

Saber Tadros, MD[∗](#page-0-0) Aleksei Kondrashov, M[D‡](#page-0-1) [§](#page-0-2) Sriya Namagiri, BA[‡](#page-0-1) Ashis Chowdhury, BHM[S‡](#page-0-1) Yeshavanth Kumar Banasavadi-Siddegowda, Ph[D‡](#page-0-1)

Abhik Ray-Chaudhury, M[D‡](#page-0-1)

∗Laboratory of Pathology, National Cancer Institute, National Institutes of Health, Bethesda, Maryland, USA; ‡National Institute of Neurological Disorders and Stroke, National Institutes of Health, Bethesda, Maryland, USA; §Faculty of Medicine, Moscow State University, Moscow, Russia

Correspondence:

Saber Tadros, MD, Laboratory of Pathology, Center for Cancer Research, National Cancer Institute, 10 Center Dr, Building 10, Room 3N248, Bethesda, MD 20814, USA. Email: tadross2@nih.gov

Received, June 7, 2020. **Accepted,** December 13, 2020.

^C Congress of Neurological Surgeons 2021. All rights reserved. For permissions, please e-mail: [journals.permissions@oup.com](file:journals.permissions@oup.com)

Pathological Features of Tumors of the Nervous System in Hereditary Cancer Predisposition Syndromes: A Review

Hereditary cancer predisposition syndromes (HCS) become more recognizable as the knowledge about them expands, and genetic testing becomes more affordable. In this review, we discussed the known HCS that predispose to central and peripheral nervous system tumors. Different genetic phenomena were highlighted, and the important cellular biological alterations were summarized. Genetic mosaicism and germline mutations are features of HCS, and recently, they were described in normal population and as modifiers for the genetic landscape of sporadic tumors. Description of the tumors arising in these conditions was augmented by representative cases explaining the main pathological findings. Clinical spectrum of the syndromes and diagnostic criteria were tabled to outline their role in defining these disorders. Interestingly, precision medicine has found its way to help these groups of patients by offering targeted preventive measures. Understanding the signaling pathway alteration of mammalian target of rapamycin (mTOR) in tuberous sclerosis helped introducing mTOR inhibitors as a prophylactic treatment in these patients. More research to define the germline genetic alterations and resulting cellular signaling perturbations is needed for effective risk-reducing interventions beyond prophylactic surgeries.

KEY WORDS: Hereditary, Germline, Mosaicism, Neurofibromatosis, NF1, NF2, P53, RB1, MEN1, PTEN, mTOR, VHL, ATM

Neurosurgery 0:1–20, 2021 DOI:10.1093/neuros/nyab019 www.neurosurgery-online.com

Hereditary cancer predisposition syn-
drome (HCS) is a condition of increasing risk of developing cancer resulting
from inherited gene mutation. The first decdrome (HCS) is a condition of increafrom inherited gene mutation. The first described germline genetic variants predisposing individuals to cancer was *BRCA1.*[1](#page-18-0) Currently, exome/whole genome sequencing allows novel genes discovery in these disorders. Many exclusive solo cancer susceptibility genes have been identified; however, epidemiological studies elucidated that susceptibility to specific tumors could be attributed to mutations in group of different genes. Conversely, some independent syndromes appeared to be manifestations of different mutations in the same gene. Development of each syndrome requires a loss of tumor-suppressor proteins – hypothesis described by $Knudson²$ $Knudson²$ $Knudson²$ in retinoblastoma. Knudson's[2](#page-18-1) "two-hit" hypothesis postulates one allele loss in germline followed by second

ABBREVIATIONS: AD, autosomal dominant; **AR,** Autosomal recessive; **ASA,** acetylsalicylic acid; **AT,** ataxiatelangiectasia;**ATM,**AT mutated;**AT/RT,** atypical teratoid/rhabdoid tumor;**CMMRD,** constitutional MMR deficiency syndrome; **CNC,** Carney complex; **CNS,** central nervous system; **CPC,** choroid plexus carcinoma; **CS,** Cowden syndrome; **DNT,** dysembryoplastic neuroepithelial tumor; **GEMM,** genetically engineered mouse model; **GFAP,** Glial fibrillary acidic protein; **HAT,** histone acetyltransferase; **HCS,** hereditary cancer predisposition syndrome; **H&E,** Hematoxylin and eosin stain; **HIF,** hypoxia-inducible factor; **IHC,** Immunohistochemistry; **LFS,** Li-Fraumeni syndrome; **LS,** Lynch syndrome; **MMR,** mismatch repair; **mTOR,** mammalian target of rapamycin; **MYC,** myelocytomatosis oncogene; **NBS,** Nijmegen breakage syndrome; **OPG,** optic pathway glioma; **PA,** pituitary adenomas; **PHTS,** PTEN hamartoma tumor syndrome; **RB,** Retinoblastoma; **RTPS,** rhabdoid tumor predisposition syndrome; **SEGA,** subependymal giant cell astrocytoma; **SGBS,** Simpson-Golabi-Behmel syndrome; **SHH,** sonic hedgehog; **SMO,** the signal transducer Smoothened; **TSC,** tuberous sclerosis complex; **VCP,** valosin-containing protein; **VHL,** Von Hippel-Lindau; **WHO,** World Health Organization

allele loss causes multiple tumors. However, the phenomenon of gene dosage sensitivity suggests that haploinsufficiency can explain tumorigenesis.^{[3-](#page-18-2)[5](#page-18-3)}

Germline mosaicism is a feature of HCS. Mosaicism could be somatic, germline, or mixed gonadal/somatic. Naturally, an average of 2:1 asymmetry of early human embryonic cells contributes to adult tissues.^{[6](#page-18-4)} Segmental syndrome/mosaicism results in some cells having 2 normal genes and other cells containing pathogenic variants in 1 copy of the same gene. People with segmental syndrome do not have affected parents. Originally, segmental syndrome is proven by molecular analysis in many cutaneous traits then expanded to hereditary cancer syndrome. Rare individuals were described having only germline mosaicism without apparent somatic features.^{[7,](#page-18-5)[8](#page-18-6)}

Germline mutations are traditionally viewed as HCS hallmark; however, increasing evidence shows a key role of inherited germline genetic variation in nonhereditary cancer risk.⁹ Recent studies suggest that germline variants affect somatic mutation profiles in individuals having sporadic cancer.^{[10](#page-18-8)[,11](#page-18-9)} For example, selective amplification of germline variants like *AGK*, *DGKB*, *EGFR*, *INSR*, *KIT*, and *RELN* are associated with increased glioblastoma risk.¹²

Studying HCS requires experimental models in addition to natural history and genetics studies of affected families. Modeling human cancers was achieved by genetically engineered mouse models (GEMMs) either by the traditional, mosaic, conditional, chimeric, or nongermline GEMMs. Furthermore, induced pluripotent stem cells technology introduces a feasible way of studying HCS.^{[13,](#page-18-11)[14](#page-18-12)}

Central nervous system (CNS) is affected by several HCS either predominantly or as a bystander. Some HCS were previously grouped under the term "Phakomatoses" because of the involvement of the eye lesions; however, it is no longer used because of lack of such lesions in many HCS. Common syndromes that predominantly involve the CNS include neurofibromatosis (NF) types 1 and 2, Von Hippel-Lindau (VHL) disease, and tuberous sclerosis complex (TSC), whereas syndromes having mostly extra-neural manifestations include Cowden, Li-Fraumeni, Turcot, and Gorlin syndromes. There is cellular, molecular, and clinical overlap between NF1, NF2, and schwanommatosis.^{[15](#page-18-13)}

Noteworthy, recent advances in understating HCS improved personalized risk assessment and develop novel interventions to prevent or intercept cancer. For example, precision prevention for *BRCA1*-mutation carriers moved toward receptor activator of $NF- κ B (RANK)$ activation interference.^{[16](#page-18-14)} Further studies are needed to expand our knowledge of germline alterations for developing and implementing effective risk-reducing interventions beyond prophylactic surgical approaches.

NEUROFIBROMATOSIS TYPE 1

NF1 (Tables [1](#page-2-0) and [2\)](#page-6-0) increases risk for neurofibroma, optic gliomas, and malignant peripheral nerve sheath tumors (MPNSTs).

The cutaneous, intraneural, diffuse, and plexiform neurofibromas (Figure [1A-1C](#page-8-0)) are commonly encountered. The plexiform variant involving multiple nerve trunks that appear early first 2 yr of life causes significant physical disfigurement. Plexiform neurofibromas have 10% lifetime risk of progression into MPNST (Figure [1D](#page-8-0) and [1E](#page-8-0)). In localized pediatric MPNST, NF1 is associated with worse survival.

NF1 develop low-grade tumors of the optic nerves, chiasm, tracts, and radiations, termed optic pathway gliomas (OPGs), which can cause vision loss. OPGs (Figure [1F](#page-8-0)) are seen in 15% to 20% of children with NF1. The second common brain tumor is the brainstem glioma, representing 18% of NF1-associated brain neoplasms. Glioblastomas could also be seen.

NF1 is caused by *NF1* germline mutations. Neurofibromin is a cytoplasmic protein that negatively regulates the RAS signaling. The *NF1* mutations are frequently seen in multiple malignancies not typically associated with NF1. Intriguingly, mouse models showed that Nf1+/- microenvironment accelerates benign tumors formation but impairs further progression to malignancy.[17](#page-18-15) In the brain, neurofibromin acts with valosin-containing protein (VCP)/P97 to control endoplasmic reticulum formation and consequent protein synthesis. Additionally, VCP/P97 regulates dendritic spine formation and brain function.^{[18](#page-18-16)}

NF1 gliomas have specific genomic signatures. Pediatric lowgrade gliomas typically exhibit mutations of the MAPK pathway, whereas high-grade gliomas are characterized by loss-of-function mutations in *ATRX*, *TP53*, and *CDKN2A*. Children with NF1 develop diffuse midline gliomas that carry worse prognosis than sporadic cases, independent of histone H3 lysine27-tomethionine (H3K27M) mutation. NF1 gliomas also reduce deoxyribonucleic acid (DNA) methylation of immune genes, which explain tumor-infiltrating lymphocytes.^{[19](#page-18-17)}

NF1 microdeletion syndrome is severe phenotype of NF1 characterized by mental retardation, developmental delay, cardiac anomalies, and dysmorphic features.²⁰ Patients have a 1.4-Mb heterozygous 17q11.2 deletion encompassing *NF1*.

Legius syndrome is an autosomal dominant (AD) disorder that results from loss-of-function *SPRED1* germline mutations and mimics NF1 clinically. It consists of multiple café-au-lait spots, axillary freckling, and macrocephaly but lack neurofibromas, typical osseous lesions, and $OPGs.²¹$ $OPGs.²¹$ $OPGs.²¹$

Melanoma-astrocytoma syndrome (MAS) is a rare condition with cutaneous and neurological manifestations. It is linked to *CDKN2A* and *CDKN2B* germline mutations.

NEUROFIBROMATOSIS TYPE 2

Bilateral vestibular schwannomas are the diagnostic hallmark. Cranial and spinal meningiomas, meningioangiomatosis, spinal ependymoma, and other gliomas are frequently seen. Tumors frequently demonstrate a saltatory growth pattern with unpre-dictable progression.^{[22](#page-18-20)}

NF2-associated schwannomas (Figure [2A](#page-9-0)) appear early in life (third decade) than sporadic cases. Though the vestibular branch of 8th cranial nerve is usually involved, 5th cranial nerve and

Downloaded from https://academic.oup.com/neurosurgery/advance-article/doi/10.1093/neuros/nyab019/6162972 by guest on 22 April 2021 Downloaded from https://academic.oup.com/neurosurgery/advance-article/doi/10.1093/neuros/nyab019/6162972 by guest on 22 April 2021

AD, autosomal dominant; AR, autosomal recessive; GFAP, Glial fibrillary acidic protein; NIH, National institutes of Health.

diagnosis.¹⁰³
IHC, Immunohistochemistry. IHC, Immunohistochemistry.

spinal nerve dorsal roots can be affected. The vestibular tumors in NF2 are polyclonal and more difficult to treat than sporadic tumors because of their multifocal nature.

Plexiform schwannoma (Figure [2B](#page-9-0)) could be associated with NF2 and no malignant transformation is reported. Schwannoma/neurofibroma hybrid nerve sheath tumors have been described in the settings of NF2. Hybrid tumors show highly ordered cellular components (Antoni-A) with loose myxoid components (Antoni-B) in addition to areas of wavy nuclei interspersed with collagen fibers (neurofibroma).

Malignant transformation in schwannoma often has epithelioid appearance but sometimes develops angiosarcoma. Collection of epithelioid cells in schwannoma suggests early malignant transformation.

Multiple meningiomas (Figure [2C-2E](#page-9-0)) (meningiomatosis) occur throughout the cranial and spinal neuraxis and maybe the presenting feature of NF2. There are 2 main hypotheses for their development, one that supports the independent tumorigenesis of these neoplasms and the other suggests the propagation of tumor cells of a unique clone through cerebrospinal fluid.

Gliomas in NF2 consist predominantly of spinal and cauda equina ependymomas (Figure [2F](#page-9-0)). These are usually multiple and slowly growing.

Glial hamartias are clusters of atypical glial cells that occur in the cortices and basal ganglia. These are common and pathognomonic features of NF2.

NF2 results from mutation in the NF2. Merlin exists in 2 forms: inactivated (unphosphorylated) and activated (phosphorylated). Once Merlin gets activated in the presence of WNT or transmembrane proteins (cadherins, integrins, and CD44), inhibits mammalian target of rapamycin (mTOR), is imported to the nucleus and suppresses the Hippo pathway (YAP/TAZ) through binding of the transcription factor CRL4.^{[23](#page-18-21)}

The discovery of LZTR1-associated schwannomatosis showed substantial diagnostic overlap with NF2, particularly in those with unilateral vestibular schwannoma and other nondermal schwannomas.

Schwannomatosis is characterized by multiple peripheral schwannomas in the absence of other diagnostic features of NF2. Bilateral vestibular schwannomas are not encountered and multiple meningiomas could happen. Unlike NF2, somatic mutations of *NF2* not germline mutations are frequently seen resulting from *SMARCB1* or *LZTR1* germline mutations.

LZTR1 inhibits RAS signaling through ubiquitination of RAS by cullin 3 ubiquitin ligase complexes. *LZTR1* mutations result in incomplete degradation of Ras-GTPase RIT1 and dysregulated growth factor signaling responses.²⁴ LZTR1 loss in Schwann cells drives dedifferentiation and proliferation.^{[25](#page-18-23)}

FAMILIAL MULTIPLE MENINGIOMAS

It is an AD disorder defined by the presence of at least 2 lesions that appear simultaneously or at different intracranial locations, without the association of NF. Heterozygous loss-offunction germline mutations in *SMARCE1* – SWI/SNF complex $-$ predisposes to spinal and intracranial clear-cell meningiomas.²⁶ Swedish study showed concordance in histology for meningioma in mother-offspring.²⁷

CARNEY COMPLEX

It is associated with clinically aggressive psammomatous melanotic schwannoma (Figure [3\)](#page-10-0).

Carney complex (CNC) is linked in 40% of familial cases to mutations in tumor-suppressor gene *PRKAR1*. Protein kinase alpha is a ubiquitous cAMP-dependent kinase that binds to Akinase anchoring proteins and allows cAMP-responsive events to occur within specific compartments of the cells and to cluster with other classes of signaling enzymes.^{[28](#page-18-26)} Missense mutation in *PRKAR1* results in decrease in the cellular pool of the regulatory subunits ($RI\alpha$) allowing the catalytic (C) subunit to roam unregulated increasing the kinase activity.

CNC was previously called NAME (nevi, atrial myxoma, ephelides) and LAMB (lentigines, atrial myxoma, blue nevi) syndrome. Because of the presence of ephelides (freckles), it may seem like Peutz-Jeughers syndrome. Carney triad consists of paragangliomas, gastric stromal tumors, and pulmonary chondromas, and should not be confused with CNC.

FIGURE 3. *Tumors of CNC (H&E staining). Psammomatous melanotic schwannoma in 10% of patients, a rare nerve sheath tumor that can involve the CNS and is characterized by high melanin pigmentation and psammoma bodies.*

TUBEROUS SCLEROSIS COMPLEX

CNS manifestations include tubers, subcortical glioneuronal nodules, subependymal glial nodules, and subependymal giant cell astrocytoma (SEGA).

SEGA (Figure [4A](#page-11-0)) occurs in 5% to 15% of TSC patients and is considered a major diagnostic criterion. The tumors are well circumscribed and project in the lateral ventricle. SEGA could present with intratumoral hemorrhage and acute hydrocephalus.

Cortical tubers (Figure [4B](#page-11-0)) are hamartomatous nodules that can be epileptogenic and are usually needed to be resected. Magnetic resonance imaging helps to identify them, and reflectance spectroscopy was attempted in small cohort for better localization.^{[29](#page-18-27)}

TSC is caused by inactivating *TSC1* and *TSC2* germline mutations. Tuberin and hamartin dimerize to suppress mTOR; that is why everolimus is used in TSC.^{[30](#page-18-28)} Tsc1 stabilizes Tsc2 by facilitating Hsp90-mediated folding of kinase and nonkinase clients, including Tsc2, thereby preventing their degradation. 31

Conversely, Polo-like kinase 1 phosphorylation of TSC1 interferes with TSC1/TSC2 binding, destabilized TSC1, promoted dissociation of the TSC complex from the lysosome, and eventually leads to mTOR activation.^{[32](#page-18-30)} Sustained activation of mTOR in oligodendrocyte progenitors in *Tsc1* mutants leads to oligodendrocyte cell death and myelination defects.³³ Hyperactive mTOR-mediated negative feedback regulation of AKT partially contributes to the benign nature of TSC-associated tumors.^{[34](#page-18-32)}

VON HIPPEL-LINDAU DISEASE

VHL syndrome is divided into 4 subtypes (1, 2A, 2B, and 2C) according to the risk of developing pheochromocytoma (type 2C has only pheochromocytomas). Type 1 is more likely to develop CNS hemangioblastomas.

CNS or retinal hemangioblastomas (Figure [5\)](#page-11-1) appear at early age (25-29 yr). CNS tumors developed in cerebellum, brain stem,

FIGURE 4. *Lesions in TSC (H&E staining).* **A***, SEGA: lesion consists of large atypical astrocytes that have copious eosinophilic cytoplasm resembling gemistocytic astrocytes and are arranged in sheets, fascicles, and perivascular clusters (blue arrow). Stromal dystrophic calcification (blue circle) is common. Mitotic figures are rare. It is a slow growing neoplasm (WHO grade 1) arising from the subependymal layer of the lateral ventricles (black arrow points to ependymal lining of lateral ventricle).* **B***, Cortical tubers: it is a hamartomatous nodule that can be multiple and epileptogenic. Large balloon cells displaying both glial and neuronal features are frequently encountered (blue arrow). Typically, lesion is composed of multinucleated giant cells, abnormal dysplastic neurons, and reactive gliosis, disrupting the normal cortical lamination (not shown here). Black arrow shows Virchow-Robin space.*

composed of 2 elements, large number of capillaries and small blood vessels, and collection of stromal cells with eosinophilic cytoplasm. Many stromal cells display larger, lipidized, and vacuolated cytoplasm. The cell of origin is not known. These are benign circumscribed tumors and belong to WHO grade 1.

and spinal cord, in that order of frequency. Multifocal CNS hemangioblastomas were also reported.

VHL is associated with inactivation *VHL* germline mutations. VHL is a tumor-suppressor protein involved in hypoxia signaling. It acts as ubiquitin E3 ligase to help the proteasomal degradation of hypoxia-inducible factors 1 and 2. These factors activate glycolysis, fatty acid metabolism, erythropoiesis, and angiogenesis. VHL is also required for proper P53 activation and mTOR regulation.

VHL-JAK-STAT signaling plays alternative pathways in hemangioblastoma to supplement VHL-hypoxia-inducible factor (HIF) pathway. 35 Jade-1 is another tumor-suppressor protein associated with histone acetyltransferase (HAT) activity and stabilized by VHL; its destabilization contributes to cancer.³⁶

Independent of HIF pathway, inactivation of $m⁶A$ ribonucleic acid (RNA) demethylase Fat mass and obesity-associated protein

in the presence of VHL loss reduces tumor cells growth. It was proposed as targeted therapy for drug-resistant clear-cell renal cell carcinoma in VHL.^{[37](#page-18-35)}

Although VHL could be caused by multiple loss-of-function mutations, C-terminal-encoding *VHL* mutations may cause polycythemia.[38](#page-18-36)

LI-FRAUMENI SYNDROME

Among the CNS neoplasms medulloblastomas, choroid plexus carcinoma (CPC) and ependymomas predominate. However, in the young adult, astrocytomas occur commonly.

Most CPC arise in the children (14% occurring in the first year of life) in the region of the lateral ventricles (Figure [6A](#page-12-0)). Hydrocephalus and increased intracranial pressure are common because of tumors location.

Although 40% of CPCs display *TP53* germline mutations in the setting of Li-Fraumeni syndrome (LFS), 90% of *TP53* wildtype CPC harbor some dysfunction of P53 because of polymorphism like R72P variant.

Medulloblastoma (Figure [6B](#page-12-0) and [6C](#page-12-0)) predominantly affects children less than 3 yr old. They are located exclusively in the cerebellum and dorsal brainstem region. Originally, 4 histological subtypes were recognized, including classic (72% of all), desmoplastic/nodular, medulloblastoma with extensive nodularity, and large cell/anaplastic. However, World Health Organization (WHO) classification is based on their molecular characteristics because of its increasing clinical utility. Five distinct genetic subtypes are identified: WNT-activated, SHHactivated, *TP53*-mutant, *TP53* wild-type, and non-WNT/non-SHH. Massive chromosome rearrangements in a 1-step catastrophic event termed "chromothripsis" were linked to P53 status.^{[39](#page-18-37)}

LFS results from *TP53* germline mutations. The core P53 pathway is the response to DNA damage either by repair

and return to homeostasis or cell death. There is redundancy and extensive communication of P53 pathway with other cellular pathways and feedback loops. The downstream genes regulated by the wild-type P53 are robust; however, lossof-function mutation of *TP53* turns off the entire pathway. Interestingly, mutations in *TP53* can function efficiently initializing malignancy rapidly in tissue of ectodermal or mesodermal-derived origin (in case of medulloblastoma and rhabdomyosarcoma), whereas same mutations in endodermalderived stem cells require other mutations in oncogenes or tumor-suppressor genes, occurring prior to the *TP53* mutations, to develop malignancy (in case of lung and colonic cancer).^{[40](#page-18-38)}

Surprisingly, P53-mutant loss of function is accompanied by 3 phenomena. First, P53 mutant can drive P53 wildtype to a mutant conformation in similar mechanism to the prions.[41](#page-18-39) Second, P53 mutant has the ability of acquiring novel functions[.42](#page-18-40) Last, mutations within *TP53* cause stabilization of P53-mutant and its overexpression that led to its discovery origi-nally.^{[43](#page-18-41)} Among the novel functions, P53 mutant significantly upregulates mevalonate pathway via SREBP2 and promotes the synthesis of ubiquinone that supports the synthesis of pyrimidine nucleotide[.44](#page-18-42)

PTEN HAMARTOMA TUMOR SYNDROMES

The defining feature of PTEN hamartoma tumor syndrome (PHTS) is the presence of hamartomatous tumors.

COWDEN SYNDROME

Multiple hamartomas arising from all three germ layers happens, of which facial trichilemmomas and adult cerebellar dysplastic gangliocytomas are highly pathognomonic.

Cowden syndrome (CS) is caused by *PTEN* germline mutations; however, somatic mutations are rarely encountered in CNS malignancies. Generally, PTEN levels are frequently downregulated in cancer, even in the absence of genetic aberrations.⁴⁵ E3 ubiquitin ligase WWP1 negatively regulates PTEN, and it was suggested to have oncogenic function.^{[46](#page-18-44)} PTEN tumorsuppressor activity depends on its lipid phosphatase activity, which antagonizes PI3K-AKT-mTOR signaling at the plasma membrane. Additionally, PTEN has phosphatase independent functions in the nucleus like DNA repair regulation and maintaining chromosomal stability and can be exported extracellularly through exosomal export or secretion to produce tumor-suppressor effects in adjacent cells.^{[47](#page-18-45)}

CS and Bannayan-Riley-Ruvalcaba syndrome are considered variable phenotypic presentations of the same disorder.

Dysplastic Cerebellar Gangliocytomas

They are benign cerebellar tumors composed of dysplastic and enlarged ganglionic cells infiltrating and enlarging the internal granular layer of the cerebellar gray matter, causing abnormal thickening of cerebellar folia. Though WHO grade 1 has been assigned, it is not clear whether this is a neoplasm or hamartoma. The tumor may spread locally but not outside the cerebellum. Most cases occur in the adults and all of them display *PTEN* mutations, which are not common in pediatric tumors. It may arise in conjunction with CS or without other PHTS signs.

Proteus-like syndrome is associated with *PTEN* pathogenic variants but lack hamartomas.

TURCOT SYNDROME

Turcot syndrome is a historical term for conditions showing brain tumors, intestinal polyps, and cancers caused by *APC* germline mutations. Recently, it was divided subsequently into 2 distinct disorders based on different genetic profiles involving distinct inheritance and cancer spectrum.

Mismatch Repair Cancer Syndrome

This AD disorder is caused by mutations in one of the mismatch repair (MMR) genes *MLH1*, *PMS2*, *MSH2*, and *MSH6*. Individuals carry biallelic homozygous or compound heterozygous deleterious germline mutations in MMR, leading to constitutional MMR deficiency syndrome (CMMRD) in contrast to heterozygous monoallelic germline loss-of-function mutations followed by somatic loss of the remaining wild-type allele, which happens in Lynch syndrome (LS).

Based on the effect of acetylsalicylic acid (ASA) in reducing cancer risk in individuals with LS, ASA is used to reduce the risk of cancer in CMMRD.^{[48](#page-18-46)}

Astrocytomas and oligodendrogliomas (Figure [7\)](#page-13-0) occur in the first 2 decades. Presence of giant cells on histology, multiple brain

tumors, and developmental brain anomalies are suggested to be characteristic features for CMMRD. In addition, medulloblastomas and primitive neuroectodermal tumors were reported.

Individuals with CMMRD may have NF1-like phenotype, presenting with café-au-lait spots, neurofibromas, Lisch nodules, and axillary freckling. Gastrointestinal polyposis and malignancies are encountered in almost all the patients.

Familial Adenomatous Polyposis

It is caused by heterozygous mutations in the *APC* tumorsuppressor gene. APC inhibits the WNT/ β -catenin signaling. Without APC, β-catenin remains undegraded, resulting in uncontrolled proliferation. β -catenin is also involved in cell migration, adhesion, transcriptional activation, and apoptosis inhibition.

Polymerase proofreading-associated polyposis syndrome is AD highly penetrant disorder caused by *POLE* and *POLD1* germline mutations. Affected individuals develop adenomatous polyposis and are at risk for colonic and endometrial cancer at early age.

NEVOID BASAL CELL CARINOMA SYNDROME (GORLIN SYNDROME)

The nevoid basal cell carcinoma syndrome is caused by germline mutations of *PTCH1* and rarely *SUFU* or *PTCH2*.

PTCH1 is a receptor for secreted hedgehog signaling molecules, including sonic hedgehog (SHH).⁴⁹ SHH binds to and inhibits PTCH1, which permits the signal trans-ducer Smoothened (SMO) activation.^{[50](#page-18-48)} Mutated *PTCH1* does not inhibit SMO, resulting in activation of SUFU-GLI. GLI is a transcription factor that causes cell proliferation.^{[51](#page-18-49)} The risk of developing medulloblastomas is substantially higher in individuals with *SUFU* pathogenic variants with a male predominance of 3:1.

Gorlin-Koutlas syndrome is defined by occurring of multiple schwannomas in an extended family who also develop multiple nevocytic nevi and vaginal leiomyomas.

RHABDOID TUMOR PREDISPOSITION SYNDROME

Rhabdoid tumor predisposition syndrome (RTPS) is caused by *SMARCB1* germline mutations. SMARCB1/INI1 is part of the BAF chromatin-remodeling complex (aka SWI/SNF complex). Knock-out of Ini1 in mice embryo results in their demise, 52 but Ini1-heterozygous mice develop aggressive cancer, including rhabdoid-like tumors and T-cell lymphomas.^{[53](#page-19-13)} Different mouse models showed that early Ini1 loss causes rhabdoid tumors, whereas Ini1 loss at later stages combined with *Nf2* inactivation causes schwannomas. Atypical teratoid/rhabdoid tumor (AT/RT) (Figure [8\)](#page-14-0) presents in patients aged 3 yr or below. Both supra and infratentorial locations are common.

FIGURE 8. *AT/RT (H&E staining). This is a poorly differentiated WHO grade 4 embryonal tumor that usually presents in patients aged 3 yr or below. Both supra and infratentorial locations are common. The characteristic feature of the tumor is presence of large number of cells having copious eosinophilic cytoplasm and eccentrically located nuclei, so-called rhabdoid cells (black arrow on the right). Apart from this the neoplastic cells can display epithelial, mesodermal (blue arrow on the left), and neuroectodermal differentiation.*

HEREDITARY RETINOBLASTOMA

Hereditary retinoblastoma is caused by *RB1* germline mutations and 90% of cases present before 3 yr of age. Trilateral retinoblastoma presents as midline intracranial neuroblastoma (in the pineal gland or in the supra/parasellar region) with bilateral intraocular retinoblastomas (Figure [9\)](#page-14-1)[.](#page-14-2) Noninheritable cases of retinoblastoma are always unilateral.

Retinoblastoma (RB) inhibits cell growth through interaction with E2F transcription factors.^{[54](#page-19-14)} The CDK–RB–E2F axis forms the core transcriptional machinery driving cell cycle progression and alterations in the components of this axis occur in virtually all cancers. Interestingly, mice with single copy of *Rb* develop multiple neuroendocrine neoplasia[.55](#page-19-15) Moreover, most sporadic cancers inactivate RB by phosphorylation rather than losing it entirely – possibly to take advantage of RB antiapoptotic role under stress.⁵⁶

Inactivation of *RB1* is usually caused by mutations affecting the coding region. Silencing by methylation of *RB1* promoter is observed in retinoblastoma as second event and is classified

FIGURE 9. *Retinoblastoma.* **A***, Tumor shows endophytic growth inward toward vitreous cavity and fill it.* **B***, Tumor consists of small round blue cells.*

FIGURE 10. Pituitary adenoma in MEN1 (H&E staining). The adenomas display proliferation of monomorphic epithelioid cells having copious cytoplasm. *The cells are arranged in sheets and cords* **A***, destroying the typical acinar (nested) architecture of the normal adenohypophysis. The effacement of the normal acinar network of the gland can be confirmed by a reticulin stain* **B** *that is diagnostic of an adenoma. No acinar outlines could be appreciated on reticulin staining.*

as somatic epimutation. However, monoallelic germline *RB1* promoter methylation has also been described.⁵⁷ RB1 itself is known epigenetic regulator interacting wit[h](#page-15-0) SWI/SNF complexes, histone deacetylase, and DNA methyltransferase DNMT_{1.[58](#page-19-18)}

ATAXIA-TELANGIECTASIA

A total of 85% of patients develop lymphomas and acute leukemias, but brain tumors may happen. Ataxia-telangiectasia (AT) is caused by AT mutated (*ATM*) germline mutations. ATM kinase regulates the cellular response to DNA double-strands breaks by activating P53 and inhibiting MDM2 (P53-specific inactivator), leading to P53 accumulation.⁵⁹ It also activates other DNA repair proteins (eg, BRCA1 and NBS1), CHK2 (control cell cycle), eIF-4E (protein translation), and PP2A (AKT phosphatase).^{60-[62](#page-19-21)} The presence of neurological symptoms likely develops because of PP2A inhibition.⁶³ In the absence of ATM, cells build up somatic mutations, leading to malignancies.

MULTIPLE ENDOCRINE NEOPLASIA TYPE 1 (MEN1)

It is characterized by parathyroid, pancreatic islet cell/gastrointestinal, and anterior pituitary tumors. Multiple endocrine neoplasia type 1 (MEN1) results from *MEN1* germline mutations. Menin is a component of MLL1/MLL2-containing histone methyltransferase complexes that trimethylate H3K4.^{[64](#page-19-23)} However, it interacts with myelocytomatosis oncogene (MYC) and enhances the transcription of MYC target genes independent from H3K4me3 inhibitory activity.⁶⁵ Menin inhibits SHH⁶⁶ and HOX signaling via PRMT5. It interacts with CHES1 in S-phase checkpoint pathway related to DNA damage response.^{[67](#page-19-26)} It has essential role in WNT/ β -catenin signaling.^{[68](#page-19-27)} Additionally, it binds transcription factor JUND, inhibiting its transcriptional activity.⁶⁹ It remains unclear why tumors arise only in neuroendocrine organs.

Pituitary adenomas (PA) (Figure [10\)](#page-14-2) are broadly categorized into macro- or micro-adenomas (1-cm cutoff), functional (hormone-secreting), or nonfunctioning. Functional adenomas secrete 1 hormone (commonly GH or ACTH), though some may secrete 2 hormones. These functiona[l](#page-16-0) adenomas are microadenomas and are detected early because of early hyperpituitarism symptoms. Nonfunctioning adenomas are usually macroadenomas, causing hypopituitarism by compressing the adjacent normal parenchyma.

MEN4

MEN4 is caused by CDKN1B germline mutations and characterized by parathyroid and anterior pituitary tumors in association with tumors of the adrenals, kidneys, and reproductive organs. 70

FIGURE 12. *Diagram illustrates signaling pathways involved in hereditary predisposition cancer syndromes pathogenesis. Many HCS genetic alterations affect cell cycling proteins (LFS, RB, and MAS), DNA repair (ATM, CMMRD, and PPAP), mTOR pathway (NF1, NF2, TSC, VHL, and Cowden), or epigenetic modifications (RTS, RTPS, Schwannomatosis, and MEN1). "*→*," stimulation; and " " inhibition. Color code: "warm" colors (yellow, orange, and peach) represent proto-oncogenic proteins, whereas "cold" colors (blue, magenta, violet, and green) represent tumor suppressors. Red patches – names of the diseases linked with the associated abnormal proteins. Golden yellow – Ras-Raf-MEK-ERK (also known as MAPK/ERK) pathway, one of the main growth signaling pathways. Yellow-green – hypoxia-inducible factors sustain growth and development in an oxygen-deprived environment. Orange – PI3K-AKT (also known as protein kinase B) pathway, the second major cellular growth, and proliferation signaling cascade. AKT phosphorylates more than a hundred of other proteins and is highly conservative throughout all eukaryotic organisms. The RAS pathway also activates it. Peach – protein kinase A/cAMP pathway; regulated by G proteins (Gs activates, Gi inhibits); involved in glucose metabolism, cellular growth, and proliferation (via CREB). Lime – proteins involved in cytoskeleton regulation and intercellular contacts (WNT/B-catenin pathway, Merlin). WNT pathway is a key regulator of cellular polarity, migration, and body axial orientation; it also controls regeneration in adult bone marrow, skin, and intestine. Merlin is a membrane-cytoskeleton scaffolding protein, normally promotes contact-mediated growth and proliferation inhibition through the Hippo pathway (not shown in a diagram). Green – DNA reparation proteins; includes single- and double-strand breaks repair systems and mismatched base repair system. Mismatch errors occur when DNA in the synthetic (S) phase, whereas others can occur throughout the cell cycle. Double-stranded breaks, as shown, mostly happen because of ultraviolet radiation, which explains sunlight sensitivity in AT. Blue – inhibitory proteins (VHL, NF1, and TSC1/TSC2). These are major negative regulators of growth signaling pathways. This group is heterogeneous by the mechanism of action. Magenta – cell cycle regulatory proteins; includes P53 and RB1 (G1-S checkpoint control). Purple – DNA binding proteins, which include SWI/SNF complexes and RNA polymerases. SWI/SNF complexes, as shown, responsible for proper chromatin packing and nucleosome sliding. Abbreviations: GF – growth factors (such factors as insulin and insulin-like growth factors, epidermal growth factor, nerve growth factor, platelet-derived growth factor, vascular endothelial growth factor, and fibroblast growth factor); UV – ultraviolet light; DSB – double-strand breaks); RTK – receptor tyrosine kinase; MAPK – mitogen-activated protein kinases; MEK – MAPK/ERK kinase; ERK – extracellular signal-regulated kinases; NF1– neurofibromin 1 protein; NF type 1 – neurofibromatosis type 1; Ras – rat sarcoma proteins; RAF – rapidly accelerated fibrosarcoma kinase; cAMP – cyclic adenosine monophosphate; PKA – protein kinase A; CREB – cyclic AMP response element-binding protein; PI3K – phosphoinositide 3-kinase; PTEN – phosphatase and tensin homolog; TSC1/TSC2 – tuberous sclerosis complex proteins 1 and 2; mTOR – mammalian target of rapamycin complex; CD44 (HCAM) – homing cell adhesion molecule; NF type 2 – neurofibromatosis type 2; VHL – von Hippel-Lindau; HIF 1a/1b – hypoxia-inducible factors 1a/1b; LRP5/6 – low-density lipoprotein receptor-related protein 5/6; APC – adenomatous polyposis coli; SWI/SNF – SWItch/sucrose nonfermentable chromatin-remodeling complexes; MLL – mixed lineage leukemia complexes; MEN1 – multiple endocrine neoplasia type 1; ATM – ataxia-telangiectasia mutated; CMMRD – constitutional mismatch repair disorder; EXO1 – exodeoxyribonuclease 1; Shh – sonic hedgehog protein; PTCH1 – protein patched homolog 1; SMO – smoothened; SUFU – suppressor of fused homolog; Gli – glioma associated oncogene; PPAP – polymerase proofreading-associated polyposis; Pole – DNA polymerase epsilon.*

FAMILIAL ISOLATED PITUITARY ADENOMA

It is a familial PA with no other associated tumors like in MEN1, MEN4, and CNC. It is caused by *AIP* germline mutations with incomplete penetrance.^{[71](#page-19-30)} AIP is associated normally with GH and prolactin secretory vesicles, but not in thyrotrophs, corticotrophs, gonadotrophs, or follicular cells.

McCune-Albright syndrome and SDHx mutations are also HCS that predispose to pituitary tumorigenesis.^{72[,73](#page-19-32)}

RUBINSTEIN-TAYBI SYNDROME

Rubinstein-Taybi syndrome (RTS) is associated with medulloblastomas, meningiomas, and oligodendrogliomas.

RTS is associated with mutations in the *CREBBP* or its homolog *EP300*. CBP has HAT activity and regulates expression of multiple proteins, such as P53 and MYC. CREBBP mutations results in tumorigenesis and immuno-evasion[.74-](#page-19-33)[76](#page-19-34)

OLLIER DISEASE AND MAFFUCCI SYNDROME

Individuals with these disorders develop multiple enchondromas with no causative gene identified to date. However, there is increased incidence of gliomas that harbor *IDH1* and rarely *IDH2* mutations[.77](#page-19-35) Notably, *IDH1* and *IDH2* mutations are reported in enchondromas and chondrosarcoma.^{[78](#page-19-36)}

WERNER SYNDROME

Werner syndrome (WS) is premature aging syndrome that develops features of aging in the second decade of life. It is caused by *WRN* RecQ helicase mutations, which is not enough alone to develop tumors.[79](#page-19-37) Lacking *WRN* results in deletion of telomeres from single sister chromatids that are replicated by lagging strand synthesis.⁸⁰ Cancer predisposition is due to cellular senescence and not mtDNA mutagenesis.⁸¹

NIJMEGEN BREAKAGE SYNDROME

Most malignancies in Nijmegen breakage syndrome (NBS) are lymphoid with reported medulloblastomas. NBS is due to *NBS1* mutations that result in genomic instability. Nbs1-deficient neuroprogenitors show proliferation defects and contain more chromosomal breaks, which are accompanied by ATM-mediated P53 activation.⁸²

SIMPSON-GOLABI-BEHMEL SYNDROME

Simpson-Golabi-Behmel syndrome (SGBS) is an overgrowth syndrome caused by *GPC3* germline mutations. Glypican-3 forms a complex with insulin-like growth factor 2.83 2.83 A sellarsuprasellar cyst, dysmorphic pituitary gland, and a cyst of the septum pellucidum are reported.

Beckwith-Wiedemann, Sotos, and Weaver syndromes are also overgrowth disorders that are not connected with GH overproduction.

DICER1 SYNDROME

DICER1 germline mutations cause variety of tumors. Loss of DICER1 in developing lung results in pleuropulmonary blastoma.[84](#page-19-42) CNS manifestations of DICER1 syndrome include pituitary blastoma, pineoblastoma, ciliary body medulloepithelioma, primary DICER1-associated CNS sarcomas, and ETMR-like infantile cerebellar embryonal tumor.^{[85](#page-19-43)}

NOONAN SYNDROME

Noonan syndrome is a RASopathy disorder due to *PTPN11* germline mutations. Brain tumors such as dysembryoplastic neuroepithelial tumor (DNT) (Figure [11A](#page-15-0)), subependymoma (Figure [11B](#page-15-0) and [11C](#page-15-0)), glioneural tumor, and gliomas were reported.^{[86](#page-19-44)}

CONCLUSION

Although most HCS are rare or extremely rare (1:40 000- 1:100 000), NF1 and TSC are quite common (1:3000-1:10 000) with high penetrance. For comparison, the incidence of glioblastoma is 1:33 000 to 1:50 000.

Family history of early-onset brain cancer gives a modest increase (2-3-fold) in the cumulative risk of brain cancer: 0.24% in general population vs 0.75% for offspring and 0.53% for siblings. Similar results were obtained in the Norwegian nationwide register-based cohort 1960 to 2001 study (2.6 million children, 2477 primary solid tumors).[87](#page-19-45) Interestingly, after excluding hereditary cancer syndromes, a family history of cancer still increased the risk of childhood CNS tumors and neuroblastomas 2.3-fold and retinoblastoma 6.1 fold. This suggests that other unknown genetic mechanisms are involved.

Our review presented the pathological and molecular features (Figure [12\)](#page-16-0) of tumors associated with HCS. Using DNA methylation classifier empowers the pathologists with more objective way of diagnosing the tumors; however, stratification of the tumors in syndrome-associated group vs the sporadic group has not been achieved yet. Further studies are needed to identify features that define syndrome-associated tumors to help diagnosing them and developing targeted therapy for them.

Funding

This study did not receive any funding or financial support.

Disclosures

The authors have no personal, financial, or institutional interest in any of the drugs, materials, or devices described in this article.

- 1. Hall JM, Lee MK, Newman B, et al. Linkage of early-onset familial breast cancer to chromosome 17q21. *Science*. 1990;250(4988):1684-1689.
- 2. Knudson AG Jr. Mutation and cancer: statistical study of retinoblastoma. *Proc Natl Acad Sci USA*. 1971;68(4):820-823.
- 3. Katayama S, Suzuki M, Yamaoka A, et al. GATA2 haploinsufficiency accelerates EVI1-driven leukemogenesis. *Blood*. 2017;130(7):908-919.
- 4. Sents W, Meeusen B, Kalev P, et al. PP2A inactivation mediated by PPP2R4 haploinsufficiency promotes cancer development. *Cancer Res*. 2017;77(24):6825- 6837.
- 5. Pemov A, Park C, Reilly KM, Stewart DR. Evidence of perturbations of cell cycle and DNA repair pathways as a consequence of human and murine NF1 haploinsufficiency. *BMC Genomics*. 2010;11(1):1-16.
- 6. Buzulina VP, Popova IA, Vetrova EG, Nosova EA. [Correlations of cardiorespiratory and metabolic reactions in humans with graded physical load]. *Kosm Biol Aviakosm Med*. 1990;24(1):17-20.
- 7. Lazaro C, Ravella A, Gaona A, Volpini V, Estivill X. Neurofibromatosis type 1 due to germ-line mosaicism in a clinically normal father. *N Engl J Med*. 1994;331(21):1403-1407.
- 8. Rose VM, Au KS, Pollom G, Roach ES, Prashner HR, Northrup H. Germline mosaicism in tuberous sclerosis: how common? *Am J Hum Genet*. 1999;64(4):986-992.
- 9. Kar SP, Beesley J, Amin Al Olama A, et al. Genome-wide meta-analyses of breast, ovarian, and prostate cancer association studies identify multiple new susceptibility loci shared by at least two cancer types. *Cancer Discov*. 2016;6(9):1052- 1067.
- 10. Carter H, Marty R, Hofree M, et al. Interaction landscape of inherited polymorphisms with somatic events in cancer. *Cancer Discov*. 2017;7(4):410-423.
- 11. Consortium ITP-CAoWG. Pan-cancer analysis of whole genomes. *Nature*. 2020;578(7793):82-93.
- 12. LaFramboise T, Dewal N, Wilkins K, Pe'er I, Freedman ML. Allelic selection of amplicons in glioblastoma revealed by combining somatic and germline analysis. *PLos Genet*. 2010;6(9):e1001086.
- 13. Lee DF, Su J, Kim HS, et al. Modeling familial cancer with induced pluripotent stem cells. *Cell*. 2015;161(2):240-254.
- 14. Blair JD, Hockemeyer D, Bateup HS. Genetically engineered human cortical spheroid models of tuberous sclerosis. *Nat Med*. 2018;24(10):1568-1578.
- 15. Ferner RE, Bakker A, Elgersma Y, et al. From process to progress-2017 International Conference on Neurofibromatosis 1, Neurofibromatosis 2 and Schwannomatosis. *Am J Med Genet A*. 2019;179(6):1098-1106.
- 16. Nolan E, Vaillant F, Branstetter D, et al. RANK ligand as a potential target for breast cancer prevention in BRCA1-mutation carriers. *Nat Med*. 2016;22(8):933- 939.
- 17. Brosseau JP, Liao CP, Wang Y, et al. NF1 heterozygosity fosters de novo tumorigenesis but impairs malignant transformation. *Nat Commun*. 2018;9(1):5014.
- 18. Shih YT, Huang TN, Hu HT, Yen TL, Hsueh YP. VCP overexpression and leucine supplementation increase protein synthesis and improve fear memory and social interaction of NF1 mutant mice. *Cell Rep*. 2020;31(13):107835.
- 19. D'Angelo F, Ceccarelli M, Tala, et al. The molecular landscape of glioma in patients with neurofibromatosis 1. *Nat Med*. 2019;25(1):176-187.
- 20. Kehrer-Sawatzki H, Kluwe L, Salamon J, et al. Clinical characterization of children and adolescents with NF1 microdeletions. *Childs Nerv Syst*. 2020;36(10):2297-2310.
- 21. Brems H, Chmara M, Sahbatou M, et al. Germline loss-of-function mutations in SPRED1 cause a neurofibromatosis 1-like phenotype. *Nat Genet*. 2007;39(9):1120-1126.
- 22. Dirks MS, Butman JA, Kim HJ, et al. Long-term natural history of neurofibromatosis type 2-associated intracranial tumors. *J Neurosurg*. 2012;117(1):109-117.
- 23. Coy S, Rashid R, Stemmer-Rachamimov A, Santagata S. An update on the CNS manifestations of neurofibromatosis type 2. *Acta Neuropathol*. 2020;139(4):643- 665.
- 24. Castel P, Cheng A, Cuevas-Navarro A, et al. RIT1 oncoproteins escape LZTR1 mediated proteolysis. *Science*. 2019;363(6432):1226-1230.
- 25. Bigenzahn JW, Collu GM, Kartnig F, et al. LZTR1 is a regulator of RAS ubiquitination and signaling. *Science*. 2018;362(6419):1171-1177.
- 26. Smith MJ, O'Sullivan J, Bhaskar SS, et al. Loss-of-function mutations in SMARCE1 cause an inherited disorder of multiple spinal meningiomas. *Nat Genet*. 2013;45(3):295-298.
- HEREDITARY CANCER PREDISPOSITION SYNDROMES
- 27. Babaei M, Fallah M, Sundquist K, Hemminki K. Histological concordance in familial central nervous system tumors: evidence from nationwide Swedish familycancer database. *Cancer Epidemiol*. 2015;39(3):334-339.
- 28. Bauman AL, Soughayer J, Nguyen BT, et al. Dynamic regulation of cAMP synthesis through anchored PKA-adenylyl cyclase V/VI complexes. *Mol Cell*. 2006;23(6):925-931.
- 29. Oh S, Stewart T, Miller I, et al. In vivo optical properties of cortical tubers in children with tuberous sclerosis complex (TSC): a preliminary investigation. *Epilepsia*. 2011;52(9):1699-1704.
- 30. Hong CH, Tu HP, Lin JR, Lee CH. An estimation of the incidence of tuberous sclerosis complex in a nationwide retrospective cohort study (1997-2010). *Br J Dermatol*. 2016;174(6):1282-1289.
- 31. Woodford MR, Sager RA, Marris E, et al. Tumor suppressor TSC1 is a new HSP90 co-chaperone that facilitates folding of kinase and non-kinase clients. *EMBO J*. 2017;36(24):3650-3665.
- 32. Li Z, Kong Y, Song L, et al. Plk1-mediated phosphorylation of TSC1 enhances the efficacy of rapamycin. *Cancer Res*. 2018;78(11):2864-2875.
- 33. Jiang M, Liu L, He X, et al. Regulation of PERK-eIF2alpha signalling by tuberous sclerosis complex-1 controls homoeostasis and survival of myelinating oligodendrocytes. *Nat Commun*. 2016;7(1):1-15.
- 34. Jin F, Jiang K, Ji S, et al. Deficient TSC1/TSC2-complex suppression of SOX9-osteopontin-AKT signalling cascade constrains tumour growth in tuberous sclerosis complex. *Hum Mol Genet*. 2017;26(2):407-419.
- 35. Kanno H, Yoshizumi T, Shinonaga M, Kubo A, Murata H, Yao M. Role of VHL-JAK-STAT signaling pathway in central nervous system hemangioblastoma associated with von Hippel-Lindau disease. *J Neurooncol*. 2020;148(1):29-38.
- 36. Zhou MI, Wang H, Foy RL, Ross JJ, Cohen HT. Tumor suppressor von Hippel-Lindau (VHL) stabilization of jade-1 protein occurs through plant homeodomains and is VHL mutation dependent. *Cancer Res*. 2004;64(4):1278- 1286.
- 37. Xiao Y, Thakkar KN, Zhao H, et al. The m(6)A RNA demethylase FTO is a HIF-independent synthetic lethal partner with the VHL tumor suppressor. *Proc Natl Acad Sci USA*. 2020;117(35):21441-21449.
- 38. Lanikova L, Lorenzo F, Yang C, et al. Novel homozygous VHL mutation in exon 2 is associated with congenital polycythemia but not with cancer. *Blood*. 2013;121(19):3918-3924.
- 39. Rausch T, Jones DT, Zapatka M, et al. Genome sequencing of pediatric medulloblastoma links catastrophic DNA rearrangements with TP53 mutations. *Cell*. 2012;148(1-2):59-71.
- 40. Levine AJ, Jenkins NA, Copeland NG. The roles of initiating truncal mutations in human cancers: the order of mutations and tumor cell type matters. *Cancer Cell*. 2019;35(1):10-15.
- 41. Milner J, Medcalf EA. Cotranslation of activated mutant p53 with wild type drives the wild-type p53 protein into the mutant conformation. *Cell*. 1991;65(5):765- 774.
- 42. Dittmer D, Pati S, Zambetti G, et al. Gain of function mutations in p53. *Nat Genet*. 1993;4(1):42-46.
- 43. Lane DP, Crawford LV. T antigen is bound to a host protein in SV40-transformed cells. *Nature*. 1979;278(5701):261-263.
- 44. Kaymak I, Maier CR, Schmitz W, et al. Mevalonate pathway provides ubiquinone to maintain pyrimidine synthesis and survival in p53-Deficient cancer cells exposed to metabolic stress. *Cancer Res*. 2020;80(2):189-203.
- 45. Salmena L, Carracedo A, Pandolfi PP. Tenets of PTEN tumor suppression. *Cell*. 2008;133(3):403-414.
- 46. Lee YR, Yehia L, Kishikawa T, et al. WWP1 gain-of-function inactivation of PTEN in cancer predisposition. *N Engl J Med*. 2020;382(22):2103-2116.
- 47. Wang X, Trotman LC, Koppie T, et al. NEDD4-1 is a proto-oncogenic ubiquitin ligase for PTEN. *Cell*. 2007;128(1):129-139.
- 48. Leenders E, Westdorp H, Bruggemann RJ, et al. Cancer prevention by aspirin in children with constitutional mismatch repair deficiency (CMMRD). *Eur J Hum Genet*. 2018;26(10):1417-1423.
- 49. Qi X, Schmiege P, Coutavas E, Li X. Two patched molecules engage distinct sites on hedgehog yielding a signaling-competent complex. *Science*. 2018; 362(6410).
- 50. Deshpande I, Liang J, Hedeen D, et al. Smoothened stimulation by membrane sterols drives hedgehog pathway activity. *Nature*. 2019;571(7764):284-288.
- 51. Taylor MD, Liu L, Raffel C, et al. Mutations in SUFU predispose to medulloblastoma. *Nat Genet*. 2002;31(3):306-310.
- 52. Guidi CJ, Sands AT, Zambrowicz BP, et al. Disruption of INI1 leads to peri-implantation lethality and tumorigenesis in mice. *Mol Cell Biol*. 2001;21(10):3598-3603.
- 53. Roberts CW, Galusha SA, McMenamin ME, Fletcher CD, Orkin SH. Haploinsufficiency of SNF5 (integrase interactor 1) predisposes to malignant rhabdoid tumors in mice. *Proc Natl Acad Sci USA*. 2000;97(25):13796-13800.
- 54. Lees JA, Saito M, Vidal M, et al. The retinoblastoma protein binds to a family of E2F transcription factors. *Mol Cell Biol*. 1993;13(12):7813-7825.
- 55. Nikitin AY, Juarez-Perez MI, Li S, Huang L, Lee WH. RB-mediated suppression of spontaneous multiple neuroendocrine neoplasia and lung metastases in Rb+/− mice. *Proc Natl Acad Sci USA*. 1999;96(7):3916-3921.
- 56. Clarke AR, Maandag ER, van Roon M, et al. Requirement for a functional RB-1 gene in murine development. *Nature*. 1992;359(6393):328-330.
- 57. Quinonez-Silva G, Davalos-Salas M, Recillas-Targa F, Ostrosky-Wegman P, Aranda DA, Benitez-Bribiesca L. "Monoallelic germline methylation and sequence variant in the promoter of the RB1 gene: a possible constitutive epimutation in hereditary retinoblastoma". *Clin Epigenetics*. 2016;8(1):1-9.
- 58. Kimura H, Nakamura T, Ogawa T, Tanaka S, Shiota K. Transcription of mouse DNA methyltransferase 1 (DNMT1) is regulated by both E2F-Rb-HDACdependent and -independent pathways. *Nucleic Acids Res*. 2003;31(12):3101- 3113.
- 59. Westphal CH, Schmaltz C, Rowan S, Elson A, Fisher DE, Leder P. Genetic interactions between ATM and p53 influence cellular proliferation and irradiationinduced cell cycle checkpoints. *Cancer Res*. 1997;57(9):1664-1667.
- 60. Li S, Ting NS, Zheng L, et al. Functional link of BRCA1 and ataxia telangiectasia gene product in DNA damage response. *Nature*. 2000;406(6792):210-215.
- 61. Yang DQ, Kastan MB. Participation of ATM in insulin signalling through phosphorylation of eIF-4E-binding protein 1. *Nat Cell Biol*. 2000;2(12):893-898.
- 62. Kalev P, Simicek M, Vazquez I, et al. Loss of PPP2R2A inhibits homologous recombination DNA repair and predicts tumor sensitivity to PARP inhibition. *Cancer Res*. 2012;72(24):6414-6424.
- 63. Wu CG, Zheng A, Jiang L, et al. Methylation-regulated decommissioning of multimeric PP2A complexes. *Nat Commun*. 2017;8(1):2272.
- 64. Hughes CM, Rozenblatt-Rosen O, Milne TA, et al. Menin associates with a trithorax family histone methyltransferase complex and with the hoxc8 locus. *Mol Cell*. 2004;13(4):587-597.
- 65. Wu G, Yuan M, Shen S, et al. Author correction: menin enhances c-Myc-mediated transcription to promote cancer progression. *Nat Commun*. 2018;8(1):1-15.
- 66. Gurung B, Feng Z, Iwamoto DV, et al. Menin epigenetically represses hedgehog signaling in MEN1 tumor syndrome. *Cancer Res*. 2013;73(8):2650-2658.
- 67. Busygina V, Kottemann MC, Scott KL, Plon SE, Bale AE. Multiple endocrine neoplasia type 1 interacts with forkhead transcription factor CHES1 in DNA damage response. *Cancer Res*. 2006;66(17):8397-8403.
- 68. Chen G, Jingbo A, Wang M, et al. Menin promotes the WNT signaling pathway in pancreatic endocrine cells. *Mol Cancer Res*. 2008;6(12):1894-1907.
- 69. Huang J, Gurung B, Wan B, et al. The same pocket in menin binds both MLL and JUND but has opposite effects on transcription. *Nature*. 2012;482(7386):542- 546.
- 70. Molatore S, Marinoni I, Lee M, et al. A novel germline CDKN1B mutation causing multiple endocrine tumors: clinical, genetic and functional characterization. *Hum Mutat*. 2010;31(11):E1825-1835.
- 71. Vierimaa O, Georgitsi M, Lehtonen R, et al. Pituitary adenoma predisposition caused by germline mutations in the AIP gene. *Science*. 2006;312(5777):1228- 1230.
- 72. Weinstein LS, Shenker A, Gejman PV, Merino MJ, Friedman E, Spiegel AM. Activating mutations of the stimulatory G protein in the Mccune-Albright syndrome. *N Engl J Med*. 1991;325(24):1688-1695.
- 73. Mougel G, Lagarde A, Albarel F, et al. Germinal defects of SDHx genes in patients with isolated pituitary adenoma. *Eur J Endocrinol*. 2020;183(4):369-379.
- 74. Tadros S, Green MR. Genomic drivers in follicular lymphoma. In: Fowler NH, ed. *Follicular Lymphoma: Current Management and Novel Approaches*. Cham: Springer International Publishing; 2020:47-64.
- 75. García-Ramírez I, Tadros S, González-Herrero I, et al. CREBBP loss cooperates with BCL2 overexpression to promote lymphoma in mice. *Blood*. 2017;129(19):2645-2656.
- 76. Mondello P, Tadros S, Teater M, et al. Selective inhibition of HDAC3 targets synthetic vulnerabilities and activates immune surveillance in lymphoma. *Cancer Discov*. 2020;10(3):440-459.
- 77. Pansuriya TC, van Eijk R, d'Adamo P, et al. Somatic mosaic IDH1 and IDH2 mutations are associated with enchondroma and spindle cell hemangioma in Ollier disease and Maffucci syndrome. *Nat Genet*. 2011;43(12): 1256-1261.
- 78. Amary MF, Bacsi K, Maggiani F, et al. IDH1 and IDH2 mutations are frequent events in central chondrosarcoma and central and periosteal chondromas but not in other mesenchymal tumours. *J Pathol*. 2011;224(3):334-343.
- 79. Kamath-Loeb AS, Zavala-van Rankin DG, Flores-Morales J, et al. Homozygosity for the WRN helicase-inactivating variant, R834C, does not confer a Werner syndrome clinical phenotype. *Sci Rep*. 2017;7(1):1-11.
- 80. Crabbe L, Verdun RE, Haggblom CI, Karlseder J. Defective telomere lagging strand synthesis in cells lacking WRN helicase activity. *Science*. 2004; 306(5703):1951-1953.
- 81. Tokita M, Kennedy SR, Risques RA, et al. Werner syndrome through the lens of tissue and tumour genomics. *Sci Rep*. 2016;6(1):1-10.
- 82. Frappart PO, Tong WM, Demuth I, et al. An essential function for NBS1 in the prevention of ataxia and cerebellar defects. *Nat Med*. 2005;11(5):538- 544.
- 83. Pilia G, Hughes-Benzie RM, MacKenzie A, et al. Mutations in GPC3, a glypican gene, cause the Simpson-Golabi-Behmel overgrowth syndrome. *Nat Genet*. 1996;12(3):241-247.
- 84. Hill DA, Ivanovich J, Priest JR, et al. DICER1 mutations in familial pleuropulmonary blastoma. *Science*. 2009;325(5943):965.
- 85. Rosen-Bronson S, Eckels DD. Longevity of human allospecific TLCs: mycoplasma infection as a cause of in vitro "suppression" of MLC. *Hum Immunol*. 1985;14(4):365-377.
- 86. Villani A, Greer MC, Kalish JM, et al. Recommendations for cancer surveillance in individuals with RASopathies and other rare genetic conditions with increased cancer risk. *Clin Cancer Res*. 2017;23(12):e83-e90.
- 87. Heikkinen SMM, Madanat-Harjuoja LM, Seppa KJM, et al. Familial aggregation of early-onset cancers. *Int J Cancer*. 2020;146(7):1791-1799.
- 88. Conley ME, Notarangelo LD, Etzioni A. Diagnostic criteria for primary immunodeficiencies. Representing PAGID (Pan-American Group for Immunodeficiency) and ESID (European Society for Immunodeficiencies). *Clin Immunol*. 1999;93(3):190-197.
- 89. Stratakis CA, Kirschner LS, Carney JA. Clinical and molecular features of the Carney complex: diagnostic criteria and recommendations for patient evaluation. *J Clin Endocrinol Metab*. 2001;86(9):4041-4046.
- 90. Pilarski R, Eng C. Will the real Cowden syndrome please stand up (again)? Expanding mutational and clinical spectra of the PTEN hamartoma tumour syndrome. *J Med Genet*. 2004;41(5):323-326.
- 91. Pilarski R, Burt R, Kohlman W, Pho L, Shannon KM, Swisher E. Cowden syndrome and the PTEN hamartoma tumor syndrome: systematic review and revised diagnostic criteria. *J Natl Cancer Inst*. 2013;105(21): 1607-1616.
- 92. Li FP, Fraumeni JF Jr, Mulvihill JJ, et al. A cancer family syndrome in twenty-four kindreds. *Cancer Res*. 1988;48(18):5358-5362.
- 93. Bougeard G, Renaux-Petel M, Flaman JM, et al. Revisiting Li-Fraumeni syndrome from TP53 mutation carriers. *J Clin Oncol*. 2015;33(21):2345- 2352.
- 94. Birch JM, Hartley AL, Tricker KJ, et al. Prevalence and diversity of constitutional mutations in the p53 gene among 21 Li-Fraumeni families. *Cancer Res*. 1994;54(5):1298-1304.
- 95. Gutmann DH, Aylsworth A, Carey JC, et al. The diagnostic evaluation and multidisciplinary management of neurofibromatosis 1 and neurofibromatosis 2. *JAMA*. 1997;278(1):51-57.
- 96. Evans DG, King AT, Bowers NL, et al. Identifying the deficiencies of current diagnostic criteria for neurofibromatosis 2 using databases of 2777 individuals with molecular testing. *Genet Med*. 2019;21(7):1525-1533.
- 97. Smith MJ, Bowers NL, Bulman M, et al. Revisiting neurofibromatosis type 2 diagnostic criteria to exclude LZTR1-related schwannomatosis. *Neurology*. 2017;88(1):87-92.
- 98. Bree AF, Shah MR, Group BC. Consensus statement from the first international colloquium on basal cell nevus syndrome (BCNS). *Am J Med Genet A*. 2011;155A(9):2091-2097.
- 99. Baser ME, Friedman JM, Evans DG. Increasing the specificity of diagnostic criteria for schwannomatosis. *Neurology*. 2006;66(5): 730-732.

HEREDITARY CANCER PREDISPOSITION SYNDROMES

- 100. Northrup H, Krueger DA, International Tuberous Sclerosis Complex Consensus G. Tuberous sclerosis complex diagnostic criteria update: recommendations of the 2012 International Tuberous Sclerosis Complex Consensus Conference. *Pediatr Neurol*. 2013;49(4):243-254.
- 101. Miettinen MM, Antonescu CR, Fletcher CDM, et al. Histopathologic evaluation of atypical neurofibromatous tumors and their transformation into malignant

peripheral nerve sheath tumor in patients with neurofibromatosis 1-a consensus overview. *Hum Pathol*. 2017;67:1-10.

- 102. Huang S, Lee L, Hanson NB, et al. The spectrum of WRN mutations in Werner syndrome patients. *Hum Mutat*. 2006;27(6):558-567.
- 103. Tadros S, Ray-Chaudhury A. Pathological features of brain metastases. *Neurosurg Clin N Am*. 2020;31(4):549-564.