# **REVIEW**

WILEY

# Immune cell landscape and immunotherapy of medulloblastoma

Jin Zhang<sup>1,2</sup> | Tianyou Wang<sup>2</sup>

<sup>1</sup>Department of Pediatrics, Beijing Shijitan Hospital, Capital Medical University, Beijing, China <sup>2</sup>Hematology Oncology Center, Beijing Children's Hospital, Capital Medical University, Beijing, China

#### Correspondence

Tianyou Wang, Beijing Children's Hospital, Capital Medical University, Beijing 100045, China Email: wangtianyou@ bch.com.cn

Received: 21 August, 2020 Accepted: 17 October, 2020

#### ABSTRACT

Medulloblastoma is the most common primary pediatric malignancy of the central nervous system. Recurrent and refractory patients account for approximately 30% of them. Immune cells are an important component of the brain tumor microenvironment, including tumor-associated macrophages, T lymphocytes, natural killer cells, dendritic cells, neutrophils and B lymphocytes. Understanding how they behave and interact is important in the investigation of the onset and progression of medulloblastoma. Here, we overview the features and recent advances of each component of immune cells in medulloblastoma. Meanwhile, immunotherapy is a promising but also challenging treatment strategy for medulloblastoma. At present, there are a growing number of immunotherapeutic approaches under investigation including immune checkpoint inhibitors, oncolytic viruses, cancer vaccines, chimeric antigen receptor T cell therapies, and natural killer cells in recurrent and refractory medulloblastoma patients.

#### **KEYWORDS**

Medulloblastoma, Tumor microenvironment, Immune cells, Immunotherapy

# Introduction

Medulloblastoma is the most common primary pediatric malignancy of the central nervous system. With advanced multidisciplinary therapy, the 5-year survival rate has been significantly improved.<sup>1,2</sup> However, approximately 30% of medulloblastoma patients with high risk stratification or recurrences remain incurable.<sup>3,4</sup>

In recent years, the role played by the tumor microenvironment (TME) in promoting or inhibiting tumor growth has attracted widespread attention.<sup>5,6</sup> Immune system cells are an important component of the brain TME. Tumorigenesis is a complex and dynamic process. Diverse immune cells establish complex interactions with each other and with tumor cells described as an intricate network. These interactions promote proliferation and invasion of the tumor by producing growth factors, chemokines and matrix-degrading enzymes.<sup>7</sup> Each of these immune cells contributes to brain tumor biology in unique ways. Understanding their functions and relationships is

critical to understanding the biology of tumor initiation, progression, and metastasis. Despite the low expression of immune cells in medulloblastoma, there are specific variations among the different subgroups. Pham et al<sup>8</sup> adapted murine models of human Sonic Hedgehog (SHH)driven and group 3 medulloblastomas for evaluation. They found that there were more dendritic cells, infiltrating lymphocytes, myeloid-derived suppressor cells, and tumor-associated macrophages in murine SHH medulloblastomas, and murine group 3 medulloblastomas had more CD8<sup>+</sup> T cells. Recently, Bockmayr et al<sup>9</sup> analyzed ten microenvironment cell populations in eight brain tumors including medulloblastoma. Eight immune cell populations were involved: T cells, CD8<sup>+</sup> T cells, cytotoxic lymphocytes, B lineage cells, natural killer (NK) cells, monocytic lineage cells, myeloid dendritic cells, and neutrophils. They discovered that medulloblastoma showed low expression of immune cells but with subgroupspecific infiltration. SHH-driven medulloblastomas had larger numbers of T cells, macrophages, and fibroblasts while group 3 and group 4 medulloblastomas had larger

DOI: 10.1002/ped4.12261

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

<sup>©2021</sup> Chinese Medical Association. Pediatric Investigation published by John Wiley & Sons Australia, Ltd on behalf of Futang Research Center of Pediatric Development.

numbers of CD8<sup>+</sup> T cells and cytotoxic lymphocytes. Interestingly, they also identified two immune stromal patterns (macrophage and regulatory T cell [Treg]mediated mechanisms vs. immunosuppressive cytokines and checkpoints), and they were accurately distributed to the known medulloblastoma subtypes, except for group 4 tumors.

The molecular characterization of medulloblastoma has been intensely researched, and many studies have been done to investigate the additional substructure within subgroups.<sup>10-12</sup> Typically, Cavalli et al<sup>11</sup> identified 12 different subtypes of medulloblastoma, and Northcott et al<sup>10</sup> discovered new subtypes enriched for specific genetic and transcriptional signatures, especially in Group 3 and Group 4. However, the contributions of the immune system to tumorigenesis, progression, response to treatment, and overall prognosis in medulloblastoma remain unclear. In addition, the immune system may provide additional approaches to therapeutic interventions. In this review, we mainly summarize the current knowledge on immune cells in medulloblastoma and immunotherapeutic strategies.

#### Immune cell landscape of medulloblastoma

#### Tumor-associated macrophages (TAMs)

TAMs are a key component of the TME that can promote tumor immune system evasion, inhibit T cell activity, and support tumor growth by promoting angiogenesis or suppressing tumor growth if they are pro-inflammatory.<sup>13</sup> They are the major immune cells within some brain tumors especially in glioma and medulloblastoma that are mainly composed of bone marrow-derived macrophages and brain-resident microglia.<sup>14,15</sup> Considerable data demonstrates that macrophage polarization plays an essential role in the growth and progression of brain tumors.<sup>16</sup> However, the role played by TAMs in medulloblastoma is still unclear.

Margol et al<sup>17</sup> first reported that infiltration of TAMs was significantly higher in SHH medulloblastomas compared with that in other medulloblastoma subgroups and that they contributed to the TME of medulloblastomas. They found that TAMs seen on immunohistochemistry (IHC) were of the M2 phenotype, which had been shown to promote tumor progression via a variety of mechanisms with increased expression of the TAM-related genes, CD163 and CSF1R. However, the roles of TAMs and their activation phenotypes are inconclusive in SHH medulloblastomas. Another study suggested that high levels of M1 rather than M2 macrophages correlated with poor prognosis in SHH medulloblastoma patients, contrary to the common view of the M1 phenotype.<sup>18</sup> These studies suggest that the M1/M2 classification may be reversed and incomplete.

TAMs probably play an important role in leptomeningeal

metastases. A recent study demonstrated that medulloblastoma cells can spread through the blood to the leptomeningeal space, resulting in leptomeningeal metastases.<sup>19</sup> Leptomeningeal metastases express high levels of monocyte chemotactic protein-1 (CCL2), which is responsible for macrophage recruitment. More interestingly, Maximov et al<sup>15</sup> demonstrated recently that TAMs played an active role in SHH medulloblastoma by inhibiting tumor growth, unlike in glioblastoma. This study indicated that TAMs were predominantly of myeloid origin recruited by CCL2 and that they could promote tumor cell death both *ex vivo* and *in vivo*. Furthermore, a reduction or repolarization of TAMs can result in accelerated tumor progression. Overall, whether TAMs promote or suppress tumor growth still remains controversial.

#### T Lymphocytes

T lymphocytes have two major subgroups: CD4<sup>+</sup> cells and CD8<sup>+</sup> cells. CD4<sup>+</sup> cells are considered as helper T lymphocytes (Th). CD8<sup>+</sup> cells are considered as an antitumor factor that leads to apoptosis. CD4<sup>+</sup> and CD8<sup>+</sup> T cell responses are part of the cancer-immunity cycle, and tumor growth is controlled by them.<sup>20</sup> In this context, Tregs are a cluster of cells with a CD4<sup>+</sup> CD25<sup>+</sup> FOXP3<sup>+</sup> phenotype regulating the body's immune response.<sup>21</sup> It has been reported that patients with medulloblastoma show overall reduced CD4<sup>+</sup> T cell counts at diagnosis, and Tregs increase during standard treatment but gradually decline after therapy.<sup>22</sup>

T lymphocytes consistently infiltrate medulloblastomas, and they are recruited to the TME only after the tumor cells have interacted with the tumor vascular endothelium. Macrophage migration inhibitory factor (MIF) is the pivotal chemokine molecule secreted by tumor cells that induces the tumor vascular endothelial cells to secrete potent T lymphocyte attractants.<sup>23</sup> Tumor-infiltrating lymphocytes (TILs) of medulloblastomas, which predominantly infiltrate into perivascular and intratumoral areas, are mainly CD3<sup>+</sup>CD8<sup>+</sup> T cells. Several studies have found that group 3 medulloblastomas have an increased number of CD8<sup>+</sup> T cells.<sup>8,9</sup> The relationship between TILs and the overall survival is unclear. A recent study showed no correlation between TILs and overall survival in medulloblastoma patients.<sup>24</sup> However, in another study, a reduction in TILs predicted a poor prognosis in medulloblastoma patients.25

Immune checkpoints are the important mechanism to escape T-cell mediated immune response. Blockade of immune checkpoints can enhance T cell responses.<sup>26</sup> The precise role of immune checkpoints is not known for multiple cancers. Programmed cell death protein 1 (PD-1) and programmed death-ligand 1 (PD-L1), as negative regulators of T cell immune function, are key immune checkpoints that decrease the cytotoxic activity of CD8<sup>+</sup> T cells towards the tumor.<sup>27</sup> Cyclin-dependent kinase 5 (CDK5) is a kinase required for PD-L1 upregulation that allows medulloblastomas to escape immune elimination. Disruption of CDK5 expression can attenuate tumor PD-L1 expression and promote antitumor immunity.<sup>28</sup> The PD-1/PD-L1 pathway has been shown to be inactive in pediatric cancers.<sup>29,30</sup> Vermeulen et al<sup>24</sup> found that there was a limited number of PD-1<sup>+</sup>T cells and a complete absence of PD-L1 in medulloblastoma.<sup>24</sup> This suggested that there would be a limited value for immunotherapy with PD1/PD-L1 blockers in medulloblastoma. Interestingly, Murata et al<sup>25</sup> observed high expression of PD-L1 in nine (56.3%) of 16 medulloblastoma samples. They showed that medulloblastoma patients with high expression of PD-L1 and a low infiltration of CD8<sup>+</sup> lymphocytes had a worse prognosis. In a recent study, it has been demonstrated that the gene expression levels of PD-L1 were not uniformly low, while single SHH and WNT medulloblastoma cases showed high PD-L1 expression, although it was still undetectable using IHC.<sup>9</sup> In murine models, PD-1 blockade appears to be more effective in group 3 compared with SHH medulloblastomas.<sup>8</sup>

Transforming growth factor  $\beta$  (TGF- $\beta$ ) plays a complex role in tumor initiation and progression. T cells are critical targets of TGF- $\beta$ . TGF- $\beta$  directly inhibits the activities of T cells by suppressing their proliferation, differentiation, and metabolism.<sup>31</sup> It also converts peripheral naive CD4<sup>+</sup> T cells into Tregs that are then recruited to the immunosuppressive TME.<sup>32,33</sup> Therefore, TGF- $\beta$  may reveal itself to be a promising therapeutic target in medulloblastoma. In addition, induction of indoleamine 2,3-dioxygenase 1 (IDO1) can strongly promote the recruitment of Tregs to prevent an immune response against tumor cells.<sup>34</sup> IDO1 inhibitors may present a possible potential for new strategies in the treatment of medulloblastomas.

#### NK cells

NK cells are large granular lymphocytes that can directly kill tumor cells without specific immunization. They can also secrete various cytokines to initiate antitumor responses and recruit other immune cells into the antitumor response.<sup>35,36</sup> The TME can also affect the metabolism and function of NK cells. The TME contains large numbers of immunosuppressive cytokines and other soluble factors that affect the function of NK cells.<sup>37,38</sup>

NK cells can be found in medulloblastomas by using IHC, flow cytometric analysis, RNA sequencing, etc.<sup>8,9,39</sup> It has been confirmed that NK cells can exhibit cytolytic activity against medulloblastomas both *in vitro* and *in vivo*, and they can also migrate from a distal location to the tumor. In a xenograft mouse model, it was shown that NK cells can suppress medulloblastoma growth.<sup>39</sup> Major histocompatibility complex class I-related chain A (MICA) and UL16-binding protein 2 (ULBP2) are tumor cell surface ligands for the NK group 2, member D

(NKG2D) activating receptor on NK cells, and they are prevalent in malignant brain tumors.<sup>40</sup> NKG2D/MICA-ULBP-2 interactions have been considered critical for NK cell cytotoxicity against tumor cells. It has been demonstrated that MICA and ULBP2 are overexpressed on medulloblastoma cells, and NK cell cytotoxicity decreases when NKG2D/NKG2DL interactions are blocked.<sup>41</sup>

Intracranial injection of *ex vivo* expanded human NK cells has therapeutic effects on medulloblastoma xenografts in mouse models. It suggests that patient-derived NK cells could be expanded *ex vivo* and be used for adoptive immunotherapy. In a phase I study, Khatua et al<sup>42</sup> demonstrated the feasibility and safety of intraventricular infusions of *ex vivo* expanded autologous NK cells in recurrent pediatric medulloblastoma patients.<sup>42</sup> It has also been reported that cord blood NK cells may be advantageous in TGF- $\beta$ -rich medulloblastoma patients.<sup>43</sup>

To sum up, because of tumor heterogeneity, NK cells with non-antigen requirements have some advantages compared with other cells in this respect, especially in pediatric brain cancers.<sup>44</sup> Although studies on NK cells in medulloblastomas are relatively few, they have shown the unique potential as a novel opportunity for exploring new treatment strategies.

### Dendritic cells (DCs)

DCs are myeloid-derived, potent antigen-presenting cells (APCs) that are divided into three major subsets: plasmacytoid DC (pDC), myeloid/conventional DC1 (cDC1), and myeloid/conventional DC2 (cDC2).<sup>45,46</sup> They play a critical role in the adaptive immune response by governing T cell immunity and tolerance with a specialized role of the cDC1 subset in CD8<sup>+</sup> T cell priming, cDC2 in CD4<sup>+</sup> T cell priming, and a role for pDCs in immune regulation.<sup>47,48</sup> DCs have been identified to impact disease progression in many malignancies. Infiltration by mature and active DCs into tumors increases immune activation and recruitment of immune effector cells and pathways.<sup>49</sup> More DCs are found in the murine SHH medulloblastoma model.<sup>8</sup>

The most potentially beneficial use of DCs is in the application as a DC vaccine in cancer therapy that may prolong survival in refractory patients. It is noteworthy that DCs are able to be efficiently loaded with a wide range of treatment agents which is a pivotal procedure in immunization strategies, without leading to undue toxicities in patients.<sup>50-52</sup> There have been a number of clinical trials examining the effectiveness of DC vaccination in malignant brain tumors, especially in gliomas. In a large phase III clinical trial of glioblastoma, the addition of an autologous tumor lysate-pulsed DC vaccine (DCVax-L) proved to be feasible and safe.<sup>53</sup> Nevertheless, reports on the application of DCs in medulloblastoma treatment are relatively few at present.

Nair et al<sup>54</sup> successfully generated DCs that met both phenotypic and functional requisites in 2 out of 5 (40%) patient samples. This study supports the feasibility of DC generation and DC-RNA-based vaccination in pediatric medulloblastoma patients. We can also use DCs pulsed with a tumor RNA transcriptome to expand polyclonal tumor-reactive T cells during adoptive T cell therapies in medulloblastoma.<sup>55</sup>

In general, DCs are the feasible option regarding the specific loading of treatment agents to achieve effective antitumor immunization, and further exploration of DCs in the immunotherapeutic treatment of medulloblastomas is warranted.

#### Neutrophils

Neutrophils account for approximately 70% of total leukocytes and are the first line of defense against pathogens. In the context of cancer, tumor-associated neutrophils (TANs) present an N1 (tumor-suppressive) or N2 (tumor-promoting) phenotype, with different characteristics of maturity, tumor cytotoxicity, and immune suppression.<sup>56-59</sup> Neutrophils can activate and regulate immune and inflammatory responses against tumor cells, and they are able to have both pro-tumor and antitumor effects on tumor development. It has been found that TANs play an antitumor role in the early stage of the tumor, but as the tumor progresses, TANs convert to a pro-tumor phenotype, while the antitumor ability decreases gradually.<sup>60,61</sup>

Multiple studies have suggested that the specific location of neutrophils within the tumor is related to prognosis. Neutrophils are often found deeper within the tumor during tumor progression and have an N2 phenotype to promote tumor growth.<sup>57</sup> Neutrophils are observed in medulloblastoma tissues using IHC, appearing with a low infiltration; meanwhile, the SHH group has lower numbers of neutrophils than tumors from other medulloblastoma subgroups.<sup>9</sup>

High counts of blood neutrophils are found in many patients with advanced-stage cancers, and the neutrophilto-lymphocyte ratio (NLR) has been introduced as a prognostic factor for survival in many tumor types. It has been found that a higher pre-operative NLR correlates with a higher histological grade of tumor.<sup>62</sup> Several studies have investigated the significance of the NLR in medulloblastomas. Patel et al<sup>63</sup> observed statistically significantly elevated NLRs in medulloblastoma patients before treatment. They showed that the tumor-induced systemic immune suppression in medulloblastoma patients was already present at the time of diagnosis. In another study, methylation-derived (md)NLRs were measured in peripheral blood samples of pediatric medulloblastoma patients.<sup>64</sup> It has been reported that an elevated mdNLR was significantly associated with mortality in adjusted models. Subsequently, Li et al<sup>65</sup> proved that a high preoperative NLR predicted unfavorable survival in childhood medulloblastoma patients and that levels of preoperative NLRs in group 3 and 4 medulloblastomas were significantly higher than in other groups.

Neutrophils are also being used to indicate a response to treatment. It has been shown that a high peripheral neutrophil count before bevacizumab treatment correlated with a positive and beneficial response to therapy in glioblastoma patients.<sup>66,67</sup> However, no relevant data on medulloblastomas has yet been shown.

Until now, our knowledge of the functions of neutrophils in medulloblastoma patients remains limited. In light of the functional plasticity of neutrophils, there will be more studies focusing on this aspect in the future.

#### **B** Lymphocytes

B lymphocytes play a dual role in the TME.<sup>68,69</sup> They can have antitumor activities by recognizing tumor-specific antigens and producing antibodies, as well as APC function or direct killing of cancer cells. They can also display pro-tumor activities through activation of myeloidderived suppression cells, production of pro-tumorigenic cytokines, or activation of immunosuppressive Tregs. Infiltrating B cells are associated with a positive or neutral prognosis in a variety of tumor types.<sup>70,71</sup>

The role of B lymphocytes in brain tumors has received little attention compared with that of T lymphocytes, which have been studied extensively. The specific role of B lymphocytes in the tumorigenesis and progression of brain tumors remains unclear. On this basis, current studies have been focusing more on their function in gliomas and meningiomas.<sup>72-74</sup> In a glioblastoma model, B cells were shown to act as APCs, playing a critical role in T cell-mediated antitumor immunity and brain tumor regression.<sup>75</sup> They are also involved in response to treatment. A potential mechanism of acquired drug resistance mediated by tumor-associated B cells has been described in melanoma.<sup>76</sup>

Actually, few studies are conducted in the associated field of medulloblastoma. Furthermore, B cells have been shown to minimally infiltrate into medulloblastomas.<sup>9</sup> Therefore, more studies are needed to analyze and explore their potential value in treating medulloblastoma patients and perhaps be used in the development of new treatment strategies.

#### **Current immunotherapy for medulloblastoma**

Medulloblastoma has a low number of infiltrating immune cells and little immunogenicity with minimal tumor mutational burden. Thus, it is challenging to design targets for immunotherapy.<sup>77</sup> Immunotherapy has proven to be a promising strategy in various preclinical research

studies. Nevertheless, no immunotherapeutic approach has revealed convincing clinical results in pediatric medulloblastoma. Still, it is a strategy worth pursuing. Current clinical trials evaluating immunotherapy in medulloblastoma patients are outlined in Table 1.

Current immunotherapy has focused predominantly on the following strategies:

#### Immune checkpoint inhibitors

As mentioned above, Overall levels of PD-1/PD-L1 in human medulloblastomas are low, but some single cases may present high expression of PD-L1.8,30 It has been demonstrated that PD-1 blockade can result in a more effective outcome in group 3 than that in SHH medulloblastomas. Therefore, PD-1/PD-L1 blockers perhaps show limited value for immunotherapy in medulloblastoma patients and the antitumor efficacy may depend on the subtype of medulloblastoma. Nivolumab is well tolerated without dose limiting toxicities in pediatric brain tumors.<sup>78</sup> Nivolumab, pembrolizumab, and durvalumab are undergoing clinical trials separately, recruiting medulloblastoma patients and patients with other brain tumors. In addition, indoximod, an IDO pathway inhibitor, combined with radiation and temozolomide has been investigated in a first-in-children phase I trial.

#### **Oncolytic viral therapy**

Oncolytic measles virus (MV) has potent oncolytic efficacy against medulloblastoma *in vitro* and in mouse models. Intraventricular injection of a modified oncolytic MV into murine xenograft models is safe and can effectively clear medulloblastoma cells.<sup>79-81</sup> G207, a genetically engineered herpes simplex virus (HSV-1) is able to target the highly aggressive MYC-overexpressed group 3 murine medulloblastomas.<sup>82</sup> Poliovirus oncolytic immunotherapy is also a novel approach to treat pediatric brain tumors. The polio:rhinovirus recombinant, PVSRIPO, can significantly decrease cellular proliferation of medulloblastoma *in vitro*.<sup>83</sup> Other effective oncolytic viruses *in vivo* include myxoma virus, reovirus, and Seneca Valley Virus-001 (SVV-001).<sup>84-86</sup> Furthermore, they are currently being tested in clinical trials.

#### Vaccine therapy

The feasibility of vaccination has been verified in preclinical trials in pediatric medulloblastoma patients, as mentioned earlier.<sup>54</sup> But as yet, clinical trials evaluating vaccines have not reached a satisfactory outcome. A phase I/II trial tried to determine the safety of the combination of decitabine and a cancer vaccine for relapsed or refractory patients including medulloblastoma patients. Unfortunately, it was terminated after enrolling one patient. Another phase I study evaluated the safety and maximum tolerated dose of autologous DCs loaded with allogeneic brain tumor stem cells administered as a vaccination with

recurrent brain tumors. Eight patients were involved, but the study was completed with no results posted. At present, several clinical trials on cancer vaccine therapy are underway in recurrent medulloblastoma patients.

#### Chimeric antigen receptor (CAR) T-cell therapy

CAR T cells were initially approved for hematological malignancies. However, CAR T cell therapy has recently begun to be used for brain tumors and has the potential to be integrated into treatment schema for aggressive pediatric malignant brain tumors in the future.<sup>87</sup> Many preclinical studies have shown efficacy for medulloblastomas. It has been reported that B7-H3 (CD276) CAR T cells can mediate significant antitumor activity in vivo and result in regression of medulloblastoma xenografts.<sup>88</sup> Regional and intravenous delivery of human epidermal growth factor receptor 2 (HER2)-CAR T cells causes durable regression in vitro and in murine medulloblastoma models, and their intraventricular delivery in non-human primates is feasible without systemic toxicity.<sup>89</sup> Recently, Donovan et al<sup>90</sup> identified three cell-surface targets, erythropoietinproducing hepatocellular receptor A2 (EPHA2), HER2, and interleukin 13 receptor  $\alpha 2$  (IL13 $\alpha 2$ ), expressed on medulloblastomas, and demonstrated that intrathecal delivery of the three CAR T cells was an effective treatment in group3 medulloblastoma xenografts in mouse models. This study also showed that intrathecal delivery appeared a superior therapeutic effect compared to intravenous delivery approach. Currently, several clinical trials are underway recruiting medulloblastoma patients.

#### Adoptive NK cell therapy

The side effects and appropriate dose of expanded NK cell infusions have been tested in children with recurrent pediatric medulloblastoma and ependymoma in a phase I clinical trial.<sup>41</sup> Nine patients received protocol therapy of up to three infusions weekly, in escalating doses from  $3 \times 10^6$ – $3 \times 10^8$  NK cells/m<sup>2</sup> each infusion, for up to 3 cycles, without dose-limiting toxicities. The study demonstrated the feasibility and safety of intraventricular infusions of autologous NK cells. But the clinical efficacy was disappointing in view of the fact that all patients showed progressive disease, except one patient who had transient stable disease. However, further investigation of NK cell infusions are needed.

#### Radioimmunotherapy

In a phase II trial, compartmental radioimmunotherapy (cRIT) with intraventricular <sup>131</sup>I-3F8 proved to be safe and had clinical utility in maintaining remission in high-risk or recurrent medulloblastoma.<sup>91</sup>

#### Antigenic targets based immunotherapy

CD47 is a protein on the surface of many solid tumors including medulloblastoma, which has been considered

Therapeutic ategory	Trial ID	Phase	Title	Treatment	Status	Enrollment	Sponsor
Immune checkpoint inhibitors	NCT02359565	Ч	Pembrolizumab in treating younger patients with recurrent, progressive, or refractory high- grade gliomas, diffuse intrinsic pontine gliomas, hypermutated brain tumors, ependymoma or medulloblastoma	Pembrolizumab	Recruiting	110	National Cancer Institute (NCI)
	NCT03173950	II	Immune checkpoint inhibitor nivolumab in Nivolumab people with select rare CNS cancers	Nivolumab	Recruiting	180	National Cancer Institute (NCI)
	NCT02793466	Ι	Durvalumab in pediatric and adolescent patients	Durvalumab	Recruiting	36	Children's Hospital Los Angeles
	NCT03130959	Ш	An investigational immuno-therapy study of nivolumab monotherapy and nivolumab in combination with ipilimumab in pediatric patients with high grade primary CNS,	Nivolumab; Ipilimumab	Active, not recruiting	170	Bristol-Myers Squibb
	NCT02502708	Ι	augnancues (CheckMate 908) Study of the IDO pathway inhibitor, indoximod, and temozolomide for pediatric patients with	Indoximod; Temozolomide; Completed Conformal Radiation;	Completed	81	NewLink Genetics Corporation
			progressive primary malignant brain tumors	Cyclophosphamide; Etoposide			
	NCT04049669	Π	Pediatric trial of indoximod with chemotherapy	Indoximod;	Recruiting	140	Theodore S. Johnson
			and radiation for relapsed brain tumors or newly diagnosed DIPG	Full-dose radiation; Temozolomide; Cyclophosphamide; Etoposide; Lomustine			
Oncolytic viral	NCT03911388	Ι	recurrent or	G207	Recruiting	15	University of Alabama at Birmingham
unerapy					:	:	
	NCT03043391	Ι	Phase 1b study PVSRIPO for recurrent PVSRIPO	PVSRIPO	Recruiting	12	Istari Oncology, Inc.
	NCT00314925	Ι	nauguant guoma in children Safety study of Seneca Valley Virus in patients	Seneca Valley Virus	Completed	60	Neotropix
	NCT02962167	Ι	with solid tumors with neuroendocrine features Modified measles virus (MV-NIS) for	Modified measles virus;	Recruiting	46	Sabine Mueller
			children and young adults with recurrent medulloblastoma or recurrent ATRT	Modified measles virus Lumbar puncture			
	NCT02444546	I	Wild-type reovirus in combination with	Laboratory biomarker analysis;	Active, not	9	Mayo Clinic
			sargramostim in treating younger patients with high-grade relapsed or refractory brain tumors	Sargramostim; Wild-type reovirus	recruiting		
Vaccine therapy	NCT02332889	II/I	Phase I/II: decitabine/vaccine therapy in	Vaccine (autologous dendritic Terminated	Terminated	1	University of Louisville
			relapsed/refractory pediatric high grade gliomas/ medulloblastomas/CNS PNETs	cells); Decitabine and Hiltonol			
	NCT01326104	11/1	Vaccine immunotherapy for recurrent medulloblastoma and primitive neuroectodermal	TTRNA-xALT; TTRNA-DCs	Active, not recruiting	17	University of Florida
	NCT03299309	Ι	tumor PEP-CMV in recurrent medulloblastoma/ PEP-CMV malignant glioma (PRiME)	PEP-CMV	Recruiting	30	Gary Archer
			5				(Continues)

wileyonlinelibrary.com/journal/ped4

	(1						
Therapeutic ategory	Trial ID	Phase	Title	Treatment	Status H	Enrollment	Sponsor
	NCT03615404	-	Cytomegalovirus (CMV) RNA-pulsed Ch dendritic cells for pediatric patients and young Tc adults with WHO Grade IV glioma, recurrent malignant glioma, or recurrent medulloblastoma (ATTAC-P)	CMV-DCs; Td (tetanus toxoid)	Active, not recruiting	Ξ	Gary Archer
	NCT01171469	П	Vaccination with dendritic cells loaded with Dendritic Cells, brain tumor stem cells for progressive malignant Imiquimod brain tumor	bendritic Cells; miquimod	Completed	×	Masonic Cancer Center, University of Minnesota
CAR T cell therapy	NCT04270461	П	NKG2D-based CAR T cells immunotherapy for NKG2D-based CAR T cells patient with r/r NKG2DL+ solid tumors		N o t y e t recruiting	10	Jiujiang University Affiliated Hospital
	NCT03638167	-	EGFR806-specific CAR T cell locoregional EGFR806-specific CAR T cell immunotherapy for EGFR-positive recurrent or refractory pediatric CNS tumors	:GFR806-specific CAR T cell	Recruiting	36	Seattle Children's Hospital
	NCT03500991	-	HER2-specific CAR T cell locoregional HER2-specific CAR T cell immunotherapy for HER2-positive recurrent/ refractory pediatric CNS tumors	HER2-specific CAR T cell	Recruiting	48	Seattle Children's Hospital
	NCT04185038	-	CAR T cell by for diffuse midline glioma bediatric CNS	SCRI-CARB7H3(s); B7H3-specific CAR T cell	Recruiting	70	Seattle Children's Hospital
Adoptive NK cell therapy	NCT02271711	н	Expanded natural killer cell infusion in treating Na younger patients with recurrent/refractory brain tumors	Natural killer cell therapy	Active, not recruiting	12	M.D. Anderson Cancer Center
	NCT02100891	Π	Phase 2 STIR Trial: Haploidentical Transplant Allogeneic HCT; and Donor Natural Killer Cells for Solid Tumors Donor NK cell infusion (STIR)	Allogeneic HCT; Donor NK cell infusion	Recruiting	20	Monica Thakar
Radioimmunotherapy	NCT00058370	=	Intrathecal radioimmunotherapy, radiation Ci therapy, and chemotherapy after surgery in Lc treating patients with medulloblastoma Ac Ac 3F	Cisplatin; Lomustine; Vincristine sulfate; Adjuvant therapy; lodine-1-131 monoclonal antibody 3F8; Radiation therapy	Completed	Q	Memorial Sloan Kettering Cancer Center
Antigenic targets based immunotherapy	NCT03652545	П	Multi-antigen T cell infusion against neuro- T <sup>A</sup> oncologic disease (REMIND)	TAA-T	Recruiting	32	Catherine Bollard

IDO, indoleamine 2,3-dioxygenase; PVSRIPO, Polio/Rhinovirus Recombinant; DIPG, diffuse intrinsic pontine glioma; TTRNA, total tumor RNA; xALT, *ex vivo* expanded Autologous Lymphocyte Transfer; PEP-CMV, peptide vaccine derived from cytomegalovirus; EGFR, epidermal growth factor receptor; HER2, human epidermal growth factor receptor 2; HCT, hematopoietic cell transplantation; TAA-T, tumor multi-antigen associated specific cytotoxic T lymphocytes.

TABLE 1 (Continued)

as a viable antigenic target. A humanized anti-CD47 antibody, Hu5F9-G4 demonstrated therapeutic efficacy *in vitro* and *in vivo* in Group 3 medulloblastoma xenograft models.<sup>92</sup> Moreover, a recent study showed that PRAME (PReferentially expressed Antigen in MElanoma) was detectable in 82% of medulloblastoma tissues independent of molecular and histopathologic subgroups, and high PRAME expression was correlated with worse overall survival.<sup>93</sup> This study demonstrated that PRAME was a viable target for adoptive immunotherapy using genetically modified T cells with a PRAME-specific T-cell receptor in mouse models. A phase I research on multi-antigen T cell infusion against neuro-oncologic disease is being carried out.

#### Conclusion

In general, there is significant immunologic heterogeneity in different subgroups of medulloblastoma. Immune cell components have important roles in medulloblastoma development. Further studies are necessary to increase our understanding of their functions and interactions. Immunotherapy is a promising strategy to increase survival with less toxicity in recurrent and refractory medulloblastoma patients. More clinical trials are being explored with the expectation of a major breakthrough in the not too distant future.

## **CONFLICT OF INTEREST**

None.

# REFERENCES

- Massimino M, Biassoni V, Gandola L, Garrè ML, Gatta G, Giangaspero F, et al. Childhood medulloblastoma. Crit Rev Oncol Hematol. 2016;105:35-51.
- von Bueren AO, Kortmann RD, von Hoff K, Friedrich C, Mynarek M, Müller K, et al. Treatment of children and adolescents with metastatic medulloblastoma and prognostic relevance of clinical and biologic parameters. J Clin Oncol. 2016;34:4151-4160.
- Gerber NU, Mynarek M, von Hoff K, Friedrich C, Resch A, Rutkowski S. Recent developments and current concepts in medulloblastoma. Cancer Treat Rev. 2014;40:356-365.
- Sabel M, Fleischhack G, Tippelt S, Gustafsson G, Doz F, Kortmann R, et al. Relapse patterns and outcome after relapse in standard risk medulloblastoma: a report from the HIT-SIOP-PNET4 study. J Neurooncol. 2016;129:515-524.
- 5. Quail DF, Joyce JA. The microenvironmental landscape of brain tumors. Cancer Cell. 2017;31:326-341.
- Byrd T, Grossman RG, Ahmed N. Medulloblastoma-biology and microenvironment: a review. Pediatr Hematol Oncol. 2012;29:495-506.
- Belli C, Trapani D, Viale G, D'Amico P, Duso BA, Della Vigna P, et al. Targeting the microenvironment in solid tumors. Cancer Treat Rev. 2018;65:22-32.
- 8. Pham CD, Flores C, Yang C, Pinheiro EM, Yearley JH, Sayour EJ, et al. Differential immune microenvironments and response to immune checkpoint blockade among

molecular subtypes of murine medulloblastoma. Clin Cancer Res. 2016;22:582-595.

- Bockmayr M, Mohme M, Klauschen F, Winkler B, Budczies J, Rutkowski S, et al. Subgroup-specific immune and stromal microenvironment in medulloblastoma. Oncoimmunology. 2018;7:e1462430.
- Northcott PA, Buchhalter I, Morrissy AS, Hovestadt V, Weischenfeldt J, Ehrenberger T, et al. The wholegenome landscape of medulloblastoma subtypes. Nature. 2017;547:311-317.
- Cavalli FMG, Remke M, Rampasek L, Peacock J, Shih DJH, Luu B, et al. Intertumoral heterogeneity within medulloblastoma subgroups. Cancer Cell. 2017;31:737-754. e6.
- Hovestadt V, Ayrault O, Swartling FJ, Robinson GW, Pfister SM, Northcott PA. Medulloblastomics revisited: biological and clinical insights from thousands of patients. Nat Rev Cancer. 2020;20:42-56.
- Dehne N, Mora J, Namgaladze D, Weigert A, Brüne B. Cancer cell and macrophage cross-talk in the tumor microenvironment. Curr Opin Pharmacol. 2017;35:12-19.
- Hambardzumyan D, Gutmann DH, Kettenmann H. The role of microglia and macrophages in glioma maintenance and progression. Nat Neurosci. 2016;19:20-27.
- Maximov V, Chen Z, Wei Y, Robinson MH, Herting CJ, Shanmugam NS, et al. Tumour-associated macrophages exhibit anti-tumoural properties in Sonic Hedgehog medulloblastoma. Nat Commun. 2019;10:2410.
- Guadagno E, Presta I, Maisano D, Donato A, Pirrone CK, Cardillo G, et al. Role of macrophages in brain tumor growth and progression. Int J Mol Sci. 2018;19:1005.
- Margol AS, Robison NJ, Gnanachandran J, Hung LT, Kennedy RJ, Vali M, et al. Tumor-associated macrophages in SHH subgroup of medulloblastomas. Clin Cancer Res. 2015;21:1457-1465.
- Lee C, Lee J, Choi SA, Kim SK, Wang KC, Park SH, et al. M1 macrophage recruitment correlates with worse outcome in SHH Medulloblastomas. BMC Cancer. 2018;18:535.
- Garzia L, Kijima N, Morrissy AS, De Antonellis P, Guerreiro-Stucklin A, Holgado BL, et al. A hematogenous route for medulloblastoma leptomeningeal metastases. Cell. 2018;172:1050-1062.e4.
- Ostroumov D, Fekete-Drimusz N, Saborowski M, Kühnel F, Woller N. CD4 and CD8 T lymphocyte interplay in controlling tumor growth. Cell Mol Life Sci. 2018;75:689-713.
- Ribeiro Franco PI, Rodrigues AP, de Menezes LB, Pacheco Miguel M. Tumor microenvironment components: Allies of cancer progression. Pathol Res Pract. 2020;216:152729.
- Gururangan S, Reap E, Schmittling R, Kocak M, Reynolds R, Grant G, et al. Regulatory T cell subsets in patients with medulloblastoma at diagnosis and during standard irradiation and chemotherapy (PBTC N-11). Cancer Immunol Immunother. 2017;66:1589-1595.
- 23. Salsman VS, Chow KK, Shaffer DR, Kadikoy H, Li XN, Gerken C, et al. Crosstalk between medulloblastoma cells and endothelium triggers a strong chemotactic signal recruiting T lymphocytes to the tumor microenvironment. PLoS One. 2011;6:e20267.
- Vermeulen JF, Van Hecke W, Adriaansen EJM, Jansen MK, Bouma RG, Villacorta Hidalgo J, et al. Prognostic relevance of tumor-infiltrating lymphocytes and immune checkpoints

in pediatric medulloblastoma. Oncoimmunology. 2017;7:e1398877.

- Murata D, Mineharu Y, Arakawa Y, Liu B, Tanji M, Yamaguchi M, et al. High programmed cell death 1 ligand-1 expression: association with CD8<sup>+</sup> T-cell infiltration and poor prognosis in human medulloblastoma. J Neurosurg. 2018;128:710-716.
- 26. Sharma P, Allison JP. The future of immune checkpoint therapy. Science. 2015;348:56-61.
- 27. Buchbinder EI, Desai A. CTLA-4 and PD-1 pathways: Similarities, differences, and implications of their inhibition. Am J Clin Oncol. 2016;39:98-106.
- Dorand RD, Nthale J, Myers JT, Barkauskas DS, Avril S, Chirieleison SM, et al. Cdk5 disruption attenuates tumor PD-L1 expression and promotes antitumor immunity. Science. 2016;353:399-403.
- 29. Aoki T, Hino M, Koh K, Kyushiki M, Kishimoto H, Arakawa Y, et al. Low frequency of programmed death ligand 1 expression in pediatric cancers. Pediatr Blood Cancer. 2016;63:1461-1464.
- Martin AM, Nirschl CJ, Polanczyk MJ, Bell WR, Nirschl TR, Harris-Bookman S, et al. PD-L1 expression in medulloblastoma: an evaluation by subgroup. Oncotarget. 2018;9:19177-19191.
- Dimeloe S, Gubser P, Loeliger J, Frick C, Develioglu L, Fischer M, et al. Tumor-derived TGF-β inhibits mitochondrial respiration to suppress IFN-γ production by human CD4<sup>+</sup> T cells. Sci Signal. 2019;12:eaav3334.
- Katoh M. Genomic testing, tumor microenvironment and targeted therapy of Hedgehog-related human cancers. Clin Sci (Lond). 2019;133:953-970.
- 33. Polanczyk MJ, Walker E, Haley D, Guerrouahen BS, Akporiaye ET. Blockade of TGF-β signaling to enhance the antitumor response is accompanied by dysregulation of the functional activity of CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> and CD4<sup>+</sup>CD25<sup>-</sup> Foxp3<sup>+</sup> T cells. J Transl Med. 2019;17:219.
- 34. Folgiero V, Miele E, Carai A, Ferretti E, Alfano V, Po A, et al. IDO1 involvement in mTOR pathway: a molecular mechanism of resistance to mTOR targeting in medulloblastoma. Oncotarget. 2016;7:52900-52911.
- Sun H, Sun C. The rise of NK cell checkpoints as promising therapeutic targets in cancer immunotherapy. Front Immunol. 2019;10:2354.
- Schuster IS, Coudert JD, Andoniou CE, Degli-Esposti MA. "Natural Regulators": NK cells as modulators of T cell immunity. Front Immunol. 2016;7:235.
- Terrén I, Orrantia A, Vitallé J, Zenarruzabeitia O, Borrego F. NK cell metabolism and tumor microenvironment. Front Immunol. 2019;10:2278.
- Viel S, Marçais A, Guimaraes FS, Loftus R, Rabilloud J, Grau M, et al. TGF-β inhibits the activation and functions of NK cells by repressing the mTOR pathway. Sci Signal. 2016;9:ra19.
- Kennis BA, Michel KA, Brugmann WB, Laureano A, Tao RH, Somanchi SS, et al. Monitoring of intracerebellarlyadministered natural killer cells with fluorine-19 MRI. J Neurooncol. 2019;142:395-407.
- Castriconi R, Dondero A, Negri F, Bellora F, Nozza P, Carnemolla B, et al. Both CD133<sup>+</sup> and CD133<sup>-</sup> medulloblastoma cell lines express ligands for triggering NK receptors and are susceptible to NK-mediated cytotoxicity. Eur J Immunol. 2007;37:3190-3196.

- Fernández L, Portugal R, Valentín J, Martín R, Maxwell H, González-Vicent M, et al. *In vitro* natural killer cell immunotherapy for medulloblastoma. Front Oncol. 2013;3:94.
- 42. Khatua S, Cooper LJN, Sandberg DI, Ketonen L, Johnson JM, Rytting ME, et al. Phase I study of intraventricular infusions of autologous *ex vivo* expanded NK cells in children with recurrent medulloblastoma and ependymoma. Neuro Oncol. 2020;22:1214-1225.
- Powell AB, Yadavilli S, Saunders D, Van Pelt S, Chorvinsky E, Burga RA, et al. Medulloblastoma rendered susceptible to NK-cell attack by TGFβ neutralization. J Transl Med. 2019;17:321.
- Pérez-Martínez A, Fernández L, Díaz MA. The therapeutic potential of natural killer cells to target medulloblastoma. Expert Rev Anticancer Ther. 2016;16:573-576.
- Mitchell D, Chintala S, Dey M. Plasmacytoid dendritic cell in immunity and cancer. J Neuroimmunol. 2018;322:63-73.
- 46. Collin M, Bigley V. Human dendritic cell subsets: an update. Immunology. 2018;154:3-20.
- Waisman A, Lukas D, Clausen BE, Yogev N. Dendritic cells as gatekeepers of tolerance. Semin Immunopathol. 2017;39:153-163.
- Hansen M, Andersen MH. The role of dendritic cells in cancer. Semin Immunopathol. 2017;39:307-316.
- Tran Janco JM, Lamichhane P, Karyampudi L, Knutson KL. Tumor-infiltrating dendritic cells in cancer pathogenesis. J Immunol. 2015;194:2985-2991.
- Seyfizadeh N, Muthuswamy R, Mitchell DA, Nierkens S, Seyfizadeh N. Migration of dendritic cells to the lymph nodes and its enhancement to drive anti-tumor responses. Crit Rev Oncol Hematol. 2016;107:100-110.
- 51. Kim JW, Kane JR, Panek WK, Young JS, Rashidi A, Yu D, et al. A dendritic cell-targeted adenoviral vector facilitates adaptive immune response against human glioma antigen (CMV-IE) and prolongs survival in a human glioma tumor model. Neurotherapeutics. 2018;15:1127-1138.
- Chandramohan V, Mitchell DA, Johnson LA, Sampson JH, Bigner DD. Antibody, T-cell and dendritic cell immunotherapy for malignant brain tumors. Future Oncol. 2013;9:977-990.
- 53. Liau LM, Ashkan K, Tran DD, Campian JL, Trusheim JE, Cobbs CS, et al. First results on survival from a large Phase 3 clinical trial of an autologous dendritic cell vaccine in newly diagnosed glioblastoma. J Transl Med. 2018;16:142.
- 54. Nair SK, Driscoll T, Boczkowski D, Schmittling R, Reynolds R, Johnson LA, et al. *Ex vivo* generation of dendritic cells from cryopreserved, post-induction chemotherapy, mobilized leukapheresis from pediatric patients with medulloblastoma. J Neurooncol. 2015;125:65-74.
- 55. Flores C, Wildes T, Dean BD, Moore G, Drake J, Abraham R, et al. Massive clonal expansion of medulloblastoma-specific T cells during adoptive cellular therapy. Sci Adv. 2019;5:eaav9879.
- Hinshaw DC, Shevde LA. The tumor microenvironment innately modulates cancer progression. Cancer Res. 2019;79:4557-4566.
- Shaul ME, Fridlender ZG. Tumour-associated neutrophils in patients with cancer. Nat Rev Clin Oncol. 2019;16:601-620.
- 58. Mishalian I, Granot Z, Fridlender ZG. The diversity

of circulating neutrophils in cancer. Immunobiology. 2017;222:82-88.

- 59. Giese MA, Hind LE, Huttenlocher A. Neutrophil plasticity in the tumor microenvironment. Blood. 2019;133:2159-2167.
- Lecot P, Sarabi M, Pereira Abrantes M, Mussard J, Koenderman L, Caux C, et al. Neutrophil heterogeneity in cancer: from biology to therapies. Front Immunol. 2019;10:2155.
- 61. Eruslanov EB. Phenotype and function of tumor-associated neutrophils and their subsets in early-stage human lung cancer. Cancer Immunol Immunother. 2017;66:997-1006.
- 62. Wilson JRF, Saeed F, Tyagi AK, Goodden JR, Sivakumar G, Crimmins D, et al. Pre-operative neutrophil count and neutrophil-lymphocyte count ratio (NLCR) in predicting the histological grade of paediatric brain tumours: a preliminary study. Acta Neurochir (Wien). 2018;160:793-800.
- Patel S, Wang S, Snuderl M, Karajannis MA. Pre-treatment lymphopenia and indication of tumor-induced systemic immunosuppression in medulloblastoma. J Neurooncol. 2018;136:541-544.
- Arroyo VM, Lupo PJ, Scheurer ME, Rednam SP, Murray J, Okcu MF, et al. Pilot study of DNA methylation-derived neutrophil-to-lymphocyte ratio and survival in pediatric medulloblastoma. Cancer Epidemiol. 2019;59:71-74.
- 65. Li K, Duan WC, Zhao HB, Wang L, Wang WW, Zhan YB, et al. Preoperative neutrophil to lymphocyte ratio and platelet to lymphocyte ratio are associated with the prognosis of Group 3 and Group 4 medulloblastoma. Sci Rep. 2019;9:13239.
- 66. Quillien V, Carpentier AF, Gey A, Avril T, Tartour E, Sejalon F, et al. Absolute numbers of regulatory T cells and neutrophils in corticosteroid-free patients are predictive for response to bevacizumab in recurrent glioblastoma patients. Cancer Immunol Immunother. 2019;68:871-882.
- Bertaut A, Truntzer C, Madkouri R, Kaderbhai CG, Derangère V, Vincent J, et al. Blood baseline neutrophil count predicts bevacizumab efficacy in glioblastoma. Oncotarget. 2016;7:70948-70958.
- Largeot A, Pagano G, Gonder S, Moussay E, Paggetti J. The B-side of cancer immunity: The Underrated Tune. Cells. 2019;8:449.
- 69. Yuen GJ, Demissie E, Pillai S. B lymphocytes and cancer: a love-hate relationship. Trends Cancer. 2016;2:747-757.
- Shen M, Wang J, Ren X. New insights into tumorinfiltrating B lymphocytes in breast cancer: clinical impacts and regulatory mechanisms. Front Immunol. 2018;9:470.
- 71. Wouters MCA, Nelson BH. Prognostic significance of tumor-infiltrating B cells and plasma cells in human cancer. Clin Cancer Res. 2018;24:6125-6135.
- Domingues P, González-Tablas M, Otero A, Pascual D, Miranda D, Ruiz L, et al. Tumor infiltrating immune cells in gliomas and meningiomas. Brain Behav Immun. 2016;53:1-15.
- 73. Fang L, Lowther DE, Meizlish ML, Anderson RC, Bruce JN, Devine L, et al. The immune cell infiltrate populating meningiomas is composed of mature, antigen-experienced T and B cells. Neuro Oncol. 2013;15:1479-1490.
- 74. Pucci F. Location-dependent B-cell function in glioblastoma. Cancer Immunol Res. 2019;7:1902.
- 75. Candolfi M, Curtin JF, Yagiz K, Assi H, Wibowo MK, Alzadeh GE, et al. B cells are critical to T-cell-

mediated antitumor immunity induced by a combined immune-stimulatory/conditionally cytotoxic therapy for glioblastoma. Neoplasia. 2011;13:947-960.

- Somasundaram R, Zhang G, Fukunaga-Kalabis M, Perego M, Krepler C, Xu X, et al. Tumor-associated B-cells induce tumor heterogeneity and therapy resistance. Nat Commun. 2017;8:607.
- Blaeschke F, Paul MC, Schuhmann MU, Rabsteyn A, Schroeder C, Casadei N, et al. Low mutational load in pediatric medulloblastoma still translates into neoantigens as targets for specific T-cell immunotherapy. Cytotherapy. 2019;21:973-986.
- Gorsi HS, Malicki DM, Barsan V, Tumblin M, Yeh-Nayre L, Milburn M, et al. Nivolumab in the treatment of recurrent or refractory pediatric brain tumors: a single institutional experience. J Pediatr Hematol Oncol. 2019;41:e235-e241.
- Lal S, Peng KW, Steele MB, Jenks N, Ma H, Kohanbash G, et al. Safety study: intraventricular injection of a modified oncolytic measles virus into measles-immune, hCD46transgenic, IFNαRko mice. Hum Gene Ther Clin Dev. 2016;27:145-151.
- Hutzen B, Bid HK, Houghton PJ, Pierson CR, Powell K, Bratasz A, et al. Treatment of medulloblastoma with oncolytic measles viruses expressing the angiogenesis inhibitors endostatin and angiostatin. BMC Cancer. 2014;14:206.
- Studebaker AW, Kreofsky CR, Pierson CR, Russell SJ, Galanis E, Raffel C. Treatment of medulloblastoma with a modified measles virus. Neuro Oncol. 2010;12:1034-1042.
- Bernstock JD, Vicario N, Li R, Nan L, Totsch SK, Schlappi C, et al. Safety and efficacy of oncolytic HSV-1 G207 inoculated into the cerebellum of mice. Cancer Gene Ther. 2020;27:246-255.
- Thompson EM, Brown M, Dobrikova E, Ramaswamy V, Taylor MD, McLendon R, et al. Poliovirus receptor (CD155) expression in pediatric brain tumors mediates oncolysis of medulloblastoma and pleomorphic xanthoastrocytoma. J Neuropathol Exp Neurol. 2018;77:696-702.
- 84. Lun XQ, Zhou H, Alain T, Sun B, Wang L, Barrett JW, et al. Targeting human medulloblastoma: oncolytic virotherapy with myxoma virus is enhanced by rapamycin. Cancer Res. 2007;67:8818-8827.
- Yang WQ, Senger D, Muzik H, Shi ZQ, Johnson D, Brasher PM, et al. Reovirus prolongs survival and reduces the frequency of spinal and leptomeningeal metastases from medulloblastoma. Cancer Res. 2003;63:3162-3172.
- Yu L, Baxter PA, Zhao X, Liu Z, Wadhwa L, Zhang Y, et al. A single intravenous injection of oncolytic picornavirus SVV-001 eliminates medulloblastomas in primary tumorbased orthotopic xenograft mouse models. Neuro Oncol. 2011;13:14-27.
- Patterson JD, Henson JC, Breese RO, Bielamowicz KJ, Rodriguez A. CAR T cell therapy for pediatric brain tumors. Front Oncol. 2020;10:1582.
- Majzner RG, Theruvath JL, Nellan A, Heitzeneder S, Cui Y, Mount CW, et al. CAR T cells targeting B7-H3, a pancancer antigen, demonstrate potent preclinical activity against pediatric solid tumors and brain tumors. Clin Cancer Res. 2019;25:2560-2574.
- Nellan A, Rota C, Majzner R, Lester-McCully CM, Griesinger AM, Mulcahy Levy JM, et al. Durable regression of medulloblastoma after regional and intravenous

delivery of anti-HER2 chimeric antigen receptor T cells. J Immunother Cancer. 2018;6:30.

- Donovan LK, Delaidelli A, Joseph SK, Bielamowicz K, Fousek K, Holgado BL, et al. Locoregional delivery of CAR T cells to the cerebrospinal fluid for treatment of metastatic medulloblastoma and ependymoma. Nat Med. 2020;26:720-731.
- Kramer K, Pandit-Taskar N, Humm JL, Zanzonico PB, Haque S, Dunkel IJ, et al. A phase II study of radioimmunotherapy with intraventricular <sup>131</sup>I-3F8 for medulloblastoma. Pediatr Blood Cancer. 2018;65:10.1002/ pbc.26754.
- 92. Gholamin S, Mitra SS, Feroze AH, Liu J, Kahn SA, Zhang

M, et al. Disrupting the CD47-SIRP $\alpha$  anti-phagocytic axis by a humanized anti-CD47 antibody is an efficacious treatment for malignant pediatric brain tumors. Sci Transl Med. 2017;9:eaaf2968.

93. Orlando D, Miele E, De Angelis B, Guercio M, Boffa I, Sinibaldi M, et al. Adoptive immunotherapy using PRAME-specific T cells in medulloblastoma. Cancer Res. 2018;78:3337-3349.

**How to cite this article:** Zhang J, Wang T. Immune cell landscape and immunotherapy of medulloblastoma. Pediatr Investig. 2021;5:299-309. https://doi.org/10.1002/ped4.12261