#### REVIEW

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### Microbiota and glioma: a new perspective from association to clinical translation

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#### ABSTRACT

Gliomas pose a significant challenge in oncology due to their malignant nature, aggressive growth, frequent recurrence, and complications posed by the blood-brain barrier. Emerging research has revealed the critical role of gut microbiota in influencing health and disease, indicating its possible impact on glioma pathogenesis and treatment responsiveness. This review focused on existing evidence and hypotheses on the relationship between microbiota and glioma from progression to invasion. By discussing possible mechanisms through which microbiota may affect glioma biology, this paper offers new avenues for targeted therapies and precision medicine in oncology.



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#### 1. Introduction

Gliomas, predominantly malignant tumors originating in the brain or spinal cord, pose significant challenges in neuro-oncology due to their rapid progression, aggressive nature, and high propensity for relapse<sup>1</sup>. With the incidence of 6.0 per 100 000 population, gliomas take up 81% of malignant brain tumors,<sup>2,3</sup> varying widely in their aggressiveness and can be classified into grade 1–4 based on their cellular characteristics and rate of growth, which profoundly influences the clinical outcome of glioma patients. Low-grade gliomas may exhibit favorable prognosis after surgery, while high-grade tumors, such as glioblastoma, the most aggressive grade 4 glioma, are associated with dismal survival rates, averaging 14 to 16 months even with comprehensive treatments, and a 5-year survival rate of under 10%.<sup>4</sup>

The diagnostic and therapeutic landscape of gliomas is fraught with hurdles. Early detection of gliomas has been difficult, as they often present nonspecific symptoms in the early stages,

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complicating timely and accurate diagnosis.<sup>5,6</sup> Moreover, chemoresistance and the impermeable nature of the blood-brain barrier (BBB) hampers the detection of tumor components, as well as drug delivery.<sup>7,8</sup> Additionally, the inherent heterogeneity of gliomas contributes to the complexity of managing the disease, as it underpins the variations in treatment response and the emergence of resistance, thereby elevating the risk of disease progression and dissemination.<sup>9</sup>

Recent research has illuminated the significant role of microbiota - communities of microorganisms including bacteria, viruses, fungi, archaea, and protozoa - that inhabit various parts of the human body, in health and disease.<sup>10</sup> Human microbiota has been observed to exert both local and systemic effects on the onset, progression and therapy response to many cancers, including colorectal cancer and and bladder cancer.<sup>11</sup> Notably, the identification of microbial components within the brain introduces the concept of a brain microbiome, implicating its probable involvement in the pathogenesis of glioma.<sup>12,13</sup> Therefore, understanding the intricate relationships between microbiota and glioma holds significant promise for revolutionizing the diagnostic and therapeutic approaches for this formidable disease. <sup>14–16</sup>

This review aims to summarize the potential contributions of microbiota to the development of gliomas. It integrates the possible mechanisms underlying the influence of microbiota on tumor growth and immune evasion, highlights the potential interplay between microbiota and glioma therapies, and discusses the prospective utility of these insights into refining diagnostic and therapeutic strategies for glioma. By integrating the current knowledge on the microbiota-glioma axis, we endeavor to shed light on novel pathways for intervention and ultimately improve outcomes for patients suffering from this challenging condition.

## 2. Relationship between microbiota and different types of glioma

Glioma may derive from diverse types of cells in the brain or spinal cord, including astrocytes, oligodendrocytes, and ependymal cells. The microbiome may play a pivotal role in the onset, progression, and invasion of different gliomas, possibly through mechanisms such as enhancing proliferative signaling, inhibiting apoptosis, and promoting angiogenesis.<sup>17</sup> Given the diverse nature of gliomas, the varying influences of different microbial communities within the brain, gut, lung, and oral cavity have sparked extensive research into their specific roles across glioma subtypes (Table 1).

### 2.1. Microbiota and glioblastoma

Glioblastoma, used to be called GBM, is classified as a WHO grade 4 glioma, standing out as the most prevalent and aggressive adult glioma with a poor prognosis.<sup>38</sup> The pivotal role of the human microbiome, particularly the gut microbiome, in glioma development has been a focus of recent research. Studies unveiled significant differences in the gut microbiota composition between glioma patients (or glioma-bearing mice) and healthy controls, highlighting a potential link to disease development.<sup>39</sup> Furthermore, clinical trials have revealed a shift in the concentration of metabolites derived from gut bacteria, suggesting a complex interplay between gut microbiota and glioblastoma progression.<sup>40</sup>

Mendelian randomization studies have advanced our understanding by investigating the association between specific gut microbiota compositions and glioblastoma risk. They identified that an increased presence of certain genera (Fusobacterium, Akkermansia, Escherichia/ Shigella) and a decreased presence of others (Lachnospira, Agathobacter, Bifidobacterium) correlate with various malignant glioma types.<sup>18</sup> Interestingly, other research pointed out that the Ruminococcaceae family, not previously highlighted, significantly correlates with a reduced glioblastoma risk, indicating the complexity of microbial influences on glioma.<sup>19</sup> The varying results may attribute to limited sample sizes, study design issues or population differences, calling for more comprehensive studies on the underlying associations.

Oral microbiome, where specific microbial features have been associated with glioma malignancy and IDH1 mutation, may also be a potential biomarker of glioblastoma. Notable findings include the ability of certain oral

Table 1. Roles of microbiome in different gliomas.

Condition	Location	Components	Effect/Association
Glioblastoma	Gut microbiome	Fusobacterium ↑ Akkermansia ↑ Escherichia/Shigella ↑ Lachnospira ↓ Agathobactera ↓ Bifidobacterium ↓	Correlation with various malignant glioma types <sup>18</sup>
		Ruminococcaceae Granulicatella Rothia mucilaginosa	Decreased risk of glioblastoma <sup>19</sup> Dysbiosis following SARS-CoV-2 infection, <sup>20</sup> potentially accelerating GBM progression <sup>21</sup>
	Oral microbiome	Porphyromonas Haemophilus Leptotrichia TM7x Capnocytophaga	Inverse correlation with glioma grades; Accurate discrimination of HGG from LGG and HCs <sup>22</sup>
		Bergeyella, Capnocytophaga Granulicatella Rothia mucilaginosa Capnocytophaga Veillonella	Positive associations with IDH1 mutation in glioma (vs. IDH1-wild-type group) <sup>22</sup> Dysbiosis following SARS-CoV-2 infection, <sup>20</sup> potentially accelerating GBM progression <sup>21,23</sup>
	Intratumoral microbiome	Proteobacteria Firmicutes SARS-CoV-2	Discovered by comprehensive histological imaging, Sudan B treatment and a 3D whole-tumor perspective <sup>24-26</sup> Direct infection and high expression of ACE2 in GBM tissues; <sup>27</sup> potential opcogenic effects via binding ACE2 <sup>28</sup>
Astrocytoma	Gut microbiome	Parabacteroides distasonis MRx0005; Megasphaera massiliensis MRx0029	Strong capacity to reduce IL-6 secretion <i>in vitro</i> ; <sup>29</sup> may be associated with low IL-6 level in grade 1-2 astrocytomas <sup>30</sup>
	Brain microbiome Injection into the dorsal hippocampus	Brucella abortus Lentivirus (family Retroviridae)	Potential role of microbes in brain tissue in glioma development <sup>31,32</sup> Selective targeting of astrocytes; <sup>33</sup> Manipulation of oncogenic pathways <i>via</i> gene transfer <sup>34,35</sup>
Oligodendroglioma	Uncertain	LPS	Damage on astrocytes through activation of TLR4-CD14/TLR2 receptors and NF-xB mediated cytokine surges <sup>36</sup>
DMG	Gut microbiome	F/B ratio↓ Flavobacteriaceae ↑ Bacillales ↑	Association with higher risk of DMG <sup>37</sup>

F/B, Firmicutes/Bacteroidetes.

microbial signatures to distinguish high-grade gliomas from low-grade ones.<sup>22</sup> External factors causing dysbiosis, including viral infections like SARS-CoV-2, may further complicate glioblas-toma outcomes by adversely affecting the microbiota.<sup>41</sup>

In addition to external microbiota, the role of intratumoral microbiota in glioblastoma progression has gained attention. Techniques such as immunohistochemistry and RNA fluorescence in situ hybridization have identified bacterial components within glioblastoma tissues, revealing predominance of Proteobacteria а and *Firmicutes.*<sup>42</sup> This evidence, supported by advanced imaging techniques, indicates the probable interplay between the microbiome and the tumor microenvironment (TME), offering new insights into glioblastoma pathology and potential therapeutic targets.<sup>24–26</sup>

#### 2.2. Microbiota and other gliomas

Although the role of microbiota in glioblastoma has been the focal point of research, its implications for other glioma types remain less explored. However, emerging studies provide a basis for the investigation of microbial characteristics in the diagnosis and treatment of diverse glioma subtypes.

Previous studies revealed that the dysbiosis of gut microbiome are closely associated with astrocytic dysfunction in mouse models, indicating similar effects on development of pilocytic astrocytoma (grade 1) and diffuse astrocytoma (grade 2).<sup>43</sup> Specifically, *Parabacteroides distasonis* MRx0005 and *Megasphaera massiliensis* MRx0029 demonstrated strong capacity to reduce IL-6 secretion *in vitro*, which may be associated with low IL-6 level in low-grade gliomas mentioned above.<sup>29,30</sup> Furthermore, a recent study revealed the activation of

microglia by *Brucella abortus*-infected astrocytes, suggesting the potential role of brain microbiome in progression and maintenance of astrocytoma.<sup>31,32</sup> In addition to bacteria, lentivirus are also capable of selective targeting and sustained gene expression in astrocytes, demonstrating the potential for microbial components to be involved in oncogenic pathways in glioma.<sup>34,35</sup>

Oligodendroglioma (grade 2 or grade 3), derived from oligodendrocytes which cover and protect nerve cells in the brain and spinal cord, grows slowly and is less common. The impact of lipopolysaccharides (LPS) on oligodendroglioma development and treatment response has been identified, highlighting how LPS-triggered activation of TLR4-CD14/TLR2 receptors may damage oligodendrocytes through NF- $\kappa$ B mediated cytokine surges.<sup>36</sup> This insight into microbe-induced cellular responses opens new avenues for understanding glioma pathology and enhancing therapeutic strategies.

In the case of diffuse midline glioma (DMG), a grade 4 pediatric brain tumor noted for its poor prognosis, recent trials have attempted to integrate microbiome analysis to aid its diagnosis and evaluate its therapeutic efficacy. Key findings suggest that *Firmicutes/Bacteroidetes* (F/B) ratio of gut microbiota, along with the presence of *Flavobacteriaceae* and *Bacillales*, correlates with DMG patients' progression-free survival and overall survival (OS) rates.<sup>37</sup>

These examples underscore the burgeoning interest in the microbiome's role beyond glioblastoma, offering promising diagnostic and therapeutic avenues for various glioma types. As research continues to unfold, the microbiota's multifaceted contributions to glioma pathogenesis and treatment responsiveness are poised to become increasingly significant.

### 3. Regulatory mechanisms of microbiome-glioma interaction

Recent studies have indicated the possible roles of microbes in modulating the development and the microenvironment of gliomas. These organisms may exert their influence through a combination of direct neural and endocrine pathways, alongside the actions of microbial metabolites. The understanding of the potential microbiota-glioma interaction offers novel insights that have profound implications for the diagnosis and treatment of gliomas (Figure 1).

## **3.1. Neural and endocrine modulation of glioma** progression by microbiota

Research into the regulation of microbiome-glioma interaction has predominantly concentrated on the gut-brain axis, a bidirectional communication network that connects the gut microbiota with the central nervous system (CNS). This axis may impact glioma progression through a variety of mechanisms. Gut microbes and their metabolites, for example, may influence CNS activity and pathologies by modulating neural signaling pathways, including those associated with the vagus nerve.<sup>44</sup> Such modulation may indirectly drive glioma growth and invasiveness by affecting neurotransmitter release and neuroinflammatory responses.<sup>45</sup> Moreover, the gut microbiome's ability to influence neuroplasticity may play a pivotal role in both the prognosis and management of glioma patients. 46,47 The interaction extends further, impacting CNS function through the regulation of hormone levels, including cortisol and sex hormones, potentially altering glioma cell behavior in terms of proliferation and survival.<sup>48,49</sup> This hormonal influence may be exacerbated by increasing levels of LPS, a potent immunogenic component of gram-negative bacteria that induces endotoxemia and compromises the BBB.<sup>50</sup>

The state of dysbiosis, especially the imbalance of bacterial composition and changes in bacterial metabolic activities, introduces additional complexities.<sup>51</sup> Various factors, such as chronic stress, depression, and the prolonged or inappropriate use of antibiotics, can significantly alter the gut microbiota's composition and diversity.<sup>52</sup> This altered microbial environment may facilitate the release of ATP and glutamate from astrocytes, creating conditions conducive to glioma emergence and invasion.<sup>53</sup> Conversely, the development of glioma itself can induce structural changes in the gut microbiota, marked by decreased abundance of Actinobacteria and Bacteroidia and increased abundance of Firmicutes. These shifts may



**Figure 1.** Potential interactions between the microbiome and glioma progression. The illustration delineates the probable modulatory pathway of the gut microbiome, mediated by hormonal release and the vagus nerve. Highlighted are the microbial metabolites' possible roles in synthesizing essential molecules like glucose, lipids, and amino acids within tumor cells. Notably, dysbiosis— triggered by factors such as chronic stress, depression, and antibiotic misuse—facilitates neurotransmitter (e.g., ATP, glutamate) release from astrocytes, may promote glioma onset and invasion. Furthermore, the feedback loop where glioma growth alters gut microbiota composition, indirectly affecting intestinal barrier integrity, suggests the bidirectional interaction between the microbiome and glioma progression.

suppress Foxp3 expression in the brain, further promoting glioma development.<sup>54</sup> By examining these intricate interactions, we gain insight into the possible impact of the microbiome on glioma pathology, offering potential pathways for innovative diagnostic and therapeutic strategies.

#### 3.2. Microbiome and glioma metabolism

Metabolic byproducts from the microbiome, such as short-chain fatty acids (SCFAs), may play a pivotal role in host cell physiology by serving as energy substrates. SCFAs, including butyrate, propionate, and acetate, not only fuel intestinal epithelial cells to support energy metabolism and cellular regeneration, but also impact the integrity of the intestinal barrier. This dual functionality underscores the significance of microbiome-derived metabolites in maintaining host health and their potential implications in disease processes. SCFAs may be harnessed by glioma cells as an energy source, which may contribute to their proliferation and suggest a complex interaction between microbiome metabolites and tumor biology.<sup>55</sup> Additionally, the growth of glioma leads to changes in lipid metabolism, indicating potential avenues for identifying lipid-based biomarkers or therapeutic targets.<sup>56</sup> The gut microbiome's metabolic pathways possibly intersect with the host's biosynthesis of glucose, fatty acids, and amino acids. This may either directly or indirectly support tumor cell growth and survival. One of the critical pathways affected by SCFAs is the insulin signaling pathway, potentially involved in glioma cells' energy metabolism and growth signal transduction.<sup>57</sup> By activating various G protein-coupled receptors (GPCRs) such as GPR41, GPR43, and GPR109A, SCFAs can modulate insulin sensitivity and inflammation, likely leading to insulin resistance and impacting glioma cells' nutrient absorption and utilization.<sup>58,59</sup>

Moreover, the microbiome may influence the host's fatty acid composition and metabolism, thereby affecting tumor cell fatty acid oxidation and synthesis. Gut microbiota-mediated lipid metabolism, through the production of metabolites like SCFAs, secondary bile acids, trimethylamine, and LPS, can alter fatty acid absorption and transport.<sup>60</sup> This modulation affect the host's fat storage and energy utilization, which may subsequently influence glioma cells' energy supply and growth conditions. The production of essential amino acids such as tryptophan, arginine, and glutamate, or the generation of energy through the metabolism of nonessential amino acids like branched-chain amino acids, further illustrate the microbiome's potential role in glioma proliferation and survival.<sup>61,62</sup>

Microbial metabolites may also have a profound impact on the intestinal barrier's integrity and function. Compromised barrier integrity facilitates the entry of more inflammatory mediators and immune cells into circulation, potentially affecting distant glioma cells.<sup>63</sup> Some metabolites, notably SCFAs like butyrate and secondary bile acids, are key to maintaining intestinal epithelial cell integrity, thereby enhancing barrier function.<sup>64,65</sup> Conversely, other metabolites, including proteases and LPS, can weaken the intestinal barrier by damaging its physical defenses and inducing inflammation, leading to increased permeability.<sup>66,67</sup> This dynamic interplay between microbial metabolites and the intestinal barrier indicates the complexity of microbiome-host interactions and their relevance to glioma development.

### 4. Role of microbiota in immunomodulation against glioma

The potential interplay between the microbiome and glioma involves a series of molecular and cellular mechanisms which may orchestrate immunomodulation (Figure 2). This interaction may encompass immune activation, immune escape, and the regulation of the immune microenvironment, creating conditions that may either support or hinder glioma progression. Microbiota's possible roles in immune stimulation and escape within the glioma context indicate the intricate balance between oncogenic and oncolytic influences, and underscore the critical potential of leveraging microbiota interactions for developing novel glioma therapies, aiming to boost immune surveillance while counteracting the tumor's strategies for immune evasion.

## **4.1.** *Microbiota may enhance immune surveillance against glioma cells*

The microbiota, particularly the gut microbiota, may exert a profound impact on the host immune system's surveillance of tumor cells, utilizing its metabolic products and immunomodulatory functions to influence both systemic immune responses and CNS immunity. Some microbial derivatives can activate the host immune system, potentially enhancing its ability to combat gliomas. Notably, bacterial peptides can stimulate tumor-infiltrating lymphocytes (TILs), such as CD4<sup>+</sup> T cells and peripheral blood memory cells, to improve against glioblastoma.68 immune reactivity Flagellin, a whip-like appendage that enables bacterial motility, can reduce the number of myeloidderived suppressor cells (MDSCs) in tumor tissues and adjust the conversion of tumor-associated macrophages (TAMs) from M2 to M1 type.<sup>69,70</sup> Additionally, the gut microbiota promotes dendritic cell maturation, augmenting their capacity to present tumor antigens to T cells, an effect attributed to the production of SCFAs like butyrate.<sup>71</sup> The microbiome may also engage the host's innate immune system via pattern recognition receptors, such as Toll-like receptors, activating innate immune cells (NK cells, macrophages, neutrophils) to enhance glioma cell recognition and elimination.



**Figure 2.** Potential effects of microbiome on immunomodulation. Microbiota may either promote or inhibit glioma growth *via* host immune system and modifications of TME. (1) microbial components and metabolites are able to initiate host immune response, inducing the polarization of TAMs into inflammatory (M1) phenotype, the maturation of dendritic cells, the inhibition of MDSCs, as well as the stimulation and differentiation of TILs. While metabolites such as polyamines potentially upregulate PD-L1 expression on APCs, which may potentiate treg cell expansion and immune escape. (2) AhR, detected in TAMs, can be activated by gut microbiota-derived indole-based metabolites, driving TAMs recruitment into GBM. *Fusobacterium nucleatum* may activate microglia *via* TLR2 and TLR4 pathway, upregulating cytokine (tnf- $\alpha$ , IL-1 $\beta$ , IL-6, and iNOS) secretion conducive to glioma development, which may also be suppressed by the release of CXCL10 induced by viral infection. Some microbe or its metabolites may be recognized as tumor antigen *via* molecular mimicry, promoting cross-reactive immune response, thereby destroying glioma cells.

Moreover, SCFAs contribute to the regulation of both innate and adaptive immunity, guiding the differentiation of T and B cells.<sup>55</sup> Specific strains, Parabacteroides as distasonis and such Megasphaera massiliensis, have been identified to modulate immune responses, with the latter showing potential in reducing IL-6 secretion and oxidastress, therefore inhibiting M2-like tive macrophage polarization.<sup>72,73</sup> To the opposite, the antibiotic treatment and the resulting alterations in gut microbiota may contribute to glioma growth by impairing the function of innate immune cells, such as NK cells and microglia.<sup>74,75</sup>

# **4.2.** Microbiota may promote immune escape of glioma

Certain microbial metabolites may facilitate immune escape, supporting glioma growth. Some gut microbes produce metabolites that inhibit immune cell functions, potentially affecting the PD-L1 response in glioma-bearing mice a mechanism observed in other non-CNS cancers.<sup>39</sup> Specifically, polyamines from the urogenital microbiota may suppress effector T cell activity by upregulating PD-L1 expression on antigen-presenting cells,<sup>76</sup> suggesting that a similar pathway may promote glioma growth. Additionally, the microbiome may foster an immunosuppressive environment by enhancing the proliferation and function of regulatory T cells (Tregs), mediated by fluctuations in Foxp3 expression, probably reducing immune surveillance against gliomas.<sup>77,78</sup>

## **4.3.** Impact of microbiota on glioma immune microenvironment

The microbiome may exert a profound influence on immune surveillance within the tumor immune microenvironment, a complex assembly of cells, molecules, and signaling pathways that are pivotal in shaping immune responses against glioma. A key mediator of this intricate host-microbiota interaction is the aryl hydrocarbon receptor (AhR). Research indicates that the receptor's ligand, a metabolite produced by gut microbiota, plays a crucial role; its knockdown has been shown to increase glioblastoma cell invasion, indicating the receptor's involvement in maintaining a balance between tumor suppression and promotion.<sup>79</sup>

The microbiome, encompassing both gut and other tumor-associated microorganisms, may dynamically influence the expression of cytokines and chemokines within the TME. Fusobacterium nucleatum, an oral commensal organism associated with pathological changes in the brain, activates microglia via TLR2 and TLR4 pathway, producing TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and iNOS. These products lead to local inflammation in CNS, potentially promoting the occurrence and progression of glioma.<sup>80</sup> Both DNA and RNA viral infections induce secretion of CXCL10, enhancing the recruitment of adoptively transferred therapeutic T cells to CNS, potentially improves the survival in a mouse model of GBM.<sup>81,82</sup> The shifts in the levels of proinflammatory and anti-inflammatory cytokines may directly impact the recruitment, activation, and efficacy of immune cells, which in turn implicates the efficiency of immune surveillance mechanisms targeting glioma cells.<sup>83</sup>

Moreover, the microbiome's regulatory capabilities extend to specific immune cell populations such as TAMs and TILs. Gut microbial bile acids, such as deoxycholic acid, lithocholic acid, chenodeoxycholic acid, and cholic acid, have been observed to induce TAMs to adopt an anti-tumor phenotype, suggesting their potential role in glioma suppression.<sup>84,85</sup> Similarly, these metabolites can augment the cytotoxic activity of TILs, which may bolster their ability to target and destroy glioma cells. For instance, a peptide from Enterococcus hirae bacteriophage can be recognized as a tumor antigen by CD8<sup>+</sup> T cells via molecular mimicry. TIL-derived CD4<sup>+</sup> T cell clones (TCCs), such as TCC88, strongly crossrecognizes tumor antigens and microbiotaderived peptides, which are mainly derived from phyla Firmicutes, Proteobacteria the and Bacteroidota.<sup>68</sup> Such mechanisms underscore the microbiome's potential as a therapeutic target, offering pathways to modulate immune responses for more effective glioma treatment.<sup>86</sup>

### 5. Microbiota and efficacy of glioma therapies

Treatment options for gliomas usually include surgery, radiation therapy, chemotherapy, and

Table 2. Interplay between microbiome and current glioma therapy.

Glioma therapy/Event	Components	Effects and associations
Surgery Glioma detection	Gut microbiome: Escherichia coli,	Involvement in host NAD <sup>+</sup> metabolism, <sup>87</sup> potentially associated with assessment of glioma infiltrative margins <sup>88</sup>
lschemic stroke following craniotomy (stroke vs. healthy controls)	Saccharomyces cereviside, etc. Gut microbiome: Streptococcus ↑ Lactobacillus ↑ Escherichia ↑ Eubacterium ↓ Roseburia ↓, etc.	Potentially associated with risk factors of stroke, <sup>89</sup> consumption of dietary fibers improves stroke outcomes, likely <i>via</i> SCFAs production; stroke can in turn induce intestinal dysbiosis <sup>90,91</sup>
Chemotherapy and radiotherapy		
Immunosuppression	CSF: Listeria monocytogenes † Cryptococcus neoformans †	Exclusion of severe complications of glioma, e.g. leptomeningeal spread with carcinomatous meningitis <sup>92</sup>
Alveolo-interstitial pneumonia in a glioblastoma patient following temozolomide treatment <sup>93</sup>	Gut commensal bacteria e.g. Bifidobacterium genus: Bifidobacterium pseudolongum; Bifidobacterium animalis	Potential role of lung-gut axis: gut microbiome involve in defense against respiratory infections; respiratory infections in turn change gut microbiota composition <sup>94</sup>
Bevacizumab+temozolomide vs. temozolomide	Firmicutes ↑ Bacteroidetes ↑ Actinobacteria ↑ Bacteroidetes ↓ Cyanobacteria ↓	Treatment-induced significant changes in gut microbiota; may foster the development of new glioma therapies, e.g. FMT <sup>95</sup>
lmmunotherapy ICB	Gut microbiome: Akkermansia muciniphila Bifidobacterium longum Faecalibacterium spp., etc.	Key taxa potentially associated with a better response to ICB in glioma patients <sup>96</sup>
	Gut microbiome: Streptococcus ↑ Paecalibacterium ↑ Stenotrophomonas ↑ Faecalibacterium ↓ Unidentified Lephoepirgeoge	Potential biomarkers for predicting the occurrence of irAEs <sup>97</sup>
CAR-T therapy	Probiotics, e.g. Prevotella loescheii	Preventing and treating irAEs in treatment of malignant glioma <sup>98</sup>
Other glioma therapies UniPR1331	Gut microbiome as a whole	Profiling of main metabolites supports a major role in the <i>in-vivo</i> clearance of UniPR1331 <sup>99</sup>
Dietary polyphenols	Staphylococcus aureus ↓ Pseudomonas aeruainosa ↓	Anti-bacteria properties influence gut microbiota compositions <sup>100</sup>
Finasteride	Faecalibacterium spp.↓ Ruminococcaceae UCG-005↓ Alloprevotella↑	Alterations in gut microbiota composition was reported in post-finasteride patients <sup>101</sup>
Yi Qi Qu Yu Jie Du Fang (YYQQJDF) Taohong Siwu Decoction	Gut microbiome (uncertain) Gut microbiome as a whole ( <i>in vivo</i> metabolism in mice)	Potential key role in metabolism of core active ingredients <sup>102</sup> Active ingredients were yielded from serum of SPF mice, suggesting the role of gut microbiota in TSD metabolism <sup>103</sup>

YYQQJDF: a TCM prescription comprised of "Huangqi" (Hedysarum Multijugum Maxim.), "Chuangxiong" (Chuanxiong Rhizoma), "Banxia" (Arum Ternatum Thunb.), "Baihuashecao" (Hedyotis Difusae Herba), "Gancao" (Liquorice), "Shancigu" (Pseudobulbus Cremastrae Seu Pleiones), "Shichangpu" (Acoritataninowii Rhizoma) and "Taizishen" (Pseudostellariae Radix). Taohong Siwu Decoction: a TCM prescription composed of "Taoren" (Persicae Semen), "Honghua" (Carthami Flos), "Shudihuang" (Rehmanniae Radix Praeparata), "Danggui" (Angelicae Sinensis Radix), "Baishao" (Paeoniae Radix Alba), and "Chuanxiong" (Chuanxiong Rhizoma).

targeted therapies, depending on the type and grade of the tumor. Emerging research underscores the potential influence of the microbiome, particularly the gut microbiome, on the efficacy of glioma treatments. This body of work puts forward a bidirectional relationship where the microbiome may impact treatment outcomes, and glioma therapies may also modify the composition and abundance of the microbiota (Table 2).

## 5.1. Microbiota and surgical management of glioma

Gross total excision is the primary treatment strategy for adult and pediatric patients with low-grade glioma. The gut microbiome's role in glioma treatment may extend to its influence on surgical outcomes. For instance, gut microbes are associated with the detection of glioma through their contribution to host NAD<sup>+</sup> pools *via* bacteria-enabled deamidated pathways and *de novo* synthesis.<sup>87</sup> This interaction may be relevant in the context of intraoperative fluorescence lifetime imaging, which leverages time-resolved NAD(P)H fluorescence to glean metabolic insights from the tumor environment. Such metabolic information may be applied to assessing glioma's infiltrative margins, thereby informing microbiopsy and tumor resection strategies.<sup>88</sup>

Furthermore, the diversity of the gut microbiome can be significantly altered by post-surgical conditions, such as ischemic stroke following craniotomy.<sup>63</sup> Case-control studies have delineated shifts in microbial taxa – identifying 62 upregulated taxa (e.g., *Streptococcus, Lactobacillus, Escherichia*) and 29 downregulated taxa (e.g., *Eubacterium, Roseburia*) in stroke patients compared to healthy individuals. These alterations in microbial abundance are linked to glioma patient risk factors, including vessel injury during brain surgery and anesthesia-induced cardiovascular changes, indicating the possible connections between the microbiome, surgical interventions, and patient outcomes.<sup>89,90</sup>

Microbiome's potential impact on preoperative assessment and surgical outcome illuminates new pathways for therapeutic intervention. By targeting microbiome composition and functionality, there exists a promising avenue for enhancing the multimodal treatment of glioma, underscoring the need for further research in this emerging field.

## 5.2. Microbiota in chemotherapy and radiotherapy for glioma

In the management of glioma recurrences or progression, adjuvant therapies such as chemotherapy and radiotherapy are indispensable. These treatments, however, can profoundly affect the body's microbiome and immune system. The immunosuppression induced by chemotherapy and radiotherapy can diminish the diversity of the gut microbiota, consequently increasing the permeability of the BBB. This alteration is exemplified by a case report, where systemic immunosuppression from glioblastoma chemotherapy prompted the need for comprehensive molecular microbiology testing of cerebrospinal fluid (CSF). Such testing is crucial not only for typical bacterial and viral pathogens but also for pathogens linked with immunosuppression, such as *Listeria monocytogenes* and *Cryptococcus neoformans*. This comprehensive analysis can aid in excluding severe complications like leptomeningeal spread with carcinomatous meningitis.<sup>92</sup>

The effects of treatment on the microbiome extend to the gut-lung axis, illustrated by a case of a 56-year-old woman who developed alveolointerstitial pneumonia following treatment with temozolomide.93 This condition indicates the potential interactions between the lung microbiome and the oropharynx and gut microbiome. Furthermore, temozolomide treatment induces notable changes in gut microbial composition, with distinct differences between combination therapy with bevacizumab and monotherapy. Increased abundance of Firmicutes, Bacteroidetes. Actinobacteria and decreased abundance of Bacteroidetes and Cyanobacteria were found in the patients who received combination treatment.95

Conversely, the microbiome's composition and diversity may play a crucial role in shaping responses to chemotherapy and radiotherapy. The gut microbiota, through immunomodulation, may influence the efficacy of Temozolomide in treating glioma.<sup>104</sup> Pilot clinical trials have highlighted the potential of the gut microbiota in shielding cancer patients from radiation injuries.<sup>105</sup> The intratumoral microbiota also emerges as a promising player in modulating chemotherapy efficacy. Researchers found that intratumoral microbes, such Gammaproteobacteria, mediate tumor resistance to the chemotherapeutic drug gemcitabine by converting it into an inactive form *via* the bacterial enzyme cytidine deaminase, which may be associated with decreased efficacy of glioma therapy.<sup>106,107</sup> Comparatively, research on the role of intratumoral microbiome in radiotherapy remains scant.<sup>108,109</sup> This underscores the microbiome, particularly the gut and intratumoral microbiota, as potential therapeutic targets in glioma treatment, offering avenues to enhance treatment effects and reduce side effects.

## 5.3. Microbiota may modulate immunotherapy efficacy in glioma

Evolving evidence indicates the microbiota's possible role in modulating glioma patients' responses

to immune checkpoint blockades (ICBs), such as anti-PD-L1 and anti-CTLA-4 therapies. The potential of gut microbiome modulation through fecal microbial transplantation (FMT) to either initiate or suppress the response to ICB therapies have been confirmed in the treatment of epithelial tumors such as melanoma.<sup>110</sup> This intricate interplay suggests the gut microbiome's possible influence on the therapeutic landscape of gliomas.

Moreover, the advent of chimeric antigen receptor (CAR) T-cell therapy has introduced new therapeutic potentials for gliomas, evidenced by p32specific CAR T cells. Although phase I clinical trials for CAR T-cell therapy in glioblastoma have only recently commenced,<sup>111</sup> the gut microbiome's correlation with CAR-T therapy's effectiveness has been well-documented in B-cell lymphoma and leukemia.<sup>112</sup> However, the specific impact of the gut microbiome on CAR-T therapy for glioma remains to be fully elucidated.

The microbiota's role extends beyond modulating therapy effectiveness to influencing immunotoxicity. For instance, antibiotic use, which reduces gut microbiota diversity, has been associated with exacerbating CAR T cell-induced cytokine release syndrome and immune effector cell-associated neurotoxicity syndrome.<sup>113</sup> In this context, probiotics such as butyrate-producing *Prevotella loescheii* emerge as promising agents for preventing and treating immune-related adverse events (irAEs) in novel immunotherapies against malignant glioma.<sup>98</sup>

These examples indicate the underlying dual role of the microbiota in both enhancing and mitigating responses to advanced glioma therapies. As research continues to unravel the complex relationships between the microbiota and cancer therapy, it becomes increasingly clear that targeting the microbiota may offer novel avenues for improving therapeutic outcomes and managing treatmentrelated side effects.

### 5.4. Drug-microbiota interaction in glioma therapy

The microbiome, particularly the gut microbiota, may play a crucial role in both directly and indirectly affecting the metabolism and efficacy of drugs used in glioma treatment. The capability of gut microbiota to metabolize certain drugs may significantly influence their bioavailability and therapeutic outcomes. For instance, the gut microbiota is involved in the *in-vivo* clearance of UniPR1331, an antagonist of the Eph-ephrin system shown to be effective *in vivo* in a murine orthotopic model of GBM.<sup>99</sup> Additionally, natural compounds like polyphenols, which hold promise for the prevention and therapy of glioma, encounter poor intestinal absorption challenges attributed to the individual's intestinal microbiota content.<sup>114</sup>

Conversely, the administration of drugs can lead to modifications in the gut microbiota composition, which may impact the therapeutic efficacy of glioma treatments through the mechanisms described. Finasteride, known for its antioxidant and antiproliferative activity against glioblastoma cells, has demonstrated long-term effects on gut microbiota composition.<sup>115</sup>

The interplay between traditional Chinese medicine (TCM) and the microbiota also implicates the possible relationship between glioma treatments and the microbiome. While the herbal ingredients of TCM are not directly absorbed by the host, they can be transformed into active metabolites by the gut microbiota,<sup>116</sup> some of which exert anti-glioma effects. Specifically, derivatives of Yi Qi Qu Yu Jie Du Fang have been applied in GBM treatment, illustrating the potential therapeutic impact of microbiota-mediated metabolism.<sup>102</sup> The gut microbiota's ability to regulate the metabolism of compounds such as those in Taohong Siwu Decoction, which inhibits glioma cell proliferation and enhances autophagy, indicate the potential influence of the microbiome on the efficacy of glioma therapies.<sup>103</sup>

Although further prospective controlled trials to confirm this potential interaction are required, these examples suggest the possible role of the microbiome in the metabolism and efficacy of glioma treatments, indicating that the microbiome itself, through its dynamic interaction with therapeutic agents, may offer novel avenues for enhancing treatment outcomes or mitigating adverse effects.



**Figure 3.** Microbiome-based strategies for glioma. Front techniques for detection, extraction, and analysis of microbe and its derivatives are advancing, and models for TME simulation are being built, enabling in-depth investigation of the interaction between microbiome and glioma. These progresses also facilitate non-invasive diagnosis of glioma, where body fluids, stool and tumor samples are collected for multi-omics analysis, hopefully building a model for glioma diagnosis in combination with public datasets. Microbiota and its derivatives may act as not only a therapeutic target, but also a weapon for killing glioma cells due to their biocompatibility, immune characteristics, and/or cytotoxicity.

### 6. Novel microbiome-based strategies for glioma diagnosis and therapy

Microbiota, or microbial community, has shown its research and application potential in multiple medical fields in recent years, especially in the diagnosis and treatment of glioma. Researches on the microbiota provide new perspectives and strategies for understanding the pathogenesis of glioma, improving the accuracy of diagnosis, and developing new treatment methods (Figure 3).

### 6.1. Technical supports for unraveling microbiota-glioma interactions

In recent years, the field of medical research has significantly benefited from advancements in microbial detection and application, contributing to understanding glioma's pathogenesis, diagnosis, and treatment strategies. The past decade has seen remarkable progress in the development of techniques for detecting microbes and their interactions with host cells. For instance, luminogens with aggregation-induced emission properties have revolutionized rapid and specific imaging of bacteria and fungi, further enabling the monitoring of bacteria-cell interactions. This advancement may be pivotal for understanding the microbial influences within the glioma TME.<sup>117</sup> Moreover, bacterial extracellular vesicles (BEVs) have shown promising potential in clinical applications, serving as diagnostic markers or therapeutic agents. Effective methods for BEV separation and extraction have been proposed and established, highlighting the growing importance of microbial components in medical research.<sup>118,119</sup> Similarly, two-dimensional porphyrin-based Metal-Organic Frameworks (MOFs) and their derivatives have become increasingly vital for biomedical sensing and drug delivery, facilitating the detection and transport of microbial components.<sup>120</sup>

The exploration of microbiota-glioma interactions may also be enhanced by new techniques and models. Synergistic therapeutic models, such as 3D cultures simulating the GBM TME,<sup>121</sup> and the development of organoids for medulloblastoma, and GBM, high-grade glioma,<sup>122,123</sup> may enable in-depth studies on the intricate relationships among microbes, glioma, and the TME.<sup>124</sup> Additionally, 3D neuronavigation utilized during surgical resection may offer a comprehensive view of GBM evolution and heterogeneity,<sup>26</sup> while microbiome engineering introduces novel solutions for glioma therapy by improving the functions of the existing microbial community through the introduction of new microbes.<sup>125</sup>

These advancements underscore the microbiota's research and application potential across multiple medical fields, offering new perspectives and strategies for diagnosing and treating glioma. The continuous development of techniques to investigate the associations and interactions between microbiota and glioma may pave the way for innovative diagnostic and therapeutic approaches, demonstrating the potentially significant role of microbiota in the medical realm.

## 6.2. Employing microbiome in liquid biopsy of glioma

The complexity of glioma's biological characteristics has traditionally posed challenges for distinguishing it from other brain diseases using standard diagnostic methods. Consequently, there is an urgent need for early, precise, and noninvasive detection techniques. Emerging research has identified distinct differences in the microbiome's distribution between glioma patients and healthy individuals, implicating the microbiota's involvement in glioma pathogenesis.<sup>39</sup> Specific types of bacteria, viruses, and fungi have been associated with the occurrence of glioma, suggesting their potential as biomarkers.<sup>126</sup> Furthermore, variations in the microbiome can reflect the tumor's development stage and prognosis, offering new avenues for patient monitoring. For instance, differences in the gut microbiome between benign meningioma, malignant glioma, and healthy controls have been observed, indicating the microbiota's relevance in glioma's biological landscape.<sup>18</sup>

The concept of a microbiome-derived liquid biopsy has emerged from these findings. This promising approach includes not only microorganisms but also their derivatives, components, and metabolites as biomarkers, necessitating multi-omics analysis for more comprehensive evaluation. By analyzing the composition of a patient's blood, CSF, or gut microbiome, this method may complement traditional imaging and histopathological diagnoses, hopefully enhancing the accuracy of early diagnosis.<sup>127</sup> The analytical process incorporates whole-genome and transcriptome sequencing, alongside machine learning models based on microbial signatures,<sup>128</sup> further refined by nanopore sequencing technology.<sup>129</sup> Moreover, the correlation between oral microbiota and glioma introduces saliva as a potential medium for liquid biopsy,<sup>130</sup> although the relationship between salivary microbiota and glioma warrants further exploration. Additionally, public cancer genome sequence data analysis has identified bacteria associated with glioma, such as the detection of Mycobacterium tuberculosis complex in GBM samples, revealing associations with the sequencing center of the samples.<sup>131</sup>

These advancements highlight the microbiota's significant research and application potential in glioma, offering new strategies for understanding pathogenesis, improving diagnostic accuracy, and developing innovative treatment methods.

### **6.3.** Harnessing the microbiome for glioma prevention and therapy

Insights into the microbiota-glioma relationship may inform new preventive and therapeutic strategies for glioma. Specific examples of current clinical trials and proposed interventions have demonstrated that microbiota may act as not only a therapeutic target, but also a weapon for glioma growth inhibition (Table 3).

#### 6.3.1. Microbiota as a therapeutic target for glioma

In the realm of glioma prevention and therapy, directly modulating the composition of the microbiota presents a promising approach. This modulation can be achieved through dietary adjustments and changes in living habits. Notably, recent studies have highlighted the potential of probiotic combinations, such as *Bifidobacterium lactis* with *Lactobacillus plantarum*, in inhibiting glioma growth in mice. This effect is mediated by suppressing the PI3K/AKT pathway and regulating the composition and metabolism of the gut microbiota.<sup>132</sup>

Prebiotics may also play a crucial role in enhancing the response to ICBs and reducing immunotoxicities by fostering the growth of beneficial microbial species. For example, fructooligosaccharides (FOS) and

#### Table 3. Promising preventive and therapeutic approaches based on microbiota-glioma relationship.

Associations	Strategies
Microbiota as a therapeutic target	
-Bifidobacterium lactis and Lactobacillus plantarum inhibit glioma growth in mice	Probiotic combination <sup>132</sup>
•FOS and GOS improve the microbial profile and neuroplasticity through the gut-brain axis	Fostering the growth of beneficial microbial species via prebiotics <sup>133</sup>
<ul> <li>Intake of tea and vegetables may reduce the risk of glioma, while the intake of grains and processed me may increase the risk<sup>134</sup></li> </ul>	at Dietary interventions, e.g. nutritional adjuncts <sup>137</sup>
<ul> <li>Ketogenic diet is associated with improved outcomes of GBM patients<sup>135</sup></li> </ul>	
<ul> <li>A high-glucose drink was observed to increase CD4<sup>+</sup> T cells in GBM mice through regulation of the gr microbiota<sup>136</sup></li> </ul>	ut
•Oral microbiota is associated with glioma malignancy <sup>22</sup>	Attention to lifestyle factors, e.g. oral hygiene <sup>138</sup>
<ul> <li>Transferring healthy bacteria from a carefully screened donor to the recipient's colon ameliorates cognitiv impairment, suggesting interconnectedness of the gut, blood, and brain</li> </ul>	re Exogenous microbial therapies, e.g. FMT and LBPs <sup>139,140</sup>
Microbial derivatives as vehicles for targeted therapies	
<ul> <li>Oncolytic viruses can infect glioma cells and elicit anti-tumor immune responses in patients with malignal glioma<sup>141</sup></li> </ul>	nt Virotherapy
<ul> <li>The filamentous bacteriophages are effective vehicle for delivering displayed peptides toward the tume target<sup>142</sup></li> </ul>	or
•Virus-like particles hold promise for delivery of glioma mRNA-based therapeutics <sup>143</sup>	
•EVs derived from probiotics can be applied to glioma treatment through genetic manipulation and/or dietary interventions <sup>144</sup>	EVs as vehicles for targeted glioma therapy
•Nanovehicle mimicking tumor-colonizing bacteria overcomes chemoresistance in mouse models of established breast cancer <sup>145</sup>	
•Microglia is a potential therapeutic target which may be modulated by exosomes <sup>146</sup>	
Development of new drugs	
p28 secreted by <i>Pseudomonas aeruginosa</i> has been approved for glioma treatment by FDA <sup>147</sup> •Extracts from mushrooms exhibit anti-glioma potential <sup>148</sup>	Microbial derivatives or components

galactooligosaccharides (GOS) not only improve the microbial profile and neuroplasticity through the gutbrain axis, but also mitigate leaky gut syndrome in the distal colon.<sup>133</sup>

Additionally, dietary interventions have demonstrated potential for glioma prevention and treatment by controlling the nutrients supply to the intestinal microorganisms.<sup>149</sup> Statistics showed that intake of tea and vegetables may reduce the risk of glioma, whereas the intake of grains and processed meat may increase the risk.<sup>134</sup> Nutritional adjuncts like the ketogenic diet have shown multiple benefits at the synaptic level, including reducing oxidative stress and neuroinflammation. Its influence on the gut microbiome-brain axis has been associated with improved outcomes for glioblastoma patients.<sup>135</sup> A recent systematic review confirmed the therapeutic benefits of such nutritional adjuncts, indicating an increase in the median OS for both newly diagnosed and recurrent glioma cases compared to historical data. However, the absence of a statistically significant difference in OS highlights the need for further research to elucidate the mechanisms underlying these benefits.<sup>150</sup> Interestingly, a high-glucose drink, typically linked to metabolic diseases, was found to increase CD4<sup>+</sup> T cells in GBM mice through modulation of the gut microbiota,<sup>136</sup> suggesting the immunomodulatory potential of dietary interventions.<sup>137</sup>

The relationship between oral microbiota and glioma malignancy also underscores oral microbiota as a potential therapeutic target. Lifestyle factors, such as oral hygiene, could influence glioma occurrence and progression,<sup>138</sup> indicating a broader context in which microbiota modulation may impact glioma therapy.

Exogenous microbial therapies, like FMT, offer another avenue for microbiota modulation. FMT involves transferring healthy bacteria from a carefully screened donor to the recipient's colon, successfully treating *Clostridioides difficile* infections<sup>151</sup> and ameliorating cognitive impairment in Alzheimer's disease models by modulating intestinal macrophage activity and circulating blood inflammatory monocytes.<sup>139</sup> This technique sheds light on the interconnectedness of the gut, blood, and brain and its potential application in glioma therapy. Live biotherapeutic products (LBPs), either oral, topical, or injectable, represent a future direction for employing live organisms in disease prevention and treatment. By consuming harmful compounds and delivering therapeutic agents to the TME, LBPs may prevent antibioticinduced dysbiosis, modulate gut microbiota, and regulate intratumoral microbiota to combat solid tumors.<sup>140,152</sup>

## 6.3.2. Glioma therapies based on viral vectors and microbial derivatives

The development of glioma vaccines has marked a significant research milestone, with various vehicles discovered or synthesized to carry therapeutic effectors across the BBB. Virotherapy stands out as one of the most promising strategies, with several oncolytic viruses, such as herpesvirus, demonstrating the capability to infect glioma cells and elicit anti-tumor immune responses in patients with malignant glioma.<sup>141</sup> Additionally, filamentous bacteriophages, which specifically infect bacteria and are harmless to humans, have been recognized for their biocompatibility and high carrying capacity. These bacteriophages may be powerful carriers for glioma therapy, targeting tumor cells or molecules within the TME.<sup>153</sup> It is noteworthy that virus-like particles hold promise for mRNA delivery targeting APCs,<sup>154</sup> which may enhance the potency of glioma mRNA vaccine currently in clinical trials <sup>143</sup>

Extracellular vesicles (EVs), including outer membrane vesicles derived from bioengineered bacteria, have also shown promise as delivery tools for glioma therapy. These exosomes can be loaded with various tumor antigens to stimulate an anti-tumor response, successfully inhibiting the growth and metastasis of lung melanoma and colorectal cancer.<sup>155</sup> Glioblastoma cellderived exosomes, for instance, can promote chemotherapy resistance by delivering long noncoding RNAs that induce microglia to produce complement C5,<sup>146</sup> suggesting microglia as a potential therapeutic target to mitigate chemotherapy resistance, which may be modulated by exosomes. Moreover, EVs derived from probiotics like Akkermansia muciniphila offer promising avenues for glioma treatment through manipulation and/or genetic dietary interventions.<sup>144</sup> A novel type of nanovehicle tumor-colonized Fusobacterium mimicking nucleatum has been developed to overcome chemoresistance by targeting tumor-colonizing bacteria, a strategy tested in mouse models of established breast cancer.<sup>145</sup> Notably, the presence of tumor microbiome in glioma has been confirmed,<sup>24</sup> indicating that synthesized nanoparticles mimicking intratumoral microbes might also be applied to inhibit glioma growth and invasion. These vehicles can carry a wide range of cargos, including antibodies, peptides, proteins, as well as naturally occurring compounds like diosgenin and polyphenols, which have been shown to modify gut microbiota composition and may influence glioma development.<sup>114,156</sup>

In addition to vaccines, an increasing number of new drugs based on microbial derivatives or components are emerging. Bacterial peptides like p28, secreted by *Pseudomonas aeruginosa* and capable of crossing the BBB, have received FDA approval for glioma treatment.<sup>147</sup> Furthermore, active ingredients from fungi, including extracts from mushrooms like *Cantharellus cibarius*, *Coprinus comatus*, *Lycoperdon perlatum*, and *Lactarius delicious*, have shown anti-glioma potential by inhibiting cancer cell DNA synthesis.<sup>148</sup>

These advancements suggest the potential of microbial components and engineered delivery systems in advancing glioma diagnosis and treatment, although challenges in translating these findings from bench to bedside are also emerging. The specific roles of microorganisms in occurrence, development, and treatment of glioma still need to be confirmed, limiting their clinical application.<sup>126</sup> Additionally, strict control is required to avoid microbial contamination and degradation in samples.<sup>157</sup> Furthermore, moving from laboratory research to clinical application requires extensive clinical trial validations. Addressing these challenges calls for more efforts in in-depth research, standardization of preprocessing methods, and further evaluation of feasibility and effectiveness of these strategies.

#### 7. Conclusions

This review has illuminated how leveraging the microbiota, particularly from the gut, could possibly advance glioma management, introducing noninvasive diagnostics, targeted therapies, and even preventive strategies. However, the focus on glioblastoma and gut microbiota to date

represents just the tip of the iceberg. The most critical areas for future research include the broader spectrum of gliomas and the diverse microbiota across different body sites. The interconnectedness of microbiota across these sites hints at a complex, systemic relationship with glioma, urging a more holistic approach to future studies.<sup>158</sup> Moreover, the exploration of nonbacterial microbiota components, such as viruses and fungi, promises to deepen our understanding of the microbiome-glioma nexus, unveiling new targets for intervention. As we move forward, it is imperative to identify the gaps of current literature, including the nuances of pediatric versus adult gliomas, tailoring interventions to the unique microbiota profiles of these populations.<sup>159</sup> New methodologies or approaches for liquid biopsy and glioma therapy, such as multi-omics analysis and microbiomebased therapeutics, require improvement and evaluation for further application. In conclusion, the burgeoning field of microbiota research in glioma heralds a new era in oncology, which may promise more effective, personalized, and less invasive options for diagnosis and treatment. The synergy between microbiota and glioma research not only offers hope for breakthroughs in glioma management, but also emphasizes the need for interdisciplinary research, combining oncology, microbiology, and neurology to unravel the complexities of human health and disease.

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#### **Author contributions**

Wenhui Wang and Zihao Ou wrote and edited the manuscript; Wenhui Wang collected published papers and generated the figures; and Zihao Ou and Lei Zheng revised the manuscript. All authors have read and approved the article.

#### References

- Weller M, Wick W, Aldape K, Brada M, Berger M, Pfister SM, Nishikawa R, Rosenthal M, Wen PY, Stupp R, et al. Glioma. Nat Rev Dis Primers. 2015;1 (1):15017. doi:10.1038/nrdp.2015.17.
- Ostrom QT, Bauchet L, Davis FG, Deltour I, Fisher JL, Langer CE, Pekmezci M, Schwartzbaum JA, Turner MC, Walsh KM, et al. The epidemiology of glioma in adults: a "state of the science" review. Neuro Oncol. 2014;16(7):896–913. doi:10.1093/neuonc/ nou087.
- 3. Ostrom QT, Cote DJ, Ascha M, Kruchko C, Barnholtz-Sloan JS. Adult glioma incidence and survival by race or ethnicity in the United States from 2000 to 2014. JAMA Oncol. 2018;4(9):1254–1262. doi:10.1001/jamaoncol. 2018.1789.
- Birzu C, French P, Caccese M, Cerretti G, Idbaih A, Zagonel V, Lombardi G. Recurrent glioblastoma: from molecular landscape to new treatment perspectives. Cancers (Basel). 2020;13(1):47. doi:10.3390/can cers13010047.
- Psimaras D, Bonnet C, Heinzmann A, Cárdenas G, Hernández José Luis S, Tungaria A, Behari S, Lacrois D, Mokhtari K, Karantoni E, et al. Solitary tuberculous brain lesions: 24 new cases and a review of the literature. Rev Neurol (Paris). 2014;170(6– 7):454–463. doi:10.1016/j.neurol.2013.12.008.
- Parbel S, Vlaho S, Gebhardt B, Porto L, Hattingen E, Klingebiel T, Böhles H, Kieslich M. Diagnostic difficulties in encephalitis and glioma. Klin Padiatr. 2007;219 (4):222–224. doi:10.1055/s-2006-933521.
- 7. Noorani I, de la Rosa J. Breaking barriers for glioblastoma with a path to enhanced drug delivery. Nat Commun. 2023;14(1):5909. doi:10.1038/s41467-023-41694-9.
- Riviere-Cazaux C, Carlstrom LP, Rajani K, Munoz-Casabella A, Rahman M, Gharibi-Loron A, Brown DA, Miller KJ, White JJ, Himes BT, et al. Bloodbrain barrier disruption defines the extracellular metabolome of live human high-grade gliomas. Commun Biol. 2023;6(1):653. doi:10.1038/s42003-023-05035-2.
- 9. Nicholson JG, Fine HA. Diffuse glioma heterogeneity and its therapeutic implications. Cancer Discov. 2021;11(3):575–590. doi:10.1158/2159-8290.Cd-20-1474.
- 10. Hou K, Wu ZX, Chen XY, Wang J-Q, Zhang D, Xiao C, Zhu D, Koya JB, Wei L, Li J, et al. Microbiota in health

and diseases. Signal Transduct Target Ther. 2022;7 (1):135. doi:10.1038/s41392-022-00974-4.

- Elinav E, Garrett WS, Trinchieri G, Wargo J. The cancer microbiome. Nat Rev Cancer. 2019;19(7):371–376. doi:10.1038/s41568-019-0155-3.
- Kunze R, Fischer S, Marti HH, Preissner KT. Brain alarm by self-extracellular nucleic acids: from neuroinflammation to neurodegeneration. J Biomed Sci. 2023;30(1):64. doi:10.1186/s12929-023-00954-y.
- Needham BD, Kaddurah-Daouk R, Mazmanian SK. Gut microbial molecules in behavioural and neurodegenerative conditions. Nat Rev Neurosci. 2020;21 (12):717–731. doi:10.1038/s41583-020-00381-0.
- Arabi TZ, Alabdulqader AA, Sabbah BN, Ouban A. Brain-inhabiting bacteria and neurodegenerative diseases: the "brain microbiome" theory. Front Aging Neurosci. 2023;15:1240945. doi:10.3389/fnagi.2023. 1240945.
- 15. Ma Q, Yao C, Wu Y, Wang H, Fan Q, Yang Q, Xu J, Dai H, Zhang Y, Xu F, et al. Neurological disorders after severe pneumonia are associated with translocation of endogenous bacteria from the lung to the brain. Sci Adv. 2023;9(42):eadi0699. doi:10.1126/sciadv.adi0699.
- 16. Zhao C, Kuraji R, Ye C, Gao L, Radaic A, Kamarajan P, Taketani Y, Kapila YL. Nisin a probiotic bacteriocin mitigates brain microbiome dysbiosis and alzheimer's disease-like neuroinflammation triggered by periodontal disease. J Neuroinflamm. 2023;20(1):228. doi:10. 1186/s12974-023-02915-6.
- 17. Mehrian-Shai R, Reichardt JKV, Harris CC, Toren A. The gut-brain axis, paving the way to brain cancer. Trends Cancer. 2019;5(4):200–207. doi:10.1016/j.tre can.2019.02.008.
- Jiang H, Zeng W, Zhang X, Pei Y, Zhang H, Li Y. The role of gut microbiota in patients with benign and malignant brain tumors: a pilot study. Bioengineered. 2022;13(3):7847-7859. doi:10.1080/21655979.2022. 2049959.
- Wang S, Yin F, Guo Z, Li R, Sun W, Wang Y, Geng Y, Sun C, Sun D. Association between gut microbiota and glioblastoma: a Mendelian randomization study. Front Genet. 2023;14:1308263. doi:10.3389/fgene.2023. 1308263.
- 20. Wu Y, Cheng X, Jiang G, Tang H, Ming S, Tang L, Lu J, Guo C, Shan H, Huang X, et al. Altered oral and gut microbiota and its association with SARS-CoV-2 viral load in COVID-19 patients during hospitalization. Npj Biofilms Microbiomes. 2021;7(1):61. doi:10.1038/ s41522-021-00232-5.
- Gregory T, Knight S, Aaroe A, O'Brien BJ, Patel CB, Weathers SPS, Majd N, Puduvalli VK, Kamiya-Matsuoka C. Analysis of tumor progression among patients with glioma after COVID-19 infection. J Clin Oncol. 2023;41(16\_suppl):2041–2041. doi:10.1200/ JCO.2023.41.16\_suppl.2041.
- 22. Wen Y, Feng L, Wang H, Zhou H, Li Q, Zhang W, Wang M, Li Y, Luan X, Jiang Z, et al. Association

between oral microbiota and human brain glioma grade: a case-control study. Front Microbiol. 2021;12:746568. doi:10.3389/fmicb.2021.746568.

- Bao L, Zhang C, Dong J, Zhao L, Li Y, Sun J. Oral microbiome and SARS-CoV-2: beware of lung Co-infection. Front Microbiol. 2020;11:1840. doi:10. 3389/fmicb.2020.01840.
- 24. Zhao J, He D, Lai HM, Xu Y, Luo Y, Li T, Liang J, Yang X, Guo L, Ke Y, et al. Comprehensive histological imaging of native microbiota in human glioma. J Biophotonics. 2022;15(4):e202100351. doi:10.1002/ jbio.202100351.
- 25. He D, Li T, Yang X, Xu Y, Sun H. Sudan black B treatment for reducing autofluorescence in human glioma tissue and improving fluorescent signals of bacterial LPS staining. J Biophotonics. 2023;16(5): e202200357. doi:10.1002/jbio.202200357.
- 26. Mathur R, Wang Q, Schupp PG, Nikolic A, Hilz S, Hong C, Grishanina NR, Kwok D, Stevers NO, Jin Q, et al. Glioblastoma evolution and heterogeneity from a 3D whole-tumor perspective. Cell. 2024;187(2):446– 463.e16. doi:10.1016/j.cell.2023.12.013.
- 27. Smirnova OA, Ivanova ON, Fedyakina IT, Yusubalieva GM, Baklaushev VP, Yanvarev DV, Kechko OI, Mitkevich VA, Vorobyev PO, Fedorov VS, et al. SARS-CoV-2 establishes a productive infection in hepatoma and glioblastoma multiforme cell lines. Cancers (Basel). 2023;15(3). doi:10.3390/cancers15030632.
- Khan I, Hatiboglu MA. Can COVID-19 induce glioma tumorogenesis through binding cell receptors? Med Hypotheses. 2020;144:110009. doi:10.1016/j.mehy. 2020.110009.
- 29. Ahmed S, Busetti A, Fotiadou P, Vincy Jose N, Reid S, Georgieva M, Brown S, Dunbar H, Beurket-Ascencio G, Delday MI, et al. In vitro characterization of gut microbiota-derived bacterial strains with neuroprotective properties. Front Cell Neurosci. 2019;13. doi:10. 3389/fncel.2019.00402.
- Weissenberger J, Loeffler S, Kappeler A, Kopf M, Lukes A, Afanasieva TA, Aguzzi A, Weis J. IL-6 is required for glioma development in a mouse model. Oncogene. 2004;23(19):3308–3316. doi:10.1038/sj.onc. 1207455.
- Hambardzumyan D, Gutmann DH, Kettenmann H. The role of microglia and macrophages in glioma maintenance and progression. Nat Neurosci. 2016;19 (1):20–27. doi:10.1038/nn.4185.
- 32. Rodríguez J, De Santis Arévalo J, Dennis VA, Rodríguez AM, Giambartolomei GH. Bystander activation of microglia by Brucella abortus-infected astrocytes induces neuronal death via IL-6 trans-signaling. Front Immunol. 2023;14:1343503. doi:10.3389/fimmu. 2023.1343503.
- 33. Fassler M, Weissberg I, Levy N, Diaz-Griffero F, Monsonego A, Friedman A, Taube R. Preferential lentiviral targeting of astrocytes in the central nervous

system. PLOS ONE. 2013;8(10):e76092. doi:10.1371/ journal.pone.0076092.

- 34. Humbel M, Ramosaj M, Zimmer V, Regio S, Aeby L, Moser S, Boizot A, Sipion M, Rey M, Déglon N. Maximizing lentiviral vector gene transfer in the CNS. Gene Ther. 2021;28(1):75–88. doi:10.1038/s41434-020-0172-6.
- 35. Ahmad F, Hyvärinen A, Pirinen A, Olsson V, Rummukainen J, Immonen A, Närväinen J, Tuunanen P, Liimatainen T, Kärkkäinen V, et al. Lentivirus vector-mediated genetic manipulation of oncogenic pathways induces tumor formation in rabbit brain. Mol Med Rep. 2021;23(6). doi:10.3892/mmr. 2021.12061.
- 36. Nandwana V, Nandwana NK, Das Y, Saito M, Panda T, Das S, Almaguel F, Hosmane NS, Das BC. The role of microbiome in brain development and neurodegenerative diseases. Molecules. 2022;27(11):3402. doi:10.3390/ molecules27113402.
- 37. Mueller S, Kline C, Franson A, van der Lugt J, Prados M, Waszak SM, Plasschaert SLA, Molinaro AM, Koschmann C, Nazarian J. Rational combination platform trial design for children and young adults with diffuse midline glioma: a report from PNOC. Neuro Oncol. 2024;26(Supplement\_2): S125–S135. doi:10.1093/neuonc/noad181.
- 38. Manterola L, Guruceaga E, Gállego Pérez-Larraya J, González-Huarriz M, Jauregui P, Tejada S, Diez-Valle R, Segura V, Samprón N, Barrena C, et al. A small noncoding RNA signature found in exosomes of GBM patient serum as a diagnostic tool. Neuro Oncol. 2014;16(4):520–527. doi:10.1093/neuonc/not218.
- 39. Dono A, Nickles J, Rodriguez-Armendariz AG, McFarland BC, Ajami NJ, Ballester LY, Wargo JA, Esquenazi Y. Glioma and the gut-brain axis: opportunities and future perspectives. Neurooncol Adv. 2022;4 (1):vdac054. doi:10.1093/noajnl/vdac054.
- Herbreteau A, Aubert P, Croyal M, Naveilhan P, Billon-Crossouard S, Neunlist M, Delneste Y, Couez D, Aymeric L. Late-stage glioma is associated with deleterious alteration of gut bacterial metabolites in mice. Metabolites. 2022;12(4):290. doi:10.3390/metabo12040290.
- 41. Vogel MME, Wagner A, Gempt J, Krenzlin H, Zeyen T, Drexler R, Voss M, Nettekoven C, Abboud T, Mielke D, et al. Impact of the SARS-CoV-2 pandemic on the survival of patients with high-grade glioma and best practice recommendations. Sci Rep. 2023;13(1):2766. doi:10.1038/s41598-023-29790-8.
- 42. Nejman D, Livyatan I, Fuks G, Gavert N, Zwang Y, Geller LT, Rotter-Maskowitz A, Weiser R, Mallel G, Gigi E, et al. The human tumor microbiome is composed of tumor type-specific intracellular bacteria. Science. 2020;368(6494):973–980. doi:10.1126/science.aay9189.
- Zhao YF, Wei DN, Tang Y. Gut microbiota regulate astrocytic functions in the brain: possible therapeutic consequences. Curr Neuropharmacol. 2021;19 (8):1354–1366. doi:10.2174/1570159x19666210215123239.

- 44. Dicks LMT. Gut bacteria and neurotransmitters. Microorganisms. 2022;10(9):1838. doi:10.3390/microor ganisms10091838.
- D'Alessandro G, Lauro C, Quaglio D, Ghirga F, Botta B, Trettel F, Limatola C. Neuro-signals from gut microbiota: perspectives for brain glioma. Cancers (Basel). 2021;13(11):2810. doi:10.3390/cancers13112810.
- Murciano-Brea J, Garcia-Montes M, Geuna S, Herrera-Rincon C. Gut microbiota and neuroplasticity. Cells. 2021;10(8):2084. doi:10.3390/cells10082084.
- 47. Lv K, Cao X, Wang R, Du P, Fu J, Geng D, Zhang J. Neuroplasticity of glioma patients: brain structure and topological network. Front Neurol. 2022;13:871613. doi:10.3389/fneur.2022.871613.
- Farzi A, Fröhlich EE, Holzer P. Gut microbiota and the neuroendocrine system. Neurotherapeutics. 2018;15 (1):5-22. doi:10.1007/s13311-017-0600-5.
- González-Mora AM, Garcia-Lopez P. Estrogen receptors as molecular targets of endocrine therapy for glioblastoma. Int J Mol Sci. 2021;22(22):12404. doi:10. 3390/ijms222212404.
- 50. Banks WA, Gray AM, Erickson MA, Salameh TS, Damodarasamy M, Sheibani N, Meabon JS, Wing EE, Morofuji Y, Cook DG, et al. Lipopolysaccharideinduced blood-brain barrier disruption: roles of cyclooxygenase, oxidative stress, neuroinflammation, and elements of the neurovascular unit. J Neuroinflamm. 2015;12(1):223. doi:10.1186/s12974-015-0434-1.
- DeGruttola AK, Low D, Mizoguchi A, Mizoguchi E. Current understanding of dysbiosis in disease in human and animal models. Inflamm Bowel Dis. 2016;22 (5):1137–1150. doi:10.1097/mib.000000000000750.
- 52. Madison A, Kiecolt-Glaser JK. Stress, depression, diet, and the gut microbiota: human-bacteria interactions at the core of psychoneuroimmunology and nutrition. Curr Opin Behav Sci. 2019;28:105–110. doi:10.1016/j. cobeha.2019.01.011.
- 53. Cubillos-Ruiz A, Alcantar MA, Donghia NM, Cárdenas P, Avila-Pacheco J, Collins JJ. An engineered live biotherapeutic for the prevention of antibiotic-induced dysbiosis. Nat Biomed Eng. 2022;6 (7):910–921. doi:10.1038/s41551-022-00871-9.
- 54. Fan Y, Su Q, Chen J, Wang Y, He S. Gut microbiome alterations affect glioma development and Foxp3 expression in tumor microenvironment in mice. Front Oncol. 2022;12:836953. doi:10.3389/fonc.2022. 836953.
- 55. Yao Y, Cai X, Fei W, Ye Y, Zhao M, Zheng C. The role of short-chain fatty acids in immunity, inflammation and metabolism. Crit Rev Food Sci Nutr. 2022;62 (1):1–12. doi:10.1080/10408398.2020.1854675.
- Abdul Rashid K, Ibrahim K, Wong JHD, Mohd Ramli N. Lipid alterations in glioma: a systematic review. Metabolites. 2022;12(12):1280. doi:10.3390/ metabol2121280.

- 57. Yan J, Charles JF. Gut microbiota and IGF-1. Calcif Tissue Int. 2018;102(4):406-414. doi:10.1007/s00223-018-0395-3.
- Aleti G, Troyer EA, Hong S. G protein-coupled receptors: a target for microbial metabolites and a mechanistic link to microbiome-immune-brain interactions. Brain Behav Immun Health. 2023;32:100671. doi:10.1016/j.bbih.2023.100671.
- Jin C, Chen H, Xie L, Zhou Y, Liu L-L, Wu J. GPCRs involved in metabolic diseases: pharmacotherapeutic development updates. Acta Pharmacol Sin. 2024;45 (7):1321–1336. doi:10.1038/s41401-023-01215-2.
- Schoeler M, Caesar R. Dietary lipids, gut microbiota and lipid metabolism. Rev Endocr Metab Disord. 2019;20(4):461–472. doi:10.1007/s11154-019-09512-0.
- 61. Dehhaghi M, Kazemi Shariat Panahi H, Heng B, Guillemin GJ. The gut microbiota, kynurenine pathway, and immune system interaction in the development of brain cancer. Front Cell Dev Biol. 2020;8:562812. doi:10.3389/fcell.2020.562812.
- Agus A, Clément K, Sokol H. Gut microbiota-derived metabolites as central regulators in metabolic disorders. Gut. 2021;70(6):1174–1182. doi:10.1136/gutjnl-2020-323071.
- Gwak MG, Chang SY. Gut-brain connection: microbiome, gut barrier, and environmental sensors. Immune Netw. 2021;21(3):e20. doi:10.4110/in.2021.21. e20.
- Chelakkot C, Ghim J, Ryu SH. Mechanisms regulating intestinal barrier integrity and its pathological implications. Exp Mol Med. 2018;50(8):1–9. doi:10. 1038/s12276-018-0126-x.
- Shi L, Jin L, Huang W. Bile acids, intestinal barrier dysfunction, and related diseases. Cells. 2023;12 (14):1888. doi:10.3390/cells12141888.
- 66. Caminero A, Guzman M, Libertucci J, Lomax AE. The emerging roles of bacterial proteases in intestinal diseases. Gut Microbes. 2023;15(1):2181922. doi:10. 1080/19490976.2023.2181922.
- Ghosh SS, Wang J, Yannie PJ, Ghosh S. Intestinal barrier dysfunction, LPS translocation, and disease development. J Endocr Soc. 2020;4(2):bvz039. doi:10. 1210/jendso/bvz039.
- 68. Naghavian R, Faigle W, Oldrati P, Wang J, Toussaint NC, Qiu Y, Medici G, Wacker M, Freudenmann LK, Bonté P-E, et al. Microbial peptides activate tumour-infiltrating lymphocytes in glioblastoma. Nature. 2023;617(7962):807–817. doi:10. 1038/s41586-023-06081-w.
- Hajam IA, Dar PA, Shahnawaz I, Jaume JC, Lee JH. Bacterial flagellin—a potent immunomodulatory agent. Exp Mol Med. 2017;49(9):e373-e373. doi:10.1038/ emm.2017.172.
- Wang J, Liu Y, Zhang A, Yu W, Lei Q, Xiao B, Luo Z. Investigational microbiological therapy for glioma. Cancers. 2022;14(23):5977. doi:10.3390/can cers14235977.

- Kim CH. Control of lymphocyte functions by gut microbiota-derived short-chain fatty acids. Cell Mol Immunol. 2021;18(5):1161–1171. doi:10.1038/s41423-020-00625-0.
- 72. Ahmed S, Busetti A, Fotiadou P, Vincy Jose N, Reid S, Georgieva M, Brown S, Dunbar H, Beurket-Ascencio G, Delday MI, et al. In vitro characterization of gut microbiota-derived bacterial strains with neuroprotective properties. Front Cell Neurosci. 2019;13:402. doi:10.3389/fncel.2019.00402.
- 73. Yang F, He Z, Duan H, Zhang D, Li J, Yang H, Dorsey JF, Zou W, Nabavizadeh SA, Bagley SJ, et al. Synergistic immunotherapy of glioblastoma by dual targeting of IL-6 and CD40. Nat Commun. 2021;12 (1):3424. doi:10.1038/s41467-021-23832-3.
- 74. Erny D, Hrabě de Angelis AL, Jaitin D, Wieghofer P, Staszewski O, David E, Keren-Shaul H, Mahlakoiv T, Jakobshagen K, Buch T, et al. Host microbiota constantly control maturation and function of microglia in the CNS. Nat Neurosci. 2015;18(7):965–977. doi:10. 1038/nn.4030.
- 75. D'Alessandro G, Antonangeli F, Marrocco F, Porzia A, Lauro C, Santoni A, Limatola C. Gut microbiota alterations affect glioma growth and innate immune cells involved in tumor immunosurveillance in mice. Eur J Immunol. 2020;50(5):705–711. doi:10.1002/eji. 201948354.
- 76. Chen C, Huang Z, Huang P, Li K, Zeng J, Wen Y, Li B, Zhao J, Wu P. Urogenital microbiota: Potentially important determinant of PD-L1 expression in male patients with non-muscle invasive bladder cancer. BMC Microbiol. 2022;22(1):7. doi:10.1186/s12866-021-02407-8.
- 77. Lin B, Ye Z, Ye Z, Wang M, Cao Z, Gao R, Zhang Y. Gut microbiota in brain tumors: an emerging crucial player. CNS Neurosci Ther. 2023;29(Suppl 1):84–97. doi:10. 1111/cns.14081.
- Togashi Y, Shitara K, Nishikawa H. Regulatory T cells in cancer immunosuppression — implications for anticancer therapy. Nat Rev Clin Oncol. 2019;16 (6):356–371. doi:10.1038/s41571-019-0175-7.
- Jin UH, Karki K, Cheng Y, Michelhaugh SK, Mittal S, Safe S. The aryl hydrocarbon receptor is a tumor suppressor-like gene in glioblastoma. J Biol Chem. 2019;294(29):11342-11353. doi:10.1074/jbc.RA119. 008882.
- 80. Fan Z, Tang P, Li C, Yang Q, Xu Y, Su C, Li L. Fusobacterium nucleatum and its associated systemic diseases: epidemiologic studies and possible mechanisms. J Oral Microbiol. 2023;15(1):2145729. doi:10.1080/20002297.2022.2145729.
- Mempel TR, Lill JK, Altenburger LM. How chemokines organize the tumour microenvironment. Nat Rev Cancer. 2024;24(1):28–50. doi:10.1038/s41568-023-00635-w.
- 82. Elemam NM, Talaat IM, Maghazachi AA. CXCL10 chemokine: a critical player in RNA and DNA viral

infections. Viruses. 2022;14(11):2445. doi:10.3390/ v14112445.

- 83. Cao Y, Xia H, Tan X, Shi C, Ma Y, Meng D, Zhou M, Lv Z, Wang S, Jin Y. Intratumoural microbiota: a new frontier in cancer development and therapy. Sig Transduct Target Ther. 2024;9(1):15. doi:10.1038/ s41392-023-01693-0.
- Michaels M, Madsen KL. Immunometabolism and microbial metabolites at the gut barrier: lessons for therapeutic intervention in inflammatory bowel disease. Mucosal Immunol. 2023;16(1):72–85. doi:10. 1016/j.mucimm.2022.11.001.
- 85. Wang L, Gong Z, Zhang X, Zhu F, Liu Y, Jin C, Du X, Xu C, Chen Y, Cai W, et al. Gut microbial bile acid metabolite skews macrophage polarization and contributes to high-fat diet-induced colonic inflammation. Gut Microbes. 2020;12(1):1–20. doi:10.1080/19490976. 2020.1819155.
- 86. Yang W, Cong Y. Gut microbiota-derived metabolites in the regulation of host immune responses and immune-related inflammatory diseases. Cell Mol Immunol. 2021;18(4):866–877. doi:10.1038/s41423-021-00661-4.
- 87. Ren Z, Xu Y, Li T, Sun W, Tang Z, Wang Y, Zhou K, Li J, Ding Q, Liang K, et al. NAD(+) and its possible role in gut microbiota: insights on the mechanisms by which gut microbes influence host metabolism. Anim Nutr. 2022;10:360–371. doi:10.1016/j.aninu.2022.06.009.
- 88. Alfonso-García A, Zhou X, Bec J, Anbunesan SN, Fereidouni F, Jin L-W, Lee HS, Bloch O, Marcu L. First in patient assessment of brain tumor infiltrative margins using simultaneous time-resolved measurements of 5-ala-induced PpIX fluorescence and tissue autofluorescence. J Biomed Opt. 2022;27(2). doi:10. 1117/1.Jbo.27.2.020501.
- 89. Ghosh MK, Chakraborty D, Sarkar S, Bhowmik A, Basu M. The interrelationship between cerebral ischemic stroke and glioma: a comprehensive study of recent reports. Signal Transduct Targeted Ther. 2019;4 (1):42. doi:10.1038/s41392-019-0075-4.
- Peh A, O'Donnell JA, Broughton BRS, Marques FZ. Gut microbiota and their metabolites in stroke: a double-edged sword. Stroke. 2022;53(5):1788–1801. doi:10.1161/strokeaha.121.036800.
- Komlodi-Pasztor E, Gilbert MR, Armstrong TS. Diagnosis and management of stroke in adults with primary brain tumor. Curr Oncol Rep. 2022;24 (10):1251–1259. doi:10.1007/s11912-022-01280-6.
- 92. Neagu A, Niculae CM, Lăpădat I, Hristea A. Challenges in the diagnosis of leptomeningeal dissemination of glioblastoma in a patient with fever and xanthochromic CSF: a case report. Rom J Intern Med. 2023;61 (3):163–166. doi:10.2478/rjim-2023-0010.
- 93. Guilleminault L, Carré P, de Luca K, Beau Salinas F, Autret-Leca E, Narciso B, Diot P. Pneumonie alvéolointerstitielle au témozolomide. Rev Mal Respir. 2008;25 (7):880–884. doi:10.1016/s0761-8425(08)74357-2.

- 94. Sencio V, Machado MG, Trottein F. The lung-gut axis during viral respiratory infections: the impact of gut dysbiosis on secondary disease outcomes. Mucosal Immunol. 2021;14(2):296–304. doi:10.1038/s41385-020-00361-8.
- 95. Zhu J, Su J. Alterations of the gut microbiome in recurrent malignant gliomas patients received Bevacizumab and temozolomide combination treatment and temozolomide monotherapy. Indian J Microbiol. 2022;62 (1):23–31. doi:10.1007/s12088-021-00962-2.
- 96. Blake SJ, Wolf Y, Boursi B, Lynn DJ. Role of the microbiota in response to and recovery from cancer therapy. Nat Rev Immunol. 2024;24(5):308–325. doi:10.1038/ s41577-023-00951-0.
- 97. Liu W, Ma F, Sun B, Liu Y, Tang H, Luo J, Chen H, Luo Z. Intestinal microbiome associated with immune-related adverse events for patients treated with anti-PD-1 inhibitors, a real-world study. Front Immunol. 2021;12:756872. doi:10.3389/fimmu.2021. 756872.
- 98. Chen Y, Liu Y, Wang Y, Chen X, Wang C, Chen X, Yuan X, Liu L, Yang J, Zhou X, et al. Prevotellaceae produces butyrate to alleviate PD-1/PD-L1 inhibitorrelated cardiotoxicity via PPARα-CYP4X1 axis in colonic macrophages. J Exp Clin Cancer Res. 2022;41(1):1. doi:10.1186/s13046-021-02201-4.
- 99. Ferlenghi F, Castelli R, Scalvini L, Giorgio C, Corrado M, Tognolini M, Mor M, Lodola A, Vacondio F. Drug-gut microbiota metabolic interactions: the case of UniPR1331, selective antagonist of the Eph-ephrin system, in mice. J Pharm Biomed Anal. 2020;180:113067. doi:10.1016/j.jpba.2019.113067.
- 100. Wang X, Qi Y, Zheng H. Dietary polyphenol, gut microbiota, and health benefits. Antioxidants (Basel). 2022;11(6):1212. doi:10.3390/antiox11061212.
- 101. Borgo F, Macandog AD, Diviccaro S, Falvo E, Giatti S, Cavaletti G, Melcangi RC. Alterations of gut microbiota composition in post-finasteride patients: a pilot study. J Endocrinol Invest. 2021;44(6):1263–1273. doi:10. 1007/s40618-020-01424-0.
- 102. Liang C, Zhang B, Li R, Guo S, Fan X. Network pharmacology -based study on the mechanism of traditional Chinese medicine in the treatment of glioblastoma multiforme. BMC Complement Med Ther. 2023;23 (1):342. doi:10.1186/s12906-023-04174-7.
- 103. Feng S, Wan Q, Wu W, Zhang C, Lu H, Lu X. Effect of gut microbiome regulated Taohong Siwu decoction metabolism on glioma cell phenotype. Front Cell Infect Microbiol. 2023;13:1192589. doi:10.3389/fcimb. 2023.1192589.
- 104. Hou X, Du H, Deng Y, Wang H, Liu J, Qiao J, Liu W, Shu X, Sun B, Liu Y, et al. Gut microbiota mediated the individualized efficacy of temozolomide via immunomodulation in glioma. J Transl Med. 2023;21(1):198. doi:10.1186/s12967-023-04042-5.
- 105. Yi Y, Lu W, Shen L, Wu Y, Zhang Z. The gut microbiota as a booster for radiotherapy: novel insights into

radio-protection and radiation injury. Exp Hematol Oncol. 2023;12(1):48. doi:10.1186/s40164-023-00410-5.

- 106. Geller LT, Barzily-Rokni M, Danino T, Jonas OH, Shental N, Nejman D, Gavert N, Zwang Y, Cooper ZA, Shee K, et al. Potential role of intratumor bacteria in mediating tumor resistance to the chemotherapeutic drug gemcitabine. Science. 2017;357 (6356):1156–1160. doi:10.1126/science.aah5043.
- 107. Kim MM, Camelo-Piragua S, Schipper M, Tao Y, Normolle D, Junck L, Mammoser A, Betz BL, Cao Y, Kim CJ, et al. Gemcitabine plus radiation therapy for high-grade glioma: long-term results of a phase 1 dose-escalation study. Int J Radiat Oncol Biol Phys. 2016;94(2):305–311. doi:10.1016/j.ijrobp.2015.10.032.
- 108. Xue C, Chu Q, Zheng Q, Yuan X, Su Y, Bao Z, Lu J, Li L. Current understanding of the intratumoral microbiome in various tumors. Cell Rep Med. 2023;4(1):100884. doi:10.1016/j.xcrm.2022.100884.
- 109. Yang L, Li A, Wang Y, Zhang Y. Intratumoral microbiota: roles in cancer initiation, development and therapeutic efficacy. Signal Transduct Targeted Ther. 2023;8(1):35. doi:10.1038/s41392-022-01304-4.
- 110. Andrews MC, Vasanthakumar A. Gut microbiota a double-edged sword in cancer immunotherapy. Trends In Cancer. 2023;9(1):3–5. doi:10.1016/j.trecan. 2022.08.003.
- 111. Rousso-Noori L, Mastandrea I, Talmor S, Waks T, Globerson Levin A, Haugas M, Teesalu T, Alvarez-Vallina L, Eshhar Z, Friedmann-Morvinski D, et al. P32-specific CAR T cells with dual antitumor and antiangiogenic therapeutic potential in gliomas. Nat Commun. 2021;12(1):3615. doi:10.1038/s41467-021-23817-2.
- 112. Smith M, Dai A, Ghilardi G, Amelsberg KV, Devlin SM, Pajarillo R, Slingerland JB, Beghi S, Herrera PS, Giardina P, et al. Gut microbiome correlates of response and toxicity following anti-CD19 CAR T cell therapy. Nat Med. 2022;28(4):713–723. doi:10.1038/ s41591-022-01702-9.
- 113. Vidal-Robau N, Caballero G, Archilla I, Ladino A, Fernández S, Ortiz-Maldonado V, Rovira M, Gómez-Hernando M, Delgado J, Suárez-Lledó M, et al. Postmortem neuropathologic examination of a 6-case series of CAR T-cell treated patients. Free Neuropathol. 2022;3. doi:10.17879/freeneuropathology-2022-4365.
- 114. Perrone L, Sampaolo S, Melone MAB. Bioactive phenolic compounds in the modulation of central and peripheral nervous system cancers: facts and misdeeds. Cancers (Basel). 2020;12(2):454. doi:10. 3390/cancers12020454.
- 115. Kim HJ, Kim TJ, Kim YG, Seong C, Cho J-H, Kim W, Lee K-H, Kim D-Y. Antioxidant and antiproliferative activity of finasteride against glioblastoma cells. Pharmaceutics. 2021;13(9):1410. doi:10.3390/pharma ceutics13091410.
- 116. Lin TL, Lu CC, Lai WF, Wu T-S, Lu J-J, Chen Y-M, Tzeng C-M, Liu H-T, Wei H, Lai H-C, et al. Role of gut

microbiota in identification of novel tcm-derived active metabolites. Protein Cell. 2021;12(5):394–410. doi:10. 1007/s13238-020-00784-w.

- 117. Ye Z, He W, Zhang Z, Qiu Z, Zhao Z, Tang BZ. Aiegens for microorganism-related visualization and therapy. Interdiscip Med. 2023;1(2):e20220011. doi:10.1002/ INMD.20220011.
- 118. Wen M, Wang J, Ou Z, Nie G, Chen Y, Li M, Wu Z, Xiong S, Zhou H, Yang Z, et al. Bacterial extracellular vesicles: a position paper by the microbial vesicles task force of the Chinese society for extracellular vesicles. Interdiscip Med. 2023;1(3):e20230017. doi:10.1002/ INMD.20230017.
- 119. Welsh JA, Goberdhan DCI, O'Driscoll L, Buzas EI, Blenkiron C, Bussolati B, Cai H, Di Vizio D, Driedonks TAP, Erdbrügger U, et al. Minimal information for studies of extracellular vesicles (MISEV2023): from basic to advanced approaches. J Extracell Vesicles. 2024;13(2):e12404. doi:10.1002/jev2.12404.
- 120. Wang Y, Qiao W, Zhao Z, Zhao Z, Li M. Preparation of two-dimensional porphyrin-based MOFs/derivatives and their potential in sensing and biomedical applications. Interdiscip Med. 2023;1(3):e20230010. doi:10.1002/INMD.20230010.
- 121. Wanigasekara J, Cullen PJ, Bourke P, Tiwari B, Curtin JF. Advances in 3D culture systems for therapeutic discovery and development in brain cancer. Drug Discov Today. 2023;28(2):103426. doi:10.1016/j. drudis.2022.103426.
- 122. Zhang C, Jin M, Zhao J, Chen J, Jin W. Organoid models of glioblastoma: advances, applications and challenges. Am J Cancer Res. 2020;10(8):2242–2257.
- 123. Lago C, Gianesello M, Santomaso L, Leva G, Ballabio C, Anderle M, Antonica F, Tiberi L. Medulloblastoma and high-grade glioma organoids for drug screening, lineage tracing, co-culture and in vivo assay. Nat Protoc. 2023;18(7):2143–2180. doi:10.1038/s41596-023-00839-2.
- 124. Porter RJ, Murray GI, McLean MH. Current concepts in tumour-derived organoids. Br J Cancer. 2020;123 (8):1209–1218. doi:10.1038/s41416-020-0993-5.
- 125. Albright MBN, Louca S, Winkler DE, Feeser KL, Haig S-J, Whiteson KL, Emerson JB, Dunbar J. Solutions in microbiome engineering: prioritizing barriers to organism establishment. Isme J. 2022;16 (2):331–338. doi:10.1038/s41396-021-01088-5.
- 126. Liang J, Li T, Zhao J, Wang C, Sun H. Current understanding of the human microbiome in glioma. Front Oncol. 2022;12:781741. doi:10.3389/fonc.2022.781741.
- 127. Li M, Xu H, Qi Y, Pan Z, Li B, Gao Z, Zhao R, Xue H, Li G. Tumor-derived exosomes deliver the tumor suppressor miR-3591-3p to induce M2 macrophage polarization and promote glioma progression. Oncogene. 2022;41(41):4618–4632. doi:10.1038/s41388-022-02457-w.
- 128. Li P, Luo H, Ji B, Nielsen J. Machine learning for data integration in human gut microbiome. Microb Cell Fact. 2022;21(1):241. doi:10.1186/s12934-022-01973-4.

- 129. Wang Y, Zhao Y, Bollas A, Wang Y, Au KF. Nanopore sequencing technology, bioinformatics and applications. Nat Biotechnol. 2021;39(11):1348–1365. doi:10.1038/s41587-021-01108-x.
- Belstrøm D. The salivary microbiota in health and disease. J Oral Microbiol. 2020;12(1):1723975. doi:10. 1080/20002297.2020.1723975.
- 131. Robinson KM, Crabtree J, Mattick JS, Anderson KE, Dunning Hotopp JC. Distinguishing potential bacteria-tumor associations from contamination in a secondary data analysis of public cancer genome sequence data. Microbiome. 2017;5(1):9. doi:10.1186/ s40168-016-0224-8.
- 132. Wang L, Li S, Fan H, Han M, Xie J, Du J, Peng F. Bifidobacterium lactis combined with lactobacillus plantarum inhibit glioma growth in mice through modulating PI3K/AKT pathway and gut microbiota. Front Microbiol. 2022;13:986837. doi:10.3389/fmicb.2022. 986837.
- 133. de Paiva IHR, da Silva RS, Mendonça IP, Duarte-Silva E, Botelho de Souza JR, Peixoto CA. Fructooligosaccharide (FOS) and galactooligosaccharide (GOS) improve neuroinflammation and cognition by up-regulating IRS/PI3K/AKT signaling pathway in diet-induced obese mice. J Neuroimmune Pharmacol. 2023;18(3):427–447. doi:10.1007/s11481-023-10069-8.
- 134. Zhang W, Jiang J, Li X, He Y, Chen F, Li W. Dietary factors and risk of glioma in adults: a systematic review and dose-response meta-analysis of observational studies. Front Nutr. 2022;9:834258. doi:10.3389/fnut. 2022.834258.
- 135. Montella L, Sarno F, Altucci L, Cioffi V, Sigona L, Di Colandrea S, De Simone S, Marinelli A, Facchini BA, De Vita F, et al. A root in synapsis and the other one in the gut microbiome-brain axis: are the two poles of ketogenic diet enough to challenge glioblastoma? Front Nutr. 2021;8:703392. doi:10.3389/fnut.2021. 703392.
- 136. Kim J, Kim Y, La J, Park WH, Kim H-J, Park SH, Ku KB, Kang BH, Lim J, Kwon MS, et al. Supplementation with a high-glucose drink stimulates anti-tumor immune responses to glioblastoma via gut microbiota modulation. Cell Rep. 2023;42(10):113220. doi:10.1016/j.celrep.2023.113220.
- 137. Ailioaie LM, Litscher G. Probiotics, photobiomodulation, and disease management: controversies and challenges. Int J Mol Sci. 2021;22(9):4942. doi:10.3390/ ijms22094942.
- 138. Puig-Saenz C, Pearson JRD, Thomas JE, McArdle SEB. A holistic approach to hard-to-treat cancers: The future of immunotherapy for glioblastoma, triple negative breast cancer, and advanced prostate cancer. Biomedicines. 2023;11(8):2100. doi:10.3390/biomedi cines11082100.
- 139. Kim MS, Kim Y, Choi H, Kim W, Park S, Lee D, Kim DK, Kim HJ, Choi H, Hyun D-W, et al. Transfer of a healthy microbiota reduces amyloid and tau

pathology in an alzheimer's disease animal model. Gut. 2020;69(2):283-294. doi:10.1136/gutjnl-2018-317431.

- 140. Sieow BF, Wun KS, Yong WP, Hwang IY, Chang MW. Tweak to treat: reprograming bacteria for cancer treatment. Trends Cancer. 2021;7(5):447–464. doi:10. 1016/j.trecan.2020.11.004.
- 141. Suryawanshi YR, Schulze AJ. Oncolytic viruses for malignant glioma: on the verge of success? Viruses. 2021;13(7):1294. doi:10.3390/v13071294.
- 142. Manivannan AC, Dhandapani R, Velmurugan P, Thangavelu S, Paramasivam R, Ragunathan L, Saravanan M. Phage in cancer treatment – biology of therapeutic phage and screening of tumor targeting peptide. Expert Opin Drug Deliv. 2022;19(7):873–882. doi:10.1080/17425247.2022.2094363.
- 143. Hameedat F, Mendes BB, Conniot J, Di Filippo LD, Chorilli M, Schroeder A, Conde J, Sousa F. Engineering nanomaterials for glioblastoma nanovaccination. Nat Rev Mater. 2024; doi:10.1038/ s41578-024-00684-z.
- 144. Sun B, Sawant H, Borthakur A, Bihl JC. Emerging therapeutic role of gut microbial extracellular vesicles in neurological disorders. Front Neurosci. 2023;17:1241418. doi:10.3389/fnins.2023.1241418.
- 145. Chen L, Shen J, Kang Z, Zhang Z, Zhang Z, Zhang L, Xiao Z, Zhang Q, Fang H, Zhou J, et al. Fusobacterium nucleatum-mimicking nanovehicles to overcome chemoresistance for breast cancer treatment by eliminating tumor-colonizing bacteria. Chem. 2024;10 (6):1783–1803. doi:10.1016/j.chempr.2024.01.030.
- 146. Li Z, Meng X, Wu P, Zha C, Han B, Li L, Sun N, Qi T, Qin J, Zhang Y, et al. Glioblastoma cell-derived lncRNA-containing exosomes induce microglia to produce complement C5, promoting chemotherapy resistance. Cancer Immunol Res. 2021;9(12):1383–1399. doi:10.1158/2326-6066.Cir-21-0258.
- 147. Nguyen VD, Nguyen TT, Pham TT, Packianather M, Le CH. Molecular screening and genetic diversity analysis of anticancer azurin-encoding and azurin-like genes in human gut microbiome deduced through cultivation-dependent and cultivation-independent studies. Int Microbiol. 2019;22(4):437–449. doi:10. 1007/s10123-019-00070-8.
- 148. Nowakowski P, Markiewicz-Żukowska R, Gromkowska-Kępka K, Naliwajko SK, Moskwa J, Bielecka J, Grabia M, Borawska M, Socha K. Mushrooms as potential therapeutic agents in the treatment of cancer: evaluation of anti-glioma effects of coprinus comatus, cantharellus cibarius, Lycoperdon perlatum and lactarius deliciosus extracts. Biomed Pharmacother. 2021;133:111090. doi:10.1016/j.biopha. 2020.111090.
- 149. Klimenko NS, Odintsova VE, Revel-Muroz A, Tyakht AV. The hallmarks of dietary intervention-resilient gut microbiome. Npj Biofilms

Microbiomes. 2022;8(1):77. doi:10.1038/s41522-022-00342-8.

- 150. Pahwa B, Leskinen S, Didia E, Huda S, D'Amico RS. Role of nutritional adjuncts in the management of gliomas: a systematic review of literature. Clin Neurol Neurosurg. 2023;231:107853. doi:10.1016/j.clineuro. 2023.107853.
- 151. Giles EM, D'Adamo GL, Forster SC. The future of faecal transplants. Nat Rev Microbiol. 2019;17 (12):719–719. doi:10.1038/s41579-019-0271-9.
- 152. Ağagündüz D, Çelik E, Cemali Ö, Bingöl FG, Özenir Ç, Özoğul F, Capasso R. Probiotics, live biotherapeutic products (LBPs), and gut-brain axis related psychological conditions: implications for research and dietetics. Probiotics Antimicrob Proteins. 2023;15(4):1014–1031. doi:10.1007/s12602-023-10092-4.
- 153. Wang Y, Sheng J, Chai J, Zhu C, Li X, Yang W, Cui R, Ge T. Filamentous bacteriophage—A powerful carrier for glioma therapy. Front Immunol. 2021;12:729336. doi:10.3389/fimmu.2021.729336.
- 154. Yin D, Zhong Y, Ling S, Lu S, Wang X, Jiang Z, Wang J, Dai Y, Tian X, Huang Q, et al. Dendritic-cell-targeting virus-like particles as potent mRNA vaccine carriers. Nat Biomed Eng. 2024; doi:10.1038/s41551-024-01208-4.

- 155. Cheng K, Zhao R, Li Y, Qi Y, Wang Y, Zhang Y, Qin H, Qin Y, Chen L, Li C, et al. Bioengineered bacteria-derived outer membrane vesicles as a versatile antigen display platform for tumor vaccination via plug-and-display technology. Nat Commun. 2021;12(1):2041. doi:10.1038/s41467-021-22308-8.
- 156. Ren QL, Wang Q, Zhang XQ, Wang M, Hu H, Tang J-J, Yang X-T, Ran Y-H, Liu H-H, Song Z-X, et al. Anticancer activity of diosgenin and its molecular mechanism. Chin J Integr Med. 2023;29(8):738–749. doi:10.1007/s11655-023-3693-1.
- 157. Weiss S, Amir A, Hyde ER, Metcalf JL, Song SJ, Knight R. Tracking down the sources of experimental contamination in microbiome studies. Genome Biol. 2014;15(12):564. doi:10.1186/s13059-014-0564-2.
- 158. Zheng D, Liwinski T, Elinav E. Interaction between microbiota and immunity in health and disease. Cell Res. 2020;30(6):492–506. doi:10.1038/s41422-020-0332-7.
- 159. Aggarwal P, Luo W, Pehlivan KC, Hoang H, Rajappa P, Cripe TP, Cassady KA, Lee DA, Cairo MS. Pediatric versus adult high grade glioma: immunotherapeutic and genomic considerations. Front Immunol. 2022;13:1038096. doi:10.3389/fimmu.2022.1038096.