplex virus variant G207, a modified measles virus (MV-NIS), and the polio/ rhinovirus recombinant PVSRIPO require local delivery and are under investigation in phase I trials. Cytomegalovirus (CMV) proteins, often selectively expressed in brain tumors, have been the target of numerous vaccine trials over the past several years. A PEP-CMV vaccine trial in pediatric patients with recurrent MB is underway [97].

Chimeric antigen receptor (CAR) T cells, an emerging therapeutic approach, has been particularly challenging for CNS tumors due to limitations in identifying tumor-specific antigens. Forty percent of medulloblastomas express human epidermal growth factor 2 (HER2), a poor prognostic indicator [98]. Patients aged 1–26 years may be eligible for a current trial, in which CAR T cells expressing various epidermal growth factor receptor (EGFR) antigens are infused locoregionally. Finally, because natural killer (NK) cells can recognize and eliminate cancer cells without antigen specificity, it became the focus of an ongoing study using intrathecal autologous NK cells [99].

Small molecular inhibitors are also being used to treat relapsed/refractory medulloblastoma, regardless of molecular subtypes. Several clinical trials currently use these agents including CDK4/6, c-Met, Wee1, PI3K/mTOR, EZH2, or CHK1/2, either as monotherapy or in combination with conventional chemotherapy.

Conclusion

Recent innovations in medulloblastoma research allowed us to better understand pathogenesis and molecular characteristics, resulting in advanced-risk stratification models, new therapeutic approaches, and overall improved survivals while preserving quality of life. Large multicenter, international trials, especially in adults with MB, are needed to further understand the biology and improve both short- and long-term outcomes. Acknowledgments We thank Joseph Olechnowicz, MA for his help with editing the manuscript; Marc Rosenblum, MD for providing histopathology images; and Sofia Haque, MD for her help providing radiologic images.

Compliance with Ethical Standards

Conflict of Interest None of the authors has any potential conflicts of interest to disclose.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- •• Of major importance
- Kunscher LJ. Harvey Cushing and medulloblastoma. Arch Neurol. 2002;59:642–5.
- [2] Paterson R, Farr RF. Cerebellar medulloblastoma: treatment by irradiation of the whole central nervous system. Acta Radiol. 1953;39:323–36.
- [3] Tait DM, Thornton-Jones H, Bloom HJ, Lemerle J, Morris-Jones P. Adjuvant chemotherapy for medulloblastoma: the first multicenter control trial of the International Society of Pediatric Oncology (SIOP I). Eur J Cancer. 1990;26:464–9.
- [4] Louis DN, Perry A, Reifenberger G, von Deimling A, Figarella-Branger D, Cavenee WK, et al. The 2016 World Health Organization classification of tumors of the central nervous system: a summary. Acta Neuropathol. 2016;131:803–20.
- [5] Ostrom QT, de Blank PM, Kruchko C, Petersen CM, Liao P, Finlay JL, et al. Alex's lemonade stand foundation infant and childhood primary brain and central nervous system tumors diagnosed in the United States in 2007–2011. Neuro-Oncology. 2015;16(Suppl 10):x1–x36.
- [6] Siegel, et al. Cancer statistics, 2015. CA Cancer J Clin. 2015;65:5– 29.
- [7] Ostrom QT, Gittleman H, Truitt G, Boscia A, Kruchko C, Barnholtz-Sloan JS. CBTRUS statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2011-2015. Neuro-Oncology. 2018;20(Suppl 4):1–86.
- [8] Sun T, Plutynski A, Ward S, Rubin JB. An integrative view on sex differences in brain tumors. Cell Mol Life Sci. 2015;72:3323–42.
- [9•] Khanna V, Achey RL, Ostrom QT, Block-Beach H, Kruchko C, Barnholtz-Sloan JS, et al. Incidence and survival trends for medulloblastomas in the United States from 2001 to 2013. J Neurooncol. 2017;135:433–41 The study examined population-level trends in MB incidence and survival using data from 2001 to 2013. Overall incidence rate remained stable with slight male predominance, and the incidence was highest in the youngest patients. Non-significant trends in the data suggested disparities by race.
- [10••] Waszak SM, et al. Spectrum and prevalence of genetic predisposition in medulloblastoma: a retrospective genetic study and prospective validation in a clinical trial cohort. Lancet Oncol. 2018;19:785–98 This international, multicenter study found that the prevalence of genetic predisposition syndromes in MB patients was 6%, with SHH MB having the highest number of cases. It concluded that genetic counselling and testing should be a standard of care for WNT and SHH MB patients.

- [11] Pomeroy SL, Tamayo P, Gaasenbeek M, Sturla LM, Angelo M, McLaughlin ME, et al. Prediction of central nervous system embryonal tumour outcome based on gene expression. Nature. 2002;415:436–42.
- [12] Cavalli FGM, Remke M, Rampasek L, Peacock J, Shih DJH, et al. Intertumoral heterogeneity within medulloblastoma subgroups. Cancer Cell. 2017;31:737–54.
- [13] Taylor MD, Northcott PA, Korshunov A, Remke M, Cho YJ, Clifford SC, et al. Molecular subgroups of medulloblastoma: the current consensus. Acta Neuropathol. 2012;123:465–72.
- [14] Morrissy AS, Cavalli FMG, Remke M, Ramaswamy V, Shih DJH, Holgado BL, et al. Spatial heterogeneity in medulloblastoma. Nat Genet. 2017;49:780–8.
- [15] Clifford SC, Lusher ME, Lindsey JC, Langdon JA, Gilbertson RJ, Straughton D, et al. Wnt/Wingless pathway activation and chromosome 6 loss characterize a distinct molecular subgroup of medulloblastomas associated with a favorable prognosis. Cell Cycle. 2006;5:2666–70.
- [16•] Remke M, Hielscher T, Northcott PA, Witt H, Ryzhova M, et al. Adult medulloblastoma comprises three major molecular variants. J Clin Oncol. 2011;29:2717–23 This study analyzed the transcriptome and DNA copy number alterations of 28 adult MB patients and that both tumor biology and prognostic impact differed from pediatric MB, suggesting the need for age specific classification system.
- [17••] Northcott PA, Buchhalter I, Morrissy AS, Hovestadt, et al. The whole-genome landscape of medulloblastoma subtypes. Nature. 2017;547:311-7 This study reported a comprehensive, genome-wide analysis of a cohort of 491 previously untreated MBs, which identified novel subgroup-specific driver genes, epigenetic subtypes, and candidate targets for therapy.
- [18] Gibson P, Tong Y, Robinson G, Thompson MC, Currle DS, Eden C, et al. Subtypes of medulloblastoma have distinct developmental origins. Nature. 2010;468:1095–9.
- [19] Robinson G, Parker M, Kranenburg TA, Lu C, Chen X, Ding L, et al. Novel mutations target distinct subgroups of medulloblastoma. Nature. 2012;488:43–8.
- [20] Iorgulescu JB, Van Ziffle J, Stevers M, Grenert JP, Bastian BC. Deep sequencing of WNT-activated medulloblastomas reveals secondary SHH pathway activation. Acta Neuropathol. 2018;135:635–8.
- [21] Choudhry Z, Rikani AA, Choudhry AM, Tariq S, Zakaria F, Asghar MW, et al. Sonic hedgehog signaling pathway: a complex network. Ann Neurosci. 2014;21:28–31.
- [22] Varjosalo M, Taipale J. Hedgehog: functions and mechanisms. Genes Dev. 2008;22:2454–72.
- [23] Pietrobono S, Gagliardi S, Stecca B. Non-canonical hedgehog signaling pathway in cancer: activation of GLI transcription factors beyond Smoothened. Front Genet. 2019;10:556.
- [24] Wetmore C, Eberhart DE, Curran T. Loss of p53 but not ARF accelerates medulloblastoma in mice heterozygous for Patched. Cancer Res. 2001;61:513–6.
- [25] Lafay-Cousin L, Bouffet E, Strother D, Rudneva V, Hawkins C, Eberhart C, et al. Phase II study of nonmetastatic desmoplastic medulloblastoma in children younger than 4 years of age: a report of the Children's Oncology Group (ACNS1221). J Clin Oncol. 2020;38:223–31.
- [26] Remke M, Ramaswamy V, Peacock J, Shih DJ, Koelsche C, et al. TERT promoter mutations are highly recurrent in SHH subgroup medulloblastoma. Acta Neuropathol. 2013;126:917–29.
- [27] Northcott PA, Shih DJ, Peacock J, Garzia L, Morrissy AS. Subgroup-specific structural variation across 1,000 medulloblastoma genomes. Nature. 2012;488:49–56.
- [28] Northcott PA, Robinson GW, Kratz CP, Mabbott DJ, Pomeroy SL, Clifford SC, et al. Medulloblastoma. Nat Rev Dis Primers. 2019;5:11.

- [29] Northcott PA, Lee C, Zichner T, Stutz AM, Erkek S, et al. Enhancer hijacking activates GFI1 family oncogenes in medulloblastoma. Nature. 2014;511:428–34.
- [30•] Shih DJ, Northcott PA, Remke M, Korshunov A, Ramaswamy V, et al. Cytogenetic prognostication within medulloblastoma subgroups. J Clin Oncol. 2014;32:886–96 This study identified molecular biomarkers from a set of 673 MB samples and designed risk stratification models based on clinical and cytogenetic information. This led to the identification of very low-risk and very high-risk patients within MB subgroups.
- [31] Brasme JF, Chalumeau M, Doz F, Lacour B, Valteau-Couanet D, Gaillard S, et al. Interval between onset of symptoms and diagnosis of medulloblastoma in children: distribution and determinants in a population-based study. Eur J Pediatr. 2012;171:25–32.
- [32•] Perrault S, Ramaswamy V, Achrol AS, Chao K, Liu TT, et al. MRI surrogates for molecular subgroups of medulloblastoma. Am J Neuroradiol. 2014;35:1263–9 This study showed that tumor location and enhancement pattern are predictive of molecular subgroups of pediatric MB and may serve as a surrogate for genomic testing.
- [33] Oh ME, Driever PH, Khajuria RK, Rueckriegel SM, Koustenis E, Bruhn H, et al. DTI fiber tractography of cerebro-cerebellar pathways and clinical evaluation of ataxia in childhood posterior fossa tumor survivors. J Neuro-Oncol. 2017;131:267–76.
- [34] Chang CH, Housepian EM, Herbert C Jr. An operative staging system and a megavoltage radiotherapeutic technic for cerebellar medulloblastomas. Radiology. 1969;93:1351–9.
- [35] Ellison DW, Dalton J, Kocak M, Nicholson SL, Fraga C, Neale G, et al. Medulloblastoma: clinicopathological correlates of SHH, WNT, and non-SHH/WNT molecular subgroups. Acta Neuropathol. 2011;121:381–96.
- [36] Kool M, Koster J, Bunt J, Hasselt NE, Lakeman A, et al. Integrated genomics identifies five medulloblastoma subtypes with distinct genetic profiles, pathway signatures and clinicopathological features. PLoS One. 2008;3:e3008.
- [37] Garre ML, Cama A, Bagnasco F, Morana G, Giangaspero F, Brisigotti M, et al. Medulloblastoma variants: age-dependent occurrence and relation to Gorlin syndrome–a new clinical perspective. Clin Cancer Res. 2009;15:2463–71.
- [38] Eberhart CG, Kepner JL, Goldthwaite PT, Kun LE, Duffner PK, Friedman HS, et al. Histopathologic grading of medulloblastomas: a pediatric oncology group study. Cancer. 2002;94:552–60.
- [39] Kool M, Korshunov A, Remke M, Jones DT, Schlanstein M, et al. Molecular subgroups of medulloblastoma: an international metaanalysis of transcriptome, genetic aberrations, and clinical data of WNT, SHH, group 3, and group 4 medulloblastoma. Acta Neuropathol. 2012;123:473–84.
- [40••] Ramaswamy V, Remke M, Bouffet E, Bailey S, Clifford SC, et al. Risk stratification of childhood medulloblastoma in the molecular era: the current consensus. Acta Neuropathol. 2016;131:821–31 This article summarizes a new proposed risk stratification scheme based on several published and unpublished clinical, histopathological and biological studies.
- [41] Northcott PA, Hielscher T, Dubuc A, Mack S, Shih D, Remke M, et al. Pediatric and adult sonic hedgehog medulloblastomas are clinically and molecularly distinct. Acta Neuropathol. 2011;122: 231–40.
- [42•] Thompson EM, Hielscher T, Bouffet E, Remke M, Luu B, et al. Prognostic value of medulloblastoma extent of resection after accounting for molecular subgroup: an integrated clinical and molecular analysis. Lancet Oncol. 2016;17:484–95 This study investigated the prognostic benefit of extent of resection and observed PFS benefit of gross total resection over subtotal resection but not between gross total and near total resection, and increased extent of resection only benefited metastatic group 4 patients.

- [43] Schmahmann JD. Neuroanatomy of pediatric postoperative cerebellar cognitive affective syndrome and mutism. Neurology. 2019;93:693–4.
- [44] Robertson PL, Muraszko KM, Holmes EJ, Sposto R, Packer RJ, Gajjar A, et al. Incidence and severity of postoperative cerebellar mutism syndrome in children with medulloblastoma: a prospective study by the Children's Oncology Group. J Neurosurg. 2006;105: 444–51.
- [45] Jabarkheel R, Amayiri N, Yecies D, Huang Y, Toescu S, et al. Molecular correlates of cerebellar mutism syndrome in medulloblastoma. Neuro Oncol. 2020;22:290–7.
- [46] Catsman-Berrevoets CE, Aarsen FK. The spectrum of neurobehavioral deficits in the posterior fossa syndrome in children after cerebellar tumour surgery. Cortex. 2010;46:933–46.
- [47] Laughton SJ, Merchant TE, Sklar CA, Kun LE, Fouladi M, Broniscer A, et al. Endocrine outcomes for children with embryonal brain tumors after risk-adapted craniospinal and conformal primary-site irradiation and high-dose chemotherapy with stemcell rescue on the SJMB-96 trial. J Clin Oncol. 2008;26:1112–8.
- [48] Packer RJ, Gajjar A, Vezina G, Rorke-Adams L, Burger PC, Robertson PL, et al. Phase III study of craniospinal radiation therapy followed by adjuvant chemotherapy for newly diagnosed average-risk medulloblastoma. J Clin Oncol. 2006;24:4202–8.
- [49] Merchant TE, Kun LE, Krasin MJ, Wallace D, Chintagumpala MM, Woo SY, et al. Multi-institutional prospective trial of reduced-dose craniospinal irradiation (23.4 Gy) followed by conformal posterior fossa (36 Gy) and primary site irradiation (55.8 Gy) and dose-intensive chemotherapy for average-risk medulloblastoma. Int J Radiat Oncol Biol Phys. 2008;70:782–7.
- [50] Chin AL, Moding EJ, Donaldson SS, Gibbs IC, Soltys SG, Hiniker SM, et al. Survival impact of postoperative radiotherapy timing in pediatric and adolescent medulloblastoma. Neuro-Oncology. 2018;20:1133–41.
- [51] Michalski J, Vezina G, Burger P, Gajjar A, Pollack I, et al. Preliminary results of COG ACNS0331: a phase III trial of involved field radiotherapy (IFRT) and low dose craniospinal irradiation (LD-CSI) with chemotherapy in average risk medulloblastoma: a report from the Children's Oncology Group. Neuro-Oncology. 2016;18(Suppl 3):122.
- [52] Lannering B, Rutkowski S, Doz F, Pizer B, Gustafsson G, Navajas A, et al. Hyperfractionated versus conventional radiotherapy followed by chemotherapy in standard-risk medulloblastoma: results from the randomized multicenter HIT-SIOP PNET 4 trial. J Clin Oncol. 2012;30:3187–93.
- [53] Hwang EI, Kool M, Burger PC, Capper D, Chavez L, Brabetz S, et al. Extensive molecular and clinical heterogeneity in patients with histologically diagnosed CNS-PNET treated as a single entity: a report from the Children's Oncology Group Randomized ACNS0332 trial. J Clin Oncol. 2018;36:3388–95.
- [54•] http://www.survivorshipguidelines.org/pdf/2018/COG_LTFU_ Guidelines_v5.pdf. Accessed 16 Apr 2020. This risk-based, exposure related clinical practice guideline provided recommendations for screening and management of late effects in MB survivors.
- [55] Sklar CA, Constine LS. Chronic neuroendocrinological sequelae of radiation therapy. Int J Radiat Oncol Biol Phys. 1995;31:1113– 21.
- [56] Uday S, Murray RD, Picton S, Chumas P, Raju M, Chandwani M, et al. Endocrine sequelae beyond 10 years in survivors of medulloblastoma. Clin Endocrinol. 2015;83:663–70.
- [57] Hanzlik E, Woodrome SE, Abdel-Baki M, Geller TJ, Elbabaa SK. A systematic review of neuropsychological outcomes following posterior fossa tumor surgery in children. Childs Nerv Syst. 2015;31:1869–75.
- [58•] Kahalley LS, Peterson R, Ris MD, Janzen L, Okcu MF, et al. Superior intellectual outcomes after proton radiotherapy compared

with photon radiotherapy for pediatric medulloblastoma. J Clin Oncol. 2020;38:454–61 This study demonstrated that proton radiotherapy is associated with more favorable intellectual outcomes when compared with photon radiotherapy.

- [59] Kralik SF, Ho CY, Finke W, Buchsbaum JC, Haskins CP, Shih CS. Radiation necrosis in pediatric patients with brain tumors treated with proton radiotherapy. AJNR Am J Neuroradiol. 2015;36: 1572–8.
- [60] Davanzo J, Greiner RJ, Barbour M, Rizk E. Radiation necrosis following proton beam therapy in the pediatric population: a case series. Cureus. 2017;9:e1785.
- [61] Kessler AT, Bhatt AA. Brain tumor post-treatment imaging and treatment-related complications. Insights Imaging. 2018;9:1057– 75.
- [62] Morgenstern PF, Shah K, Dunkel IJ, Reiner AS, Khakoo Y, Rosenblum MK, et al. Meningioma after radiotherapy for malignancy. J Clin Neurosci. 2016;30:93–7.
- [63] Packer RJ, Goldwein J, Nicholson HS, Vezina LG, Allen JC, Ris MD, et al. Treatment of children with medulloblastomas with reduced-dose craniospinal radiation therapy and adjuvant chemotherapy: a Children's Cancer Group Study. J Clin Oncol. 1999;17: 2127–36.
- [64] Hoff KV, Hinkes B, Gerber NU, Deinlein F, Mittler U, et al. Longterm outcome and clinical prognostic factors in children with medulloblastoma treated in the prospective randomized multicenter trial HIT'91. Eur J Cancer. 2009;45:1209–17.
- [65] Bailey CC, Gnekow A, Wellek S, Jones M, Round C, Brown J, et al. Prospective randomized trial of chemotherapy given before radiotherapy in childhood medulloblastoma. International Society of Pediatric Oncology (SIOP) and the (German) Society of Pediatric Oncology (GPO): SIOP II. Med Pediatr Oncol. 1995;25:166–78.
- [66] Allen J, Donahue B, Mehta M, Miller DC, Rorke LB, Jakacki R, et al. A phase II study of preradiotherapy chemotherapy followed by hyperfractionated radiotherapy for newly diagnosed high-risk medulloblastoma/ primitive neuroectodermal tumor: a report from the Children's Oncology Group (CCG 9931). Int J Radiat Oncol Biol Phys. 2009;74:1006–11.
- [67] Taylor RE, Bailey CC, Robinson KJ, Weston CL, Walker DA, Ellison D, et al. Outcome for patients with metastatic (M2-3) medulloblastoma treated with SIOP/UKCCSG PNET-3 chemotherapy. Eur J Cancer. 2005;41:727–34.
- [68] Sexauer CL, Khan A, Burger PC, Krischer JP, van Eys J, Vats T, et al. Cisplatin in recurrent pediatric brain tumors. A POG phase II study. A Pediatric Oncology Group Study. Cancer. 1985;56:1497– 501.
- [69] Gajjar A, Chintagumpala M, Ashley D, Kellie S, Kun LE, Merchant TE, et al. Risk-adapted craniospinal radiotherapy followed by high-dose chemotherapy and stem-cell rescue in children with newly diagnosed medulloblastoma (St Jude Medulloblastoma-96): long-term results from a prospective, multicentre trial. Lancet Oncol. 2006;7:813–20.
- [70] Valteau-Couanet D, Fillipini B, Benhamou E, Grill J, Kalifa C, Couanet D, et al. High-dose busulfan and thiotepa followed by autologous stem cell transplantation (ASCT) in previously irradiated medulloblastoma patients: high toxicity and lack of efficacy. Bone Marrow Transplant. 2005;36:939–45.
- [71] Nazemi KJ, Shen V, Finlay JL, Boyett J, Kocak M, Lafond D, et al. High incidence of veno-occlusive disease with myeloablative chemotherapy following craniospinal irradiation in children with newly diagnosed high-risk CNS embryonal tumors: a report from the Children's Oncology Group (CCG-99702). Pediatr Blood Cancer. 2016;63:1563–70.
- [72] Argyriou AA, Kyritsis AP, Makatsoris T, Kalofonos HP. Chemotherapy-induced peripheral neuropathy in adults: a

comprehensive update of the literature. Cancer Manag Res. 2014;6:135-47.

- [73] Packer RJ, Zhou T, Holmes E, Vezina G, Gajjar A. Survival and secondary tumors in children with medulloblastoma receiving radiotherapy and adjuvant chemotherapy: results of Children's Oncology Group trial A9961. Neuro-Oncology. 2013;15:97–103.
- [74] Dhall G, Grodman H, Jo L, Sands S, Gardner S, et al. Outcome of children less than three years old at diagnosis with non-metastatic medulloblastoma treated with chemotherapy on the "Head Start" I and II protocols. Pediatr Blood Cancer. 2008;50:1169–75.
- [75] Dhall G, Ji L, Haley K, Grimm JP, Gilles FH, et al. Outcome of infants and young children with newly diagnosed medulloblastoma treated on Head Start III protocol. J Clin Oncol. 2011;29(15 suppl):2011.
- [76] Rutkowski S, Bode U, Deinlein F, Ottensmeier H, Warmuth-Metz M, Soerensen N, et al. Treatment of early childhood medulloblastoma by postoperative chemotherapy alone. N Engl J Med. 2005;352:978–86.
- [77] Grundy RG, Wilne SH, Robinson KJ, Ironside JW, Cox T, Chong WK, et al. Primary postoperative chemotherapy without radiotherapy for treatment of brain tumours other than ependymoma in children under 3 years: results of the first UKCCSG/SIOP CNS 9204 trial. Eur J Cancer. 2010;46:120–33.
- [78] von Bueren AO, von Hoff K, Pietsch T, Gerber NU, Warmuth-Metz M, Deinlein F, et al. Treatment of young children with localized medulloblastoma by chemotherapy alone: results of the prospective, multicenter trial HIT 2000 confirming the prognostic impact of histology. Neuro-Oncology. 2011;13:669–79.
- [79] Merchant TE, Pollack IF, Loeffler JS, et al. Brain tumors across the age spectrum: biology, therapy, and late effects. Semin Radiat Oncol. 2010;20:58–66.
- [80] Friedrich C, von Bueren AO, von Hoff K, Kwiecien R, Pietsch T, Warmuth-Metz M, et al. Treatment of adult nonmetastatic medulloblastoma patients according to the pediatric HIT 2000 protocol: a prospective observational multicentre study. Eur J Cancer. 2013;49:893–903.
- [81] Taylor RE, Bailey CC, Robinson K, Weston CL, Ellison D, Ironside J, et al. Results of a randomized study of preradiation chemotherapy versus radiotherapy alone for nonmetastatic medulloblastoma: the International Society of Paediatric Oncology/ United Kingdom Children's Cancer Study Group PNET-3 Study. J Clin Oncol. 2003;21:1581–91.
- [82] Beier D, Proescholdt M, Reinert C, Pietsch T, Jones DTW, Pfister SM, et al. Multicenter pilot study of radiochemotherapy as firstline treatment for adults with medulloblastoma (NOA-07). Neuro-Oncology. 2018;20:400–10.
- [83] Pizer BL, Clifford SC. The potential impact of tumor biology on improved clinical practice for medulloblastoma: progress towards biologically driven clinical trials. Br J Neurosurg. 2009;23:364– 75.
- [84] Pizer B, Donachie PH, Robinson K, Taylor RE, Michalski A, et al. Treatment of recurrent central nervous system primitive neuroectodermal tumours in children and adolescents: results of a Children's Cancer and Leukemia Group study. Eur J Cancer. 2011;47:1389–97.
- [85] Gruber ML, Buster WP. Temozolomide in combination with irinotecan for treatment of recurrent malignant glioma. Am J Clin Oncol. 2004;27:33–8.
- [86] Grill J, Geoerger B, Gesner L, Perek D, Leblond P, Canete A, et al. Phase II study of irinotecan in combination with temozolomide (TEMIRI) in children with recurrent or refractory medulloblastoma: a joint ITCC and SIOPE brain tumor study. Neuro-Oncology. 2013;15:1236–43.
- [87] Levy A, Krailo M, Chi S, Williams-Hughes C, Bancroft M, et al. Temozolomide with irinotecan versus temozolomide, irinotecan plus bevacizumab for recurrent medulloblastoma/CNS PNET of

- [88] Phoenix TN, Patmore DM, Boop S, Boulos N, Jacus MO, Patel YT, et al. Medulloblastoma genotype dictates blood brain barrier phenotype. Cancer Cell. 2016;29:508–22.
- [89] Sharpe HJ, Pau G, Dijkgraaf GJ, Basset-Seguin N, Modrusan Z, Januario T, et al. Genomic analysis of Smoothened inhibitor resistance in basal cell carcinoma. Cancer Cell. 2015;27:327–41.
- [90] Pei Y, Moore CE, Wang J, Tewari AK, Eroshkin A, Cho YJ, et al. An animal model of MYC-driven medulloblastoma. Cancer Cell. 2012;21:155–67.
- [91] Hanaford AR, Archer TC, Price A, Kahlert UD, Maciaczyk J, Nikkhah G, et al. DiSCoVERing innovative therapies for rare tumors: combining genetically accurate disease models with in silico analysis to identify novel therapeutic targets. Clin Cancer Res. 2016;22:3903–14.
- [92] Delmore JE, Issa GC, Lemieux ME, Rahl PB, Shi J, Jacobs HM, et al. BET bromodomain inhibition as a therapeutic strategy to target c-Myc. Cell. 2011;146:904–17.
- [93] Perrinjaquet C, Desbaillets N, Hottinger AF. Neurotoxicity associated with cancer immunotherapy: immune checkpoint inhibitors and chimeric antigen receptor T-cell therapy. Curr Opin Neurol. 2019;32:500–10.
- [94] Kramer K, Pandit-Taskar N, Zanzonico P, Wolden SL, Humm JL, DeSelm C, et al. Low incidence of radionecrosis in children treated

with conventional radiation therapy and intrathecal radioimmunotherapy. J Neuro-Oncol. 2015;123:245–9.

- [95] Kramer K, Pandit-Taskar N, Donzelli M, Wolden SL, Zanzonico P, et al. Safety and efficacy of intraventricular 1311-labeled monoclonal antibody 8H9 targeting the surface glycoprotein B7-H3. Pediatr Blood Cancer. 2019;66(S4):V557 SIOP19–1597.
- [96] Marelli G, Howells A, Lemoine NR, Wang Y. Oncolytic viral therapy and the immune system: a double-edged sword against cancer. Front Immunol. 2018;9:866.
- [97] Batich KA, Reap EA, Archer GE, Sanchez-Perez L, Nair SK, Schmittling RJ, et al. Long-term survival in glioblastoma with cytomegalovirus pp65-targeted vaccination. Clin Cancer Res. 2017;23:1898–909.
- [98] Ahmed N, Ratnayake M, Savolodo B, Perlaky L, Dotti G, et al. Regression of experimental medulloblastoma following transfer of HER2-specific T cells. Cancer Res. 2007;67:5957–64.
- [99] Fares J, Fares MY, Fares Y. Natural killer cells in the brain tumor microenvironment: defining a new era in neuro-oncology. Surg Neurol Int. 2019;10:43.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.